



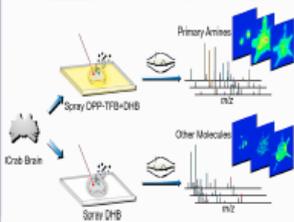
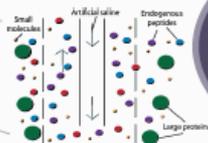
AP MALDI-Quadrupole-Orbitrap MS Platform for High Spatial and High Mass Spectral Resolution In Situ Analysis of Biomolecules

Lingjun Li, Bingming Chen, Gongyu Li, Caitlin Keller, Yatao Shi, Qinjingwen Cao, Chuanzi Ouyang, Jill Johnson
Vilas Distinguished Achievement Professor
Charles Melbourne Johnson Distinguished Chair
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**MassTech Inc. Breakfast Seminar, ASMS 2019
Atlanta, GA, USA, June 3, 2019**

Li Research Group

Crustacean Neuropeptidomic Studies



Neuropeptide changes induced by food intake, environmental stress, etc. in crustacean models are profiled and compared in this project. Microdialysis is performed to monitor neuropeptide secretion in live animals.

Research Focus

Development of Mass Spectrometry-Based Tools for Neuroscience and Bioanalytical Research



Group Accomplishments

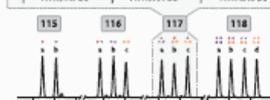
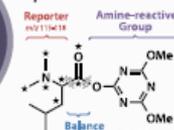


Graduate students in the lab have received more than 130 awards, including multiple training grants (RFP, CRTI, CVAC, CNTP, METP, NSF and NIH fellowships). Our lab members have given more than 300 research presentations at national and international meetings. We have published more than 100 peer-reviewed research journal articles and successfully graduated 40 Ph.Ds.

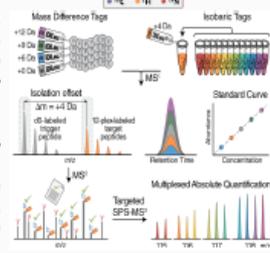
Our research has been supported by NIH, NSF, DOD, Sloan Research Foundation, American Society for Mass Spectrometry, UW-Madison School of Pharmacy, UW Carbone Cancer Center, UW Institute for Clinical and Translational Research, Wisconsin Alzheimer's Disease Research Center and Wisconsin Alumni Research Foundation.

Quantitative MS Strategies

12-plex DiLeu Isobaric Tag

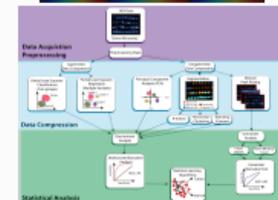
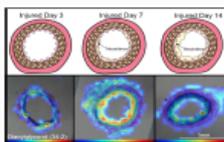


We created DiLeu isobaric and isotopic tags as a cost-effective alternative to widely-used, but expensive, commercial reagents. Paired with high-resolution MS, up to 12 biological samples can be simultaneously identified and quantified in a single experiment.



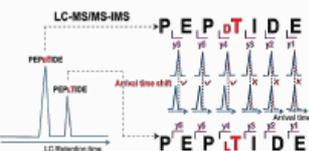
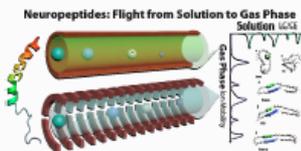
Mass Spectrometry Imaging

Mass spectrometry imaging enables scientists to obtain 2D and 3D maps of biological analytes in their native tissues without the need for antibodies. The Li group implements this technology to examine distribution patterns of neuropeptides, proteins, lipids, and metabolites in a variety of tissues.



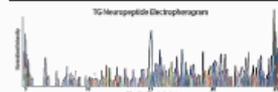
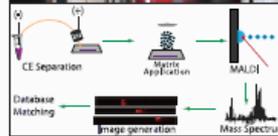
Ion Mobility-MS

IM-MS separates gas-phase ions by mass, charge, size, and shape through collisions with inert gas molecules. The Li group is using it to develop novel methods for glycan separation, peptide identification, and structural analyses.



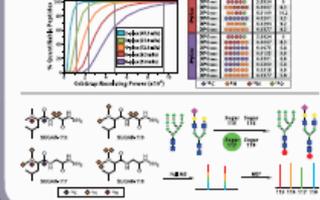
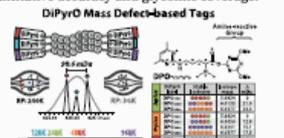
Capillary Electrophoresis MS

Capillary zone electrophoresis and capillary isotachic focusing are being developed to couple with MS detection and MS imaging, both off-line and on-line, for enhanced neuropeptidomic and glycomic analysis.



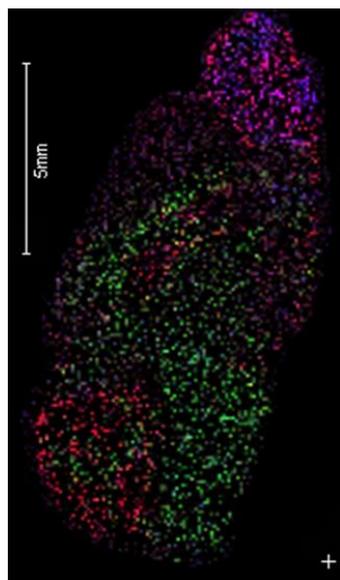
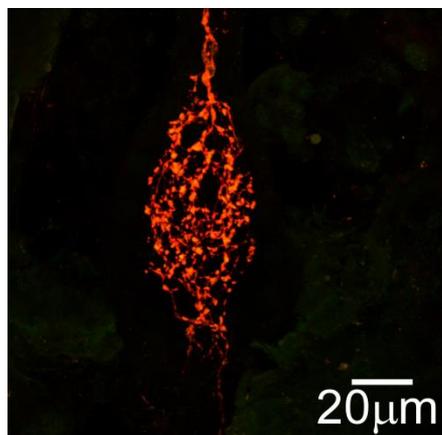
Glycan Analysis

In conjunction with CE, LC, or IM-MS separation, our group investigates glycan structure and quantitation using commercial (e.g. aminoxyTMT) and novel labeling strategies. MultiNotch MS² has been applied to improve quantitative accuracy and glycomic coverage.



Instrumentation





MS imaging advantages

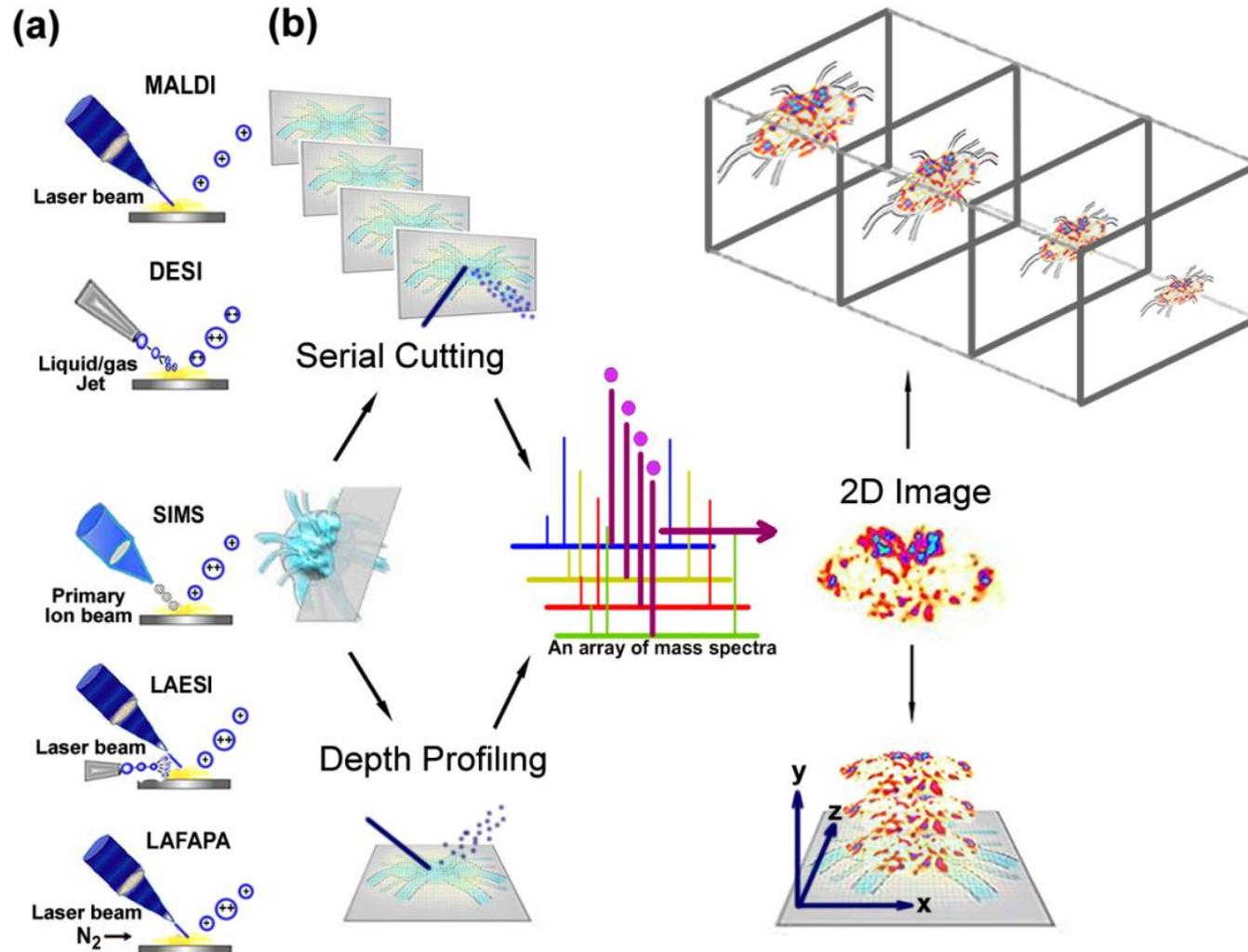
- No labeling required
 - ✓ Biomolecules are functionally unmodified
- Image biomolecular modifications
 - ✓ PTM's
 - ✓ Metabolites
- Detailed information on molecular identity
- Large scope of different elements and molecules, discovery of unknowns
- Extend histopathology to a molecular level
- Multiplexing and highly parallel

Imaging Mass Spectrometry

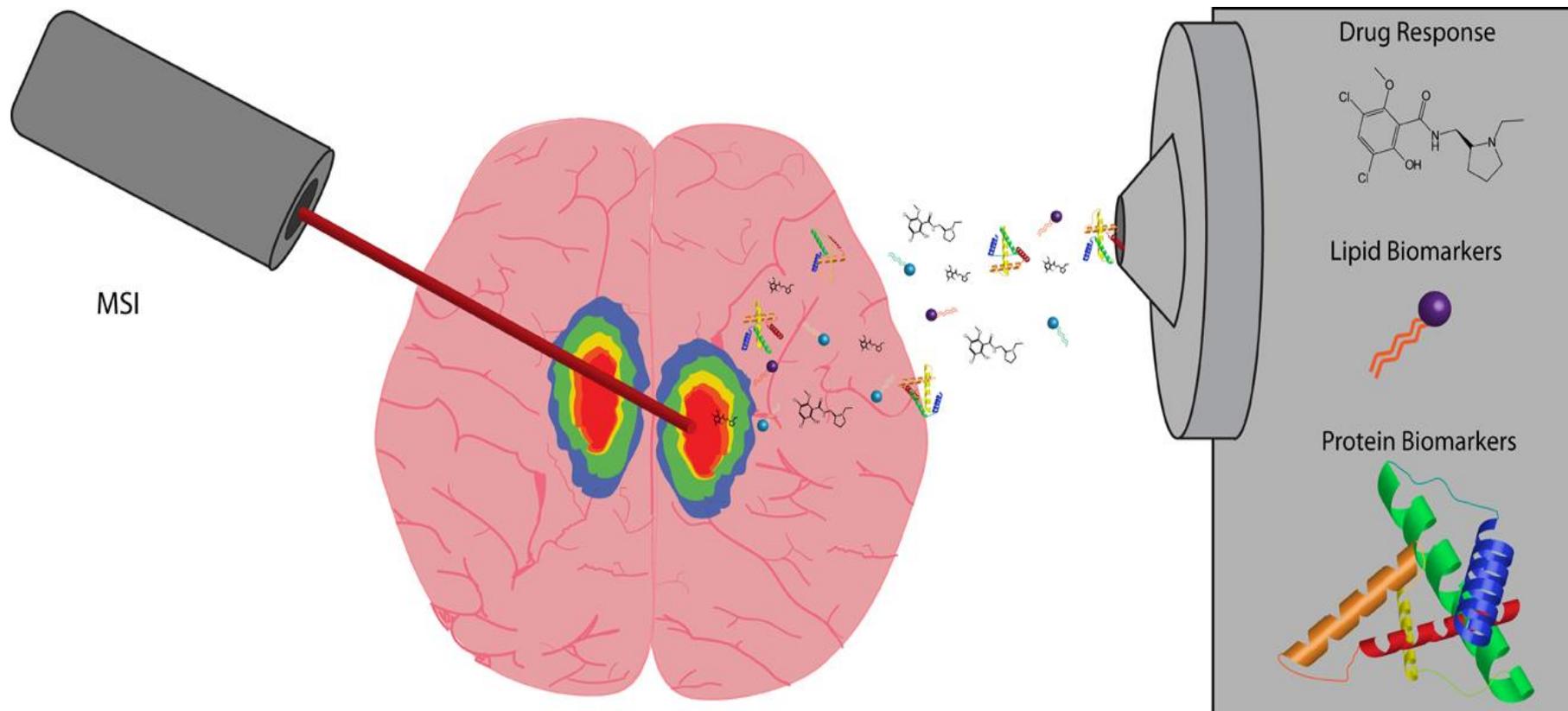
Liam A. McDonnell, Ron M. A. Heeren

Mass Spectrometry Reviews (2007) **26** 606-643

Mass spectral imaging: Ionization methods and tissue preparation strategies

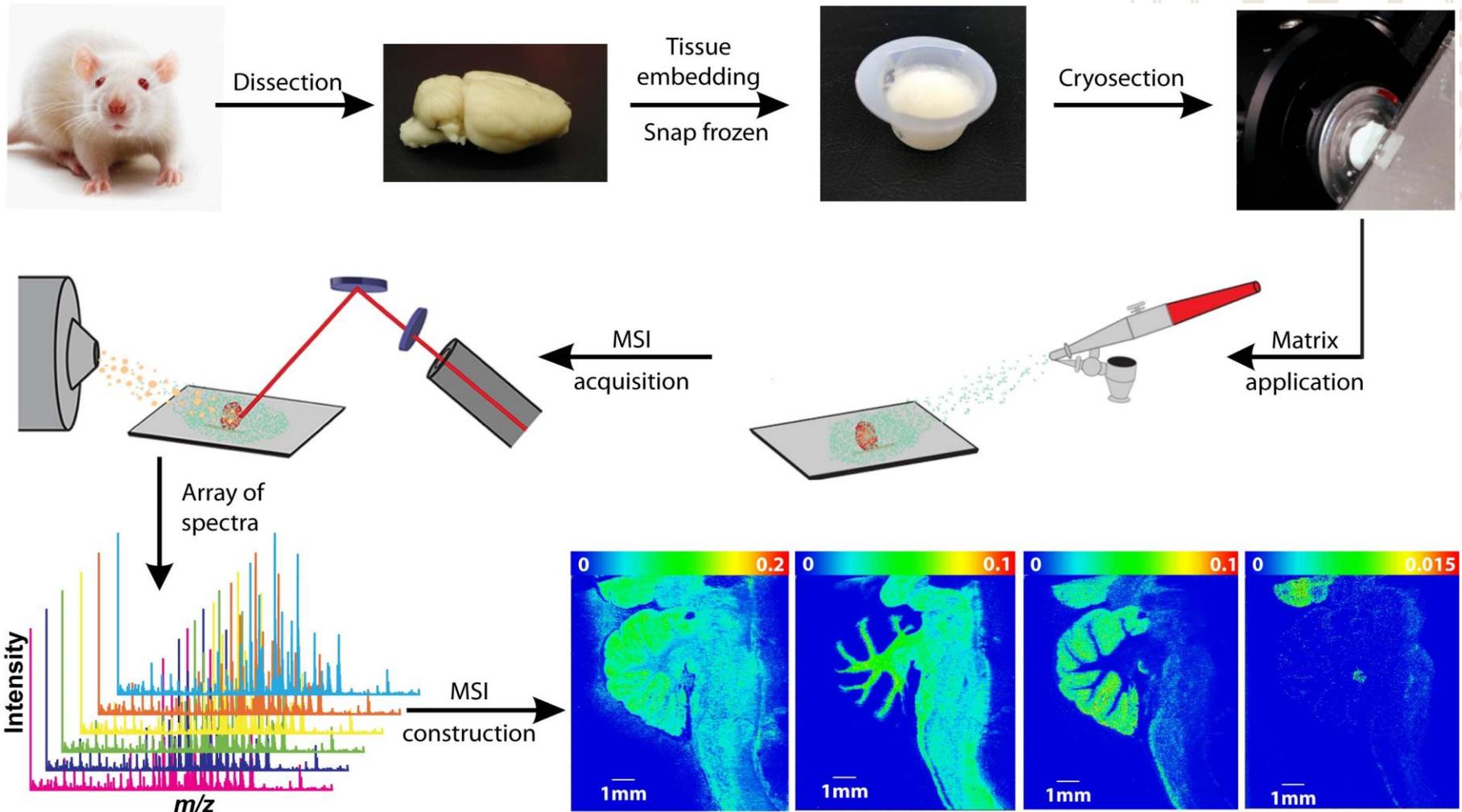


With MALDI MSI, we can...



H. Ye, E. Gemperline, and L. Li (2013). A vision for better health: Mass spectrometry imaging for clinical diagnostics. *Clinica Chimica Acta*. 420, 11-22.

Typical MS Imaging Workflow



Chen[#], Gemperline[#] & Li (2014), *Bioanalysis*, 6(4), 525-540; ([#]Co-First authors)
Angel & Caprioli (2013), *Biochemistry*, 52, 3818-3828; Gessel, Norris & Caprioli (2014), *J. Proteom.*, 107, 71-82.
Part of artwork by Dr. Erin Gemperline.

Image Acquisition Animation

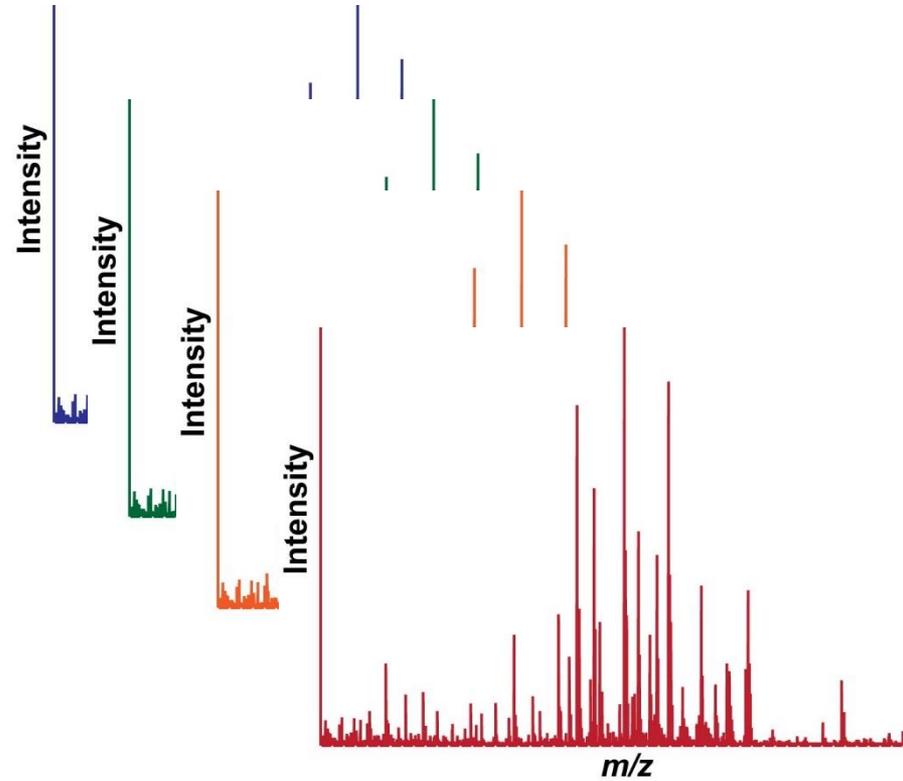
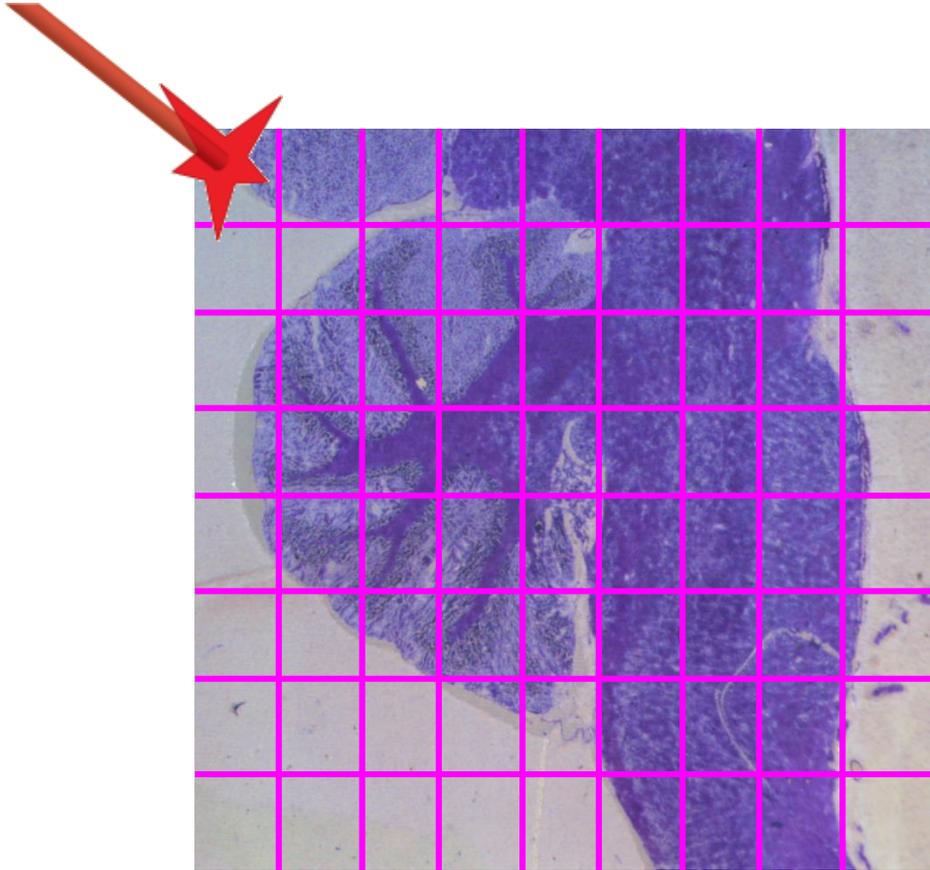
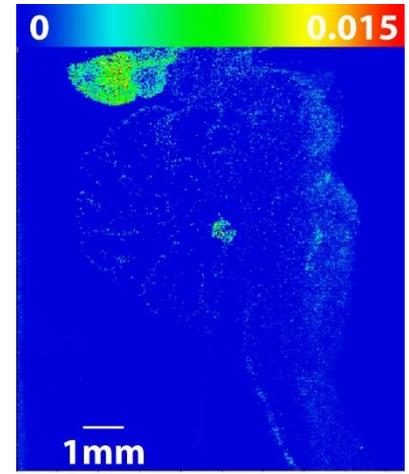
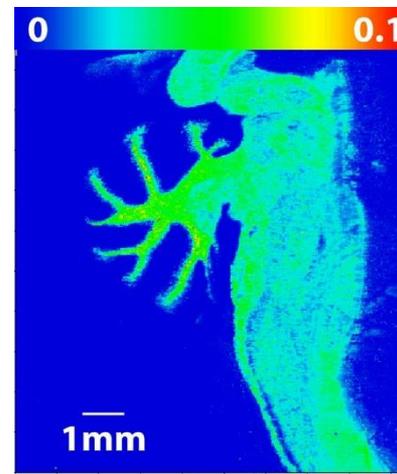
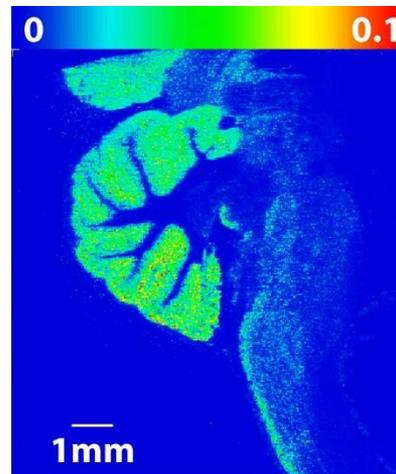
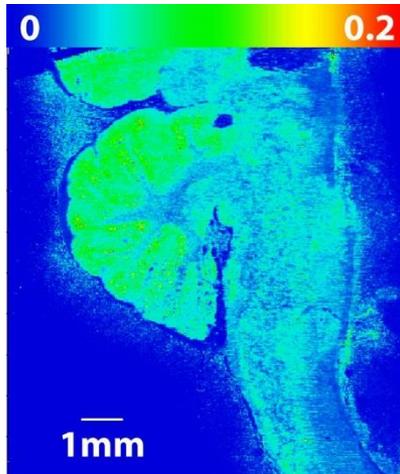
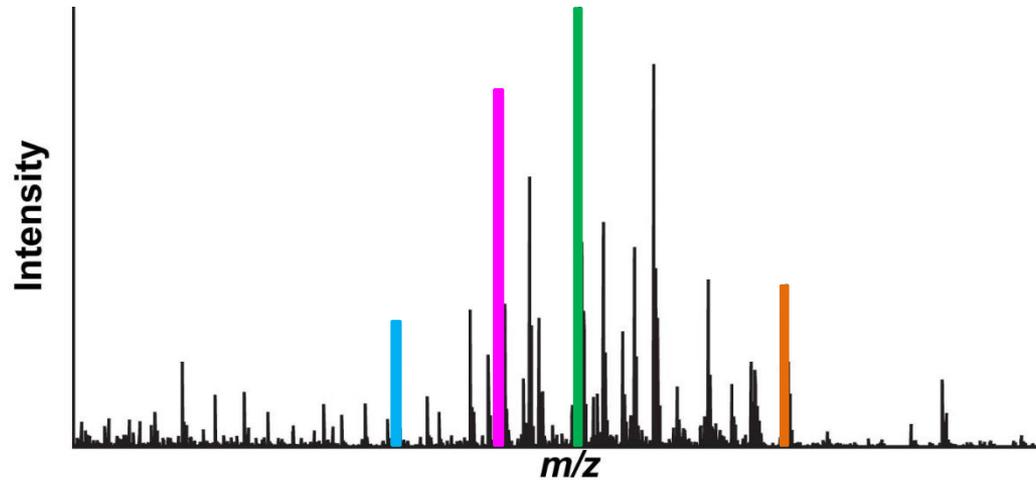
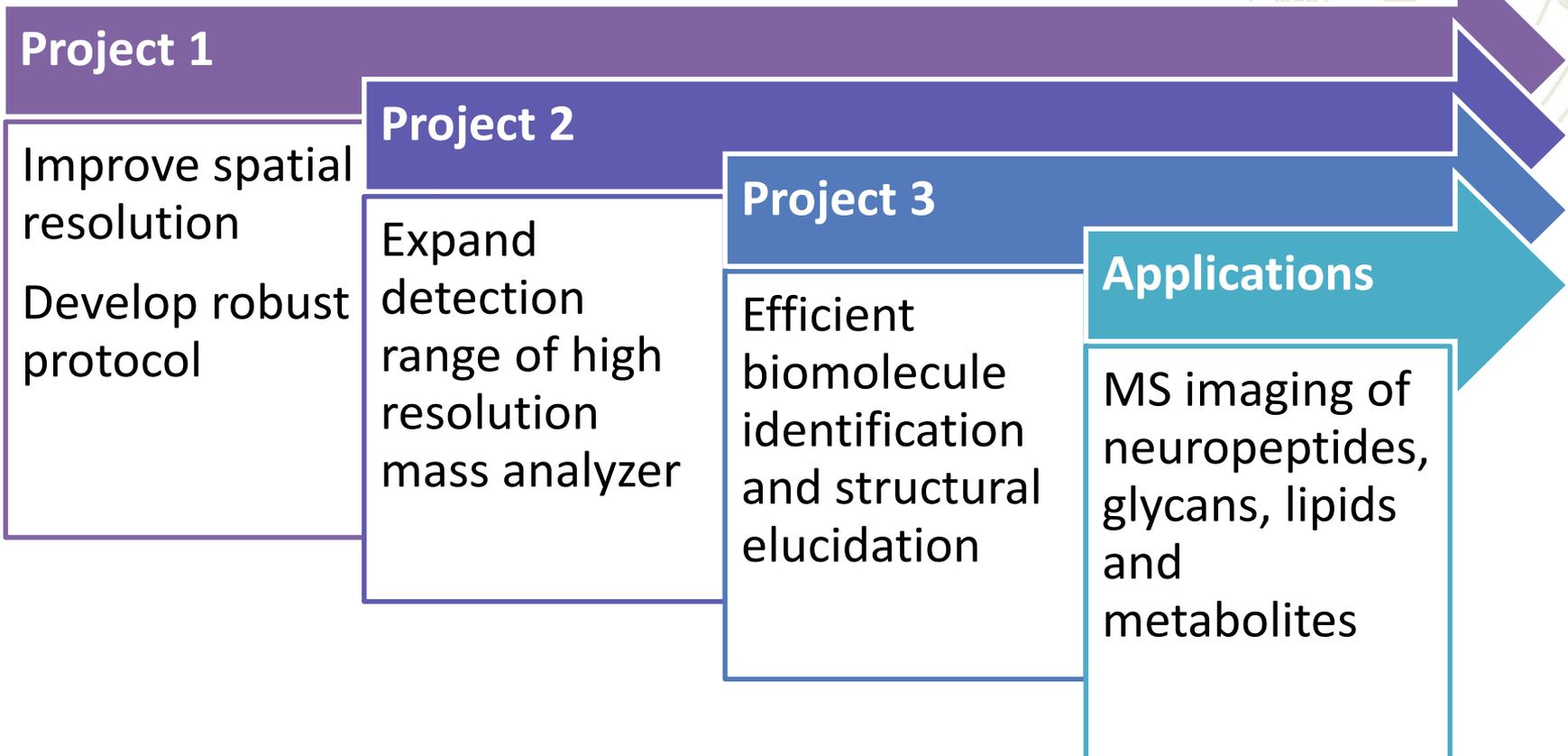


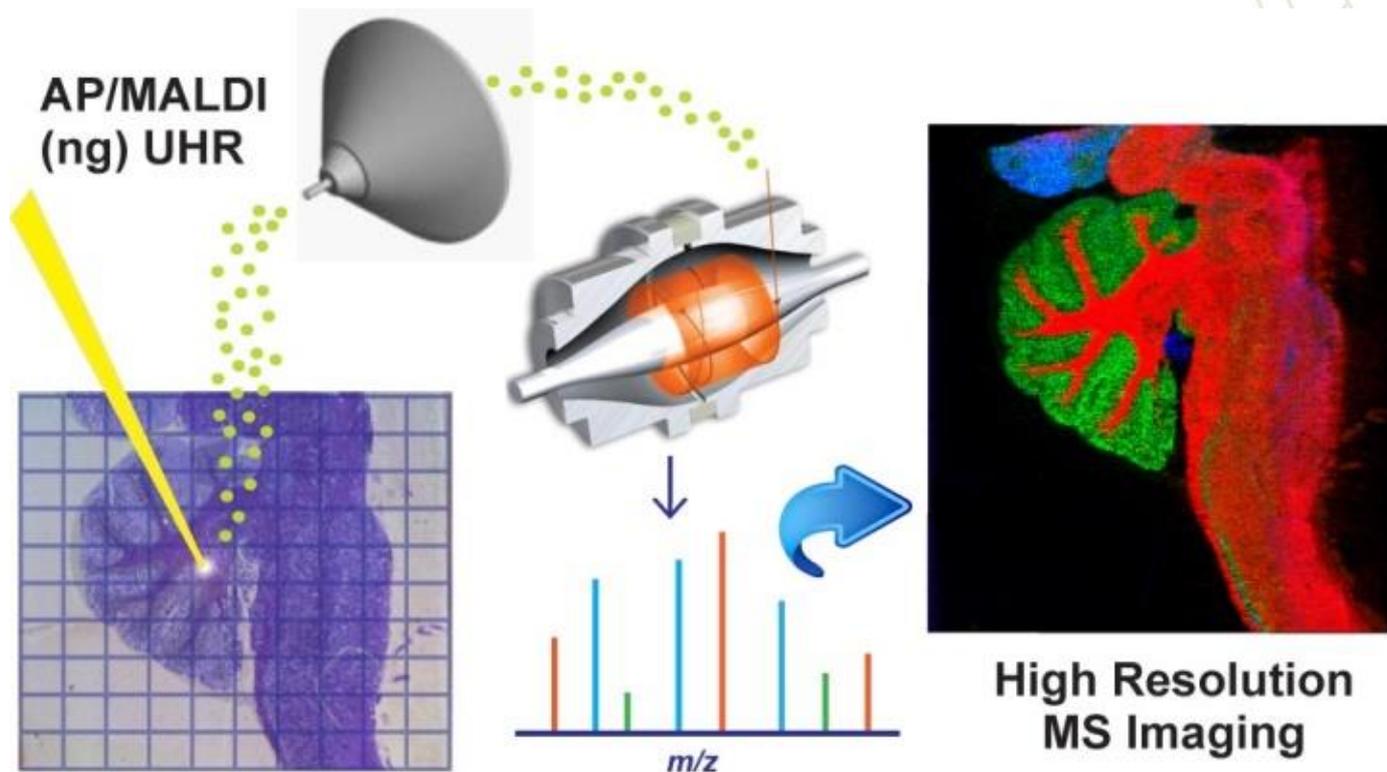
Image Data Analysis Animation



Ongoing Interest in MS Imaging

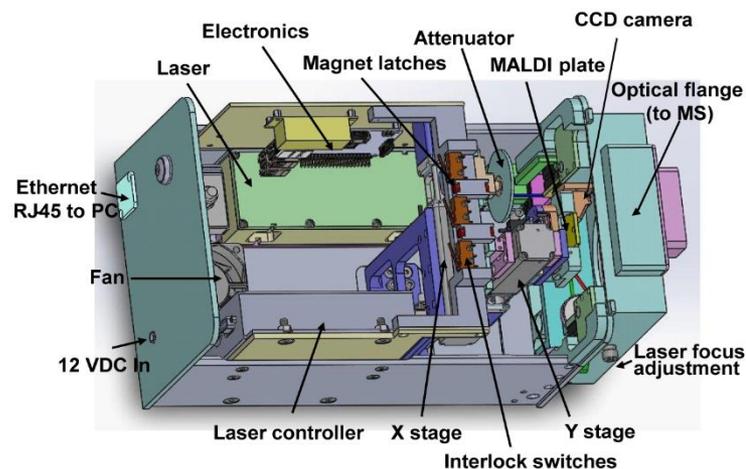


Project 1. A High Resolution Atmospheric Pressure MALDI-Quadrupole-Orbitrap Platform Enables *In Situ* Analysis of Biomolecules by Multi-Mode Ionization and Acquisition

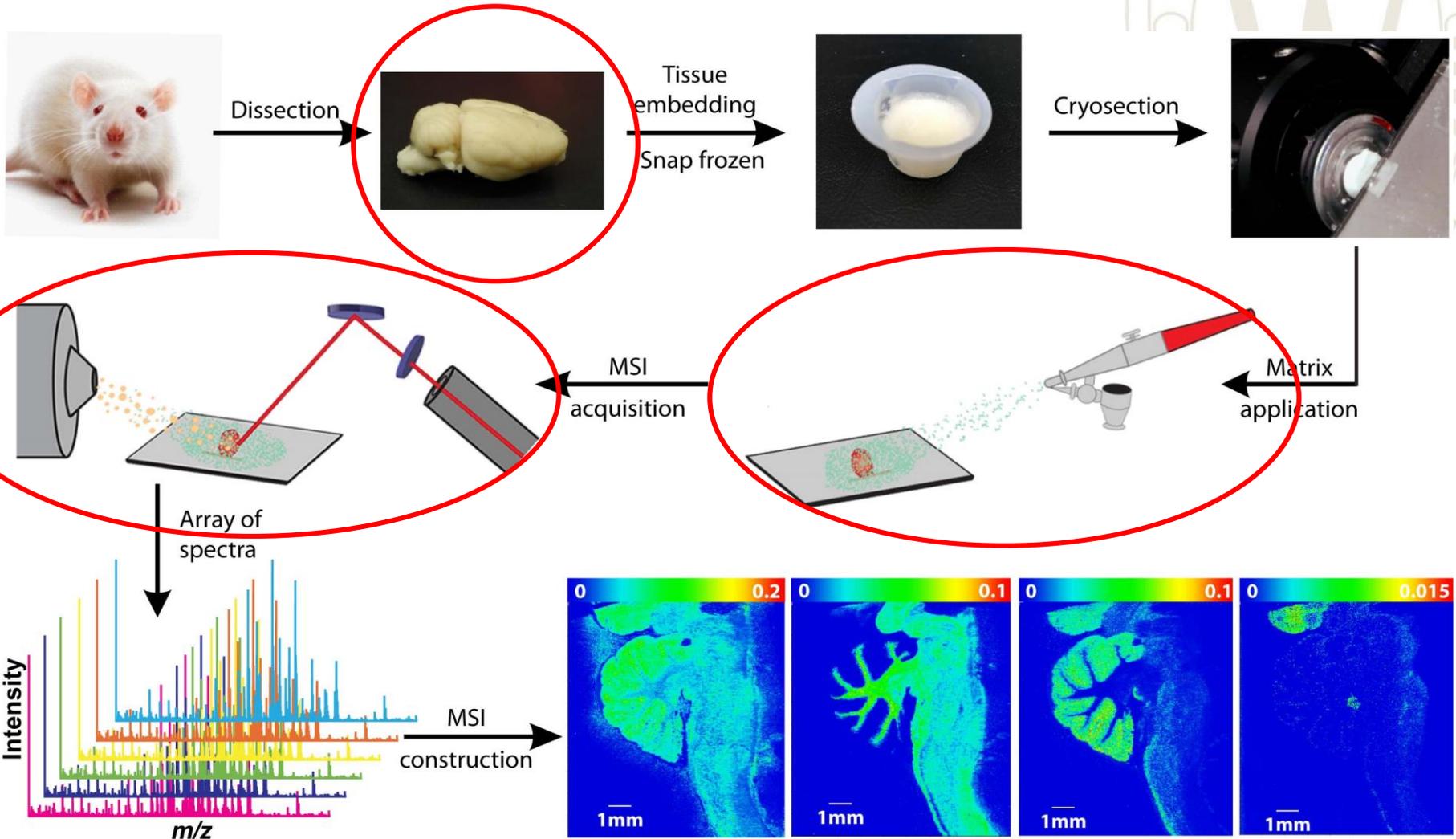


AP/MALDI-Q-Orbitrap Capability

- **AP/MALDI: an alternative to vacuum MALDI**
 - Interchangeable with ESI source
 - Ease of sample introduction and handling at AP condition
 - Capable of analyzing volatile molecules
 - Optimized source geometry design and ion transfer efficiency
- **Multiple types of ionization**
 - MALDI, novel ionizations for multiply charged ions (LSI and MAI/MAIV)
- **High resolution MS imaging**
 - In mass: 240k mass resolution at m/z 200
 - In space: $< 10 \mu\text{m}$ spatial resolution



Optimization for MS Imaging

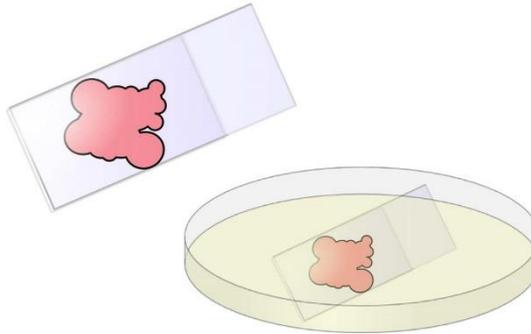


Chen[#], Gemperline[#] & Li (2014), *Bioanalysis*, 6(4), 525-540; ([#]Co-First authors)
Angel & Caprioli (2013), *Biochemistry*, 52, 3818-3828; Gessel, Norris & Caprioli (2014), *J. Proteom.*, 107, 71-82.
Part of artwork by Dr. Erin Gemperline.

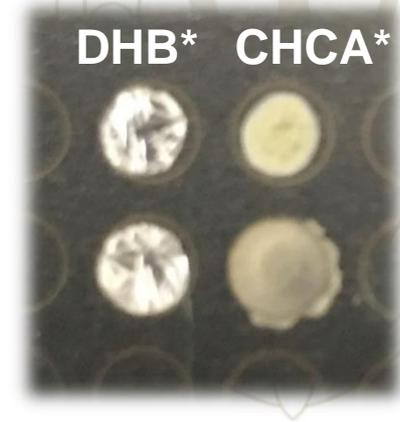
Optimization for MS Imaging



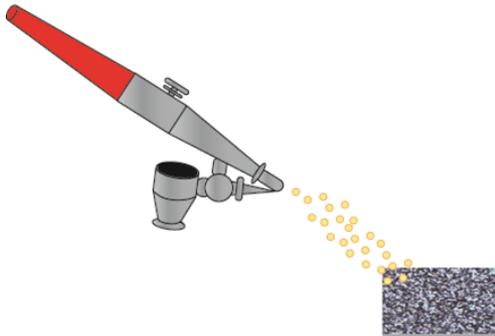
Protease Inhibition



Tissue Rinse



Matrix



Matrix Application Method

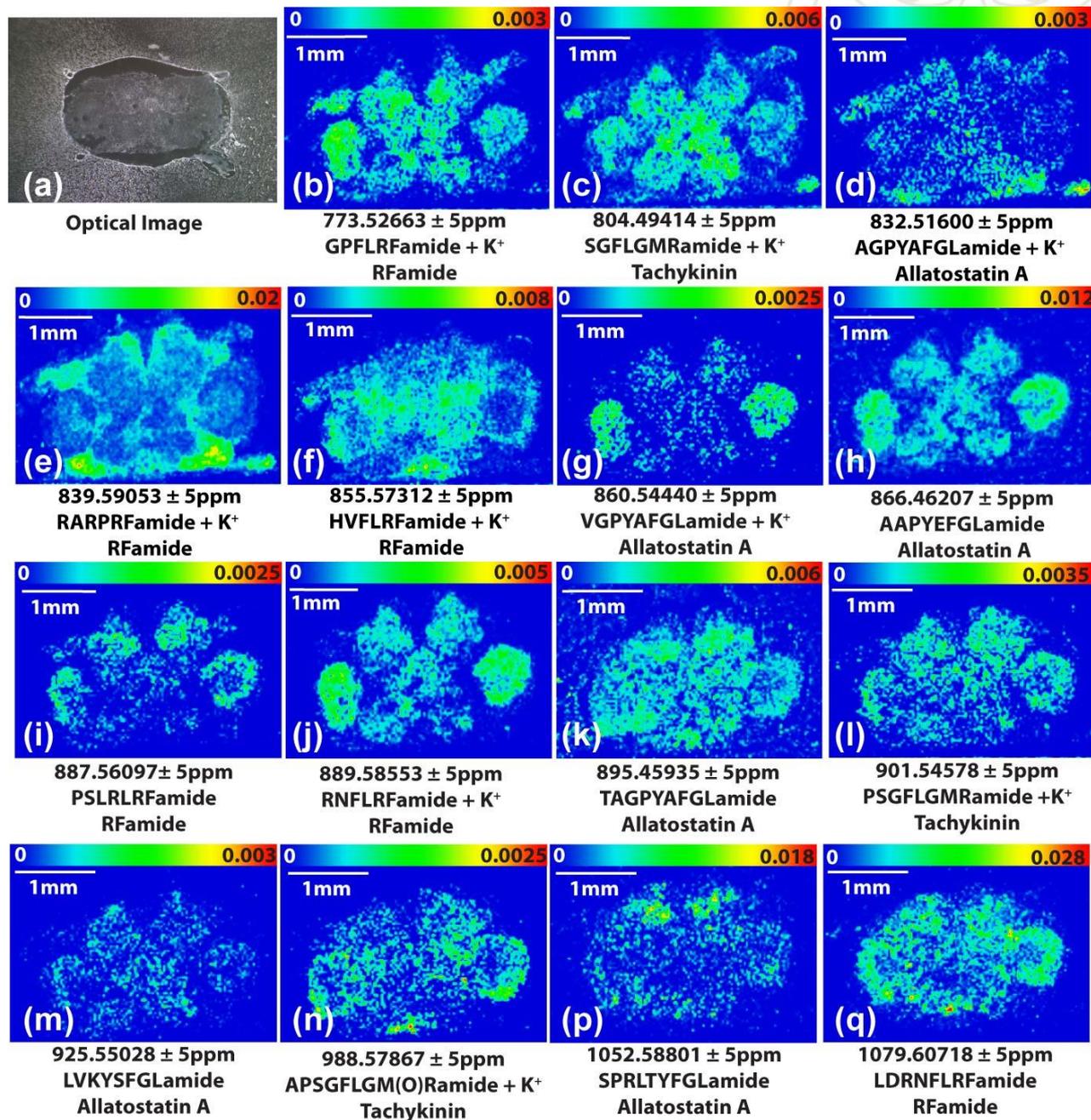


Instrument



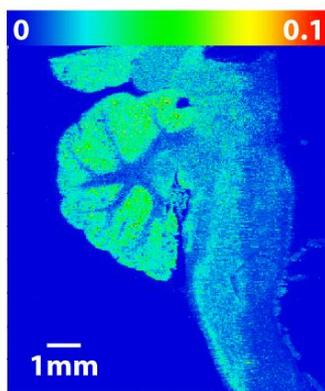
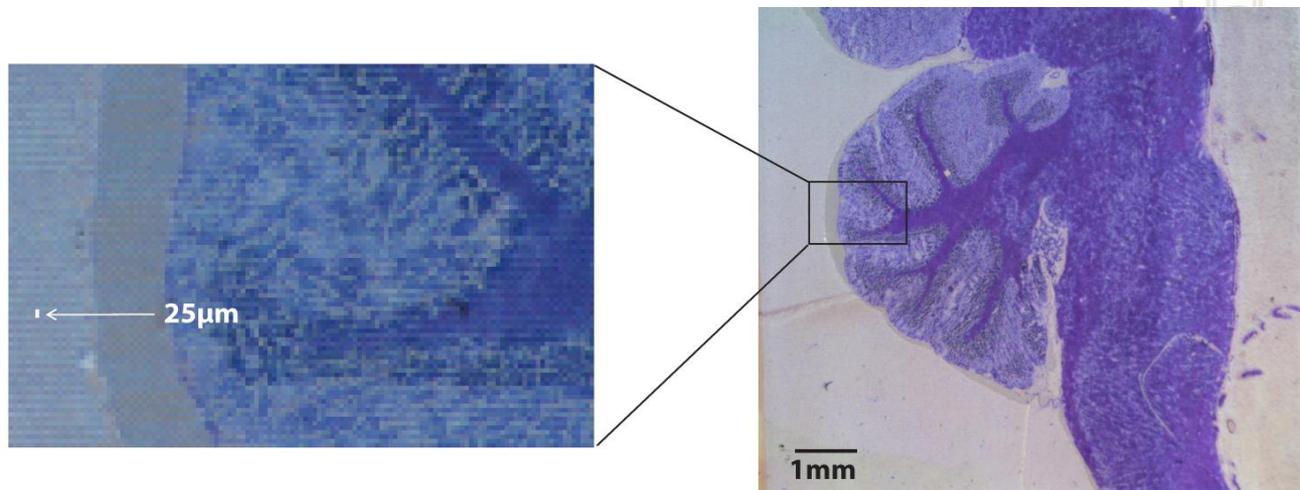
*DHB: 2,5-Dihydroxybenzoic acid
*CHCA: α -Cyano-4-hydroxycinnamic acid

High Resolution MSI of Neuropeptides in Crustacean Brain Tissue Section by AP MALDI Q-Orbitrap Platform

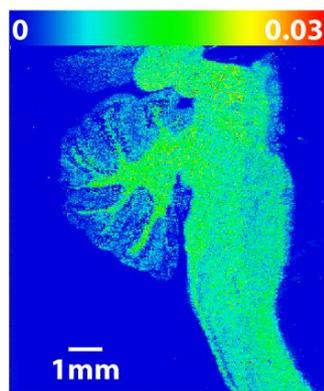


Chen et al., *Analytica Chimica Acta*, 1007, 16-25 (2018).

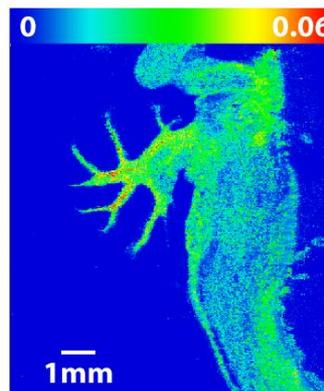
High Resolution MS Imaging



$739.46657 \pm 5\text{ppm}$
PA (36:2) + K⁺



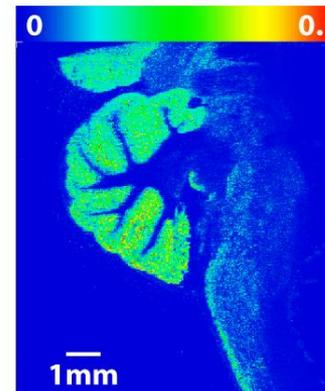
$767.49623 \pm 5\text{ppm}$
PA (38:2) + K⁺



$850.65692 \pm 5\text{ppm}$
GlcCer(42:1) + K⁺



$866.64593 \pm 5\text{ppm}$
PC (P-40:1) + K⁺



$713.45162 \pm 5\text{ppm}$
PA (34:1) + K⁺

High Resolution Trade-off: Limited Mass Range



MALDI-LTQ-Orbitrap
m/z 4000



QE HF
m/z 6000



solarix XR 7T FTICR
m/z 10000

*metabolite
peptide
glycan*

protein

mAb

Molecular Weight

1k

5k

10k

50k

100k



oxytocin 1.0K

bradykinin 1.1K

neuropeptide Y 4.3K

insulin 5.7K

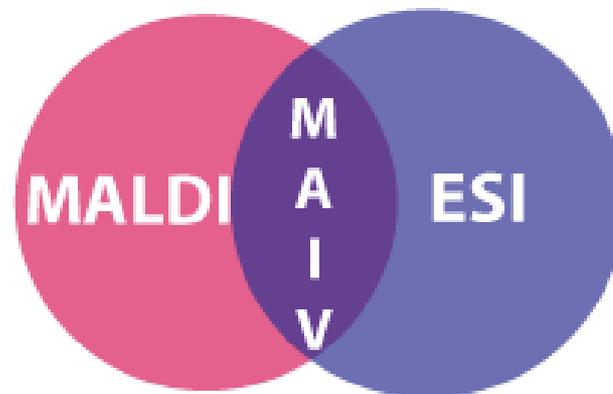
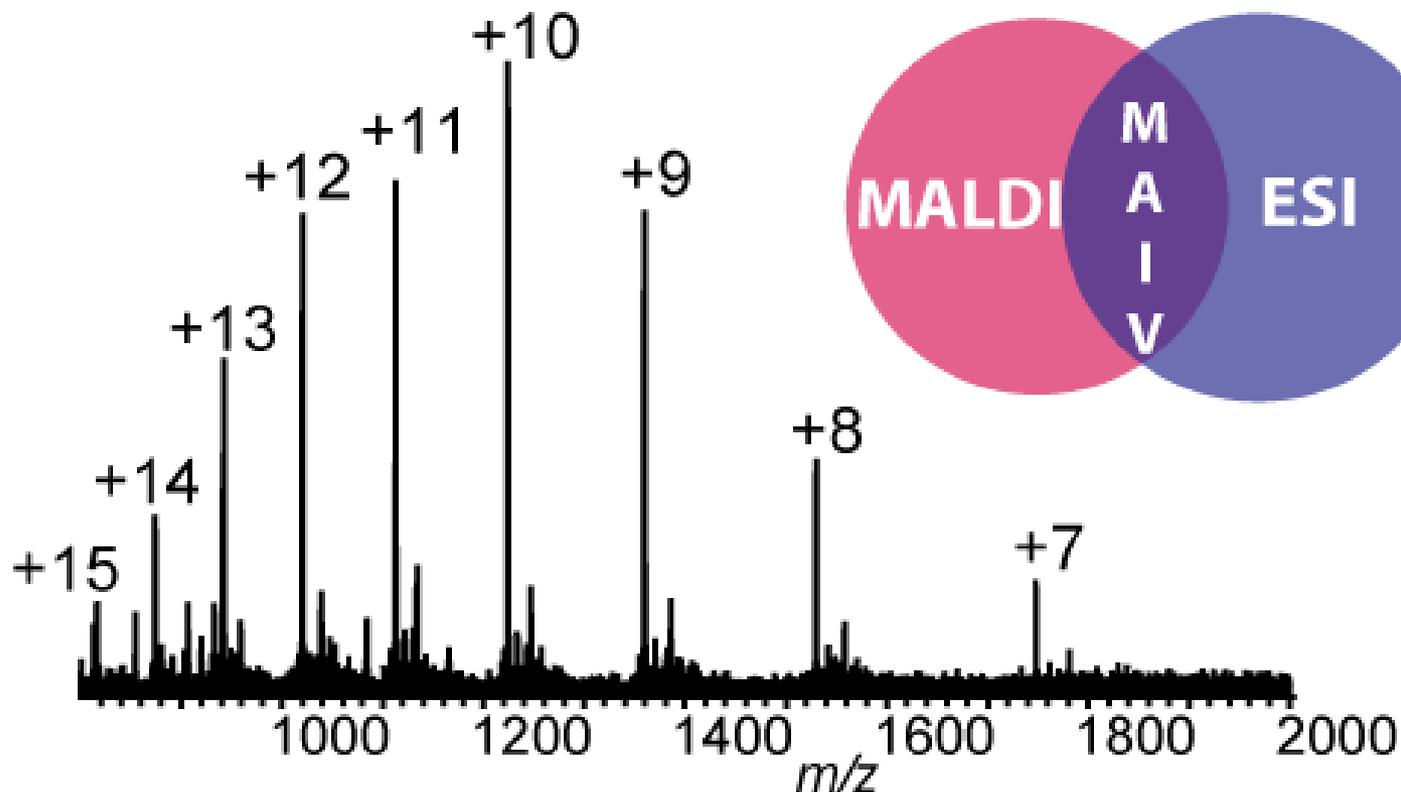
cytochrome C 12.3K

lysozyme 14.3K

myoglobin 17.0K

mAb 150K

Project 2. “Magic” Ionization on High Performance Mass Spectrometers

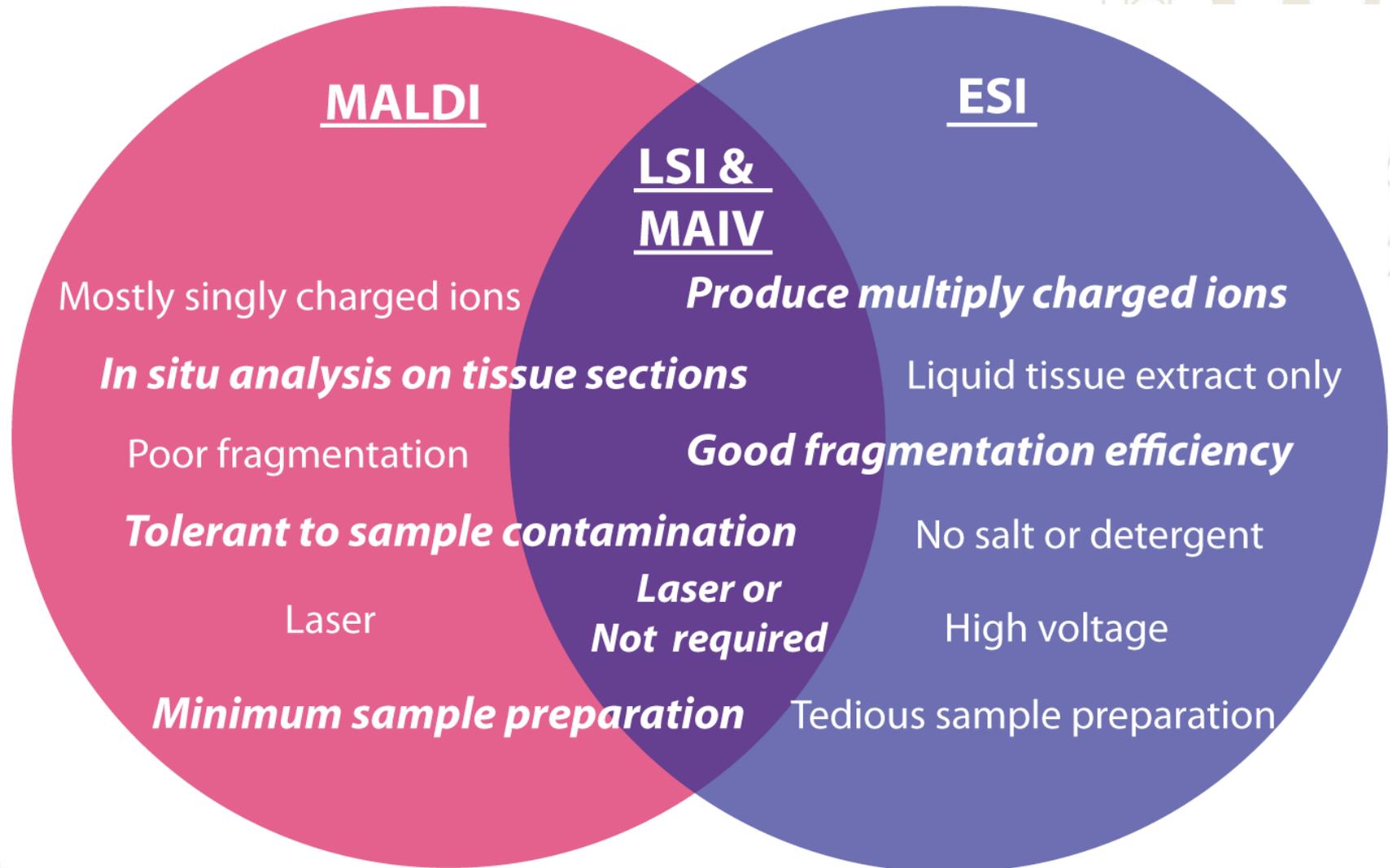


Dr. Bingming Chen



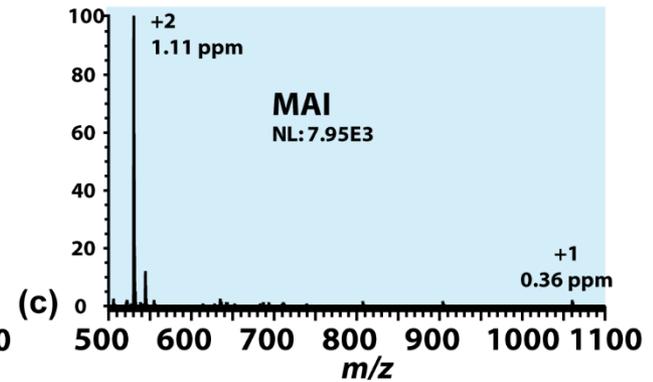
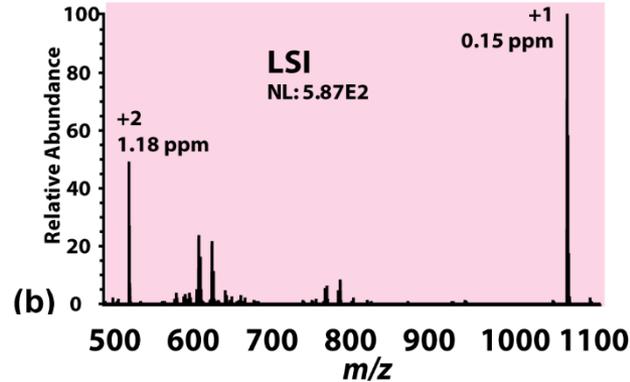
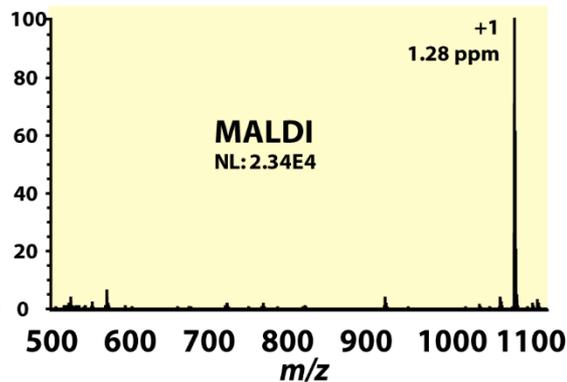
B.Chen, C.B. Lietz, C.Ouyang, X. Zhong, M. Xu and L. Li (2016). *Analyt. Chim. Acta.*, vol 916. p.52-59; B.Chen, C.B. Lietz, L. Li (2014). *J. Am. Soc. Mass Spectrom.*, vol. 25 (12). p. 2177-80; B. Chen[#], C. Ouyang[#], Z. Tian, M. Xu and L. Li, *To be submitted to Analyt. Chim. Acta.* ([#]Co-First authors);

“Magic” Ionization – Multiply Charged MALDI

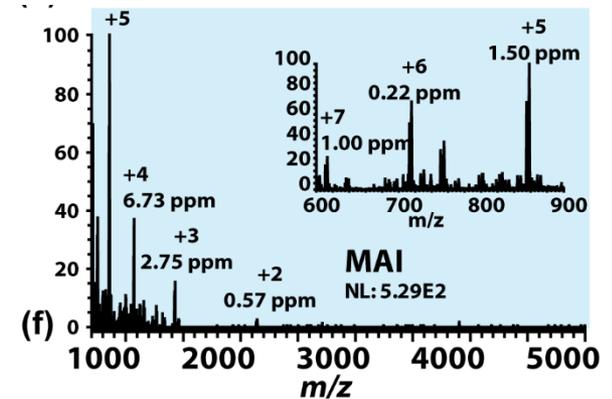
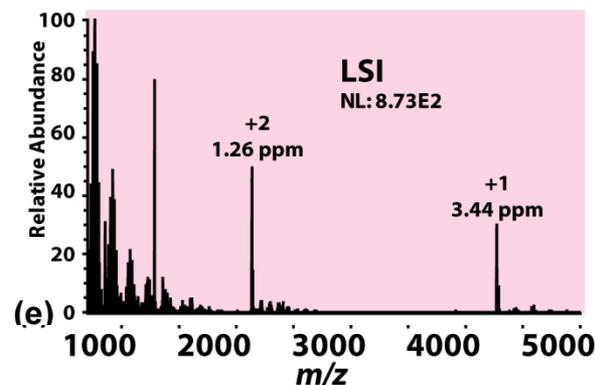
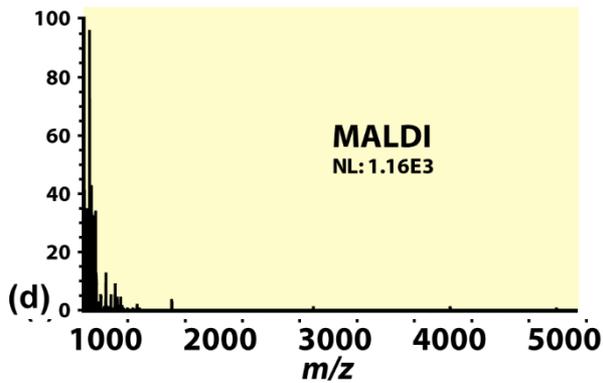


Multiply Charged Ions on AP/MALDI MS

Bradykinin 1059.5608 Da

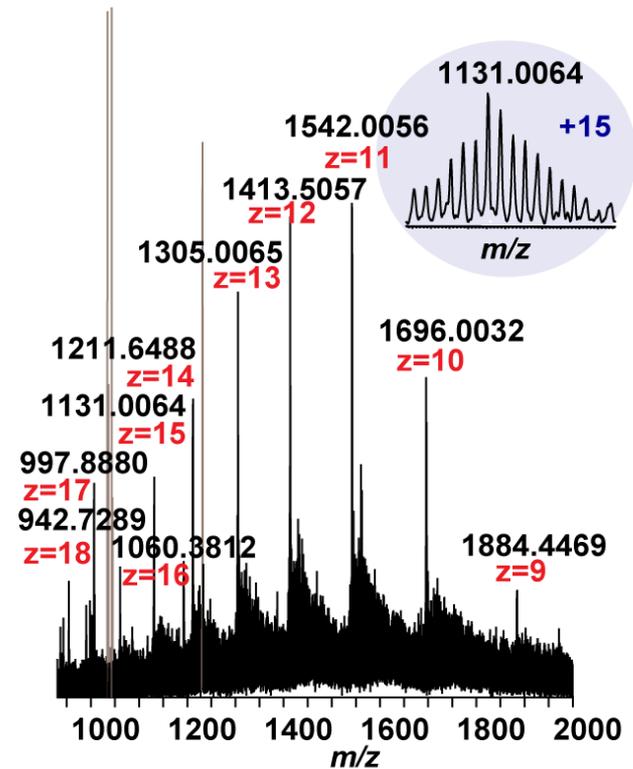


Neuropeptide Y 4269.0808 Da

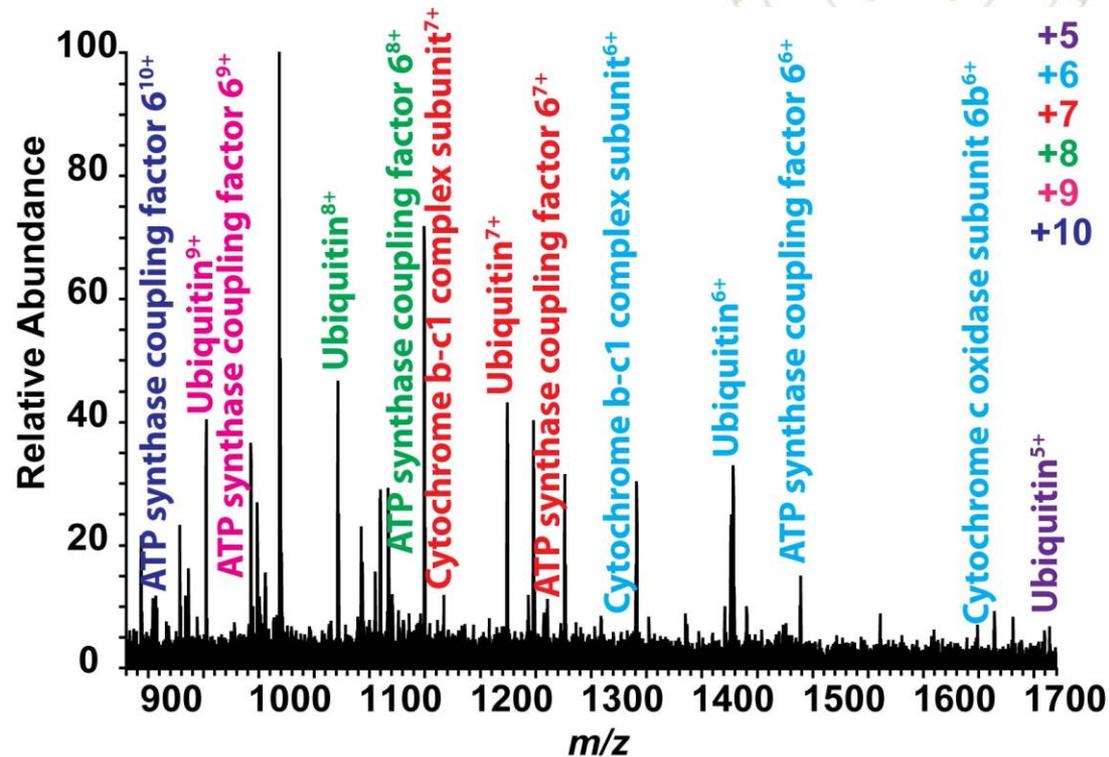


Extended Mass Range Enables Protein Detection

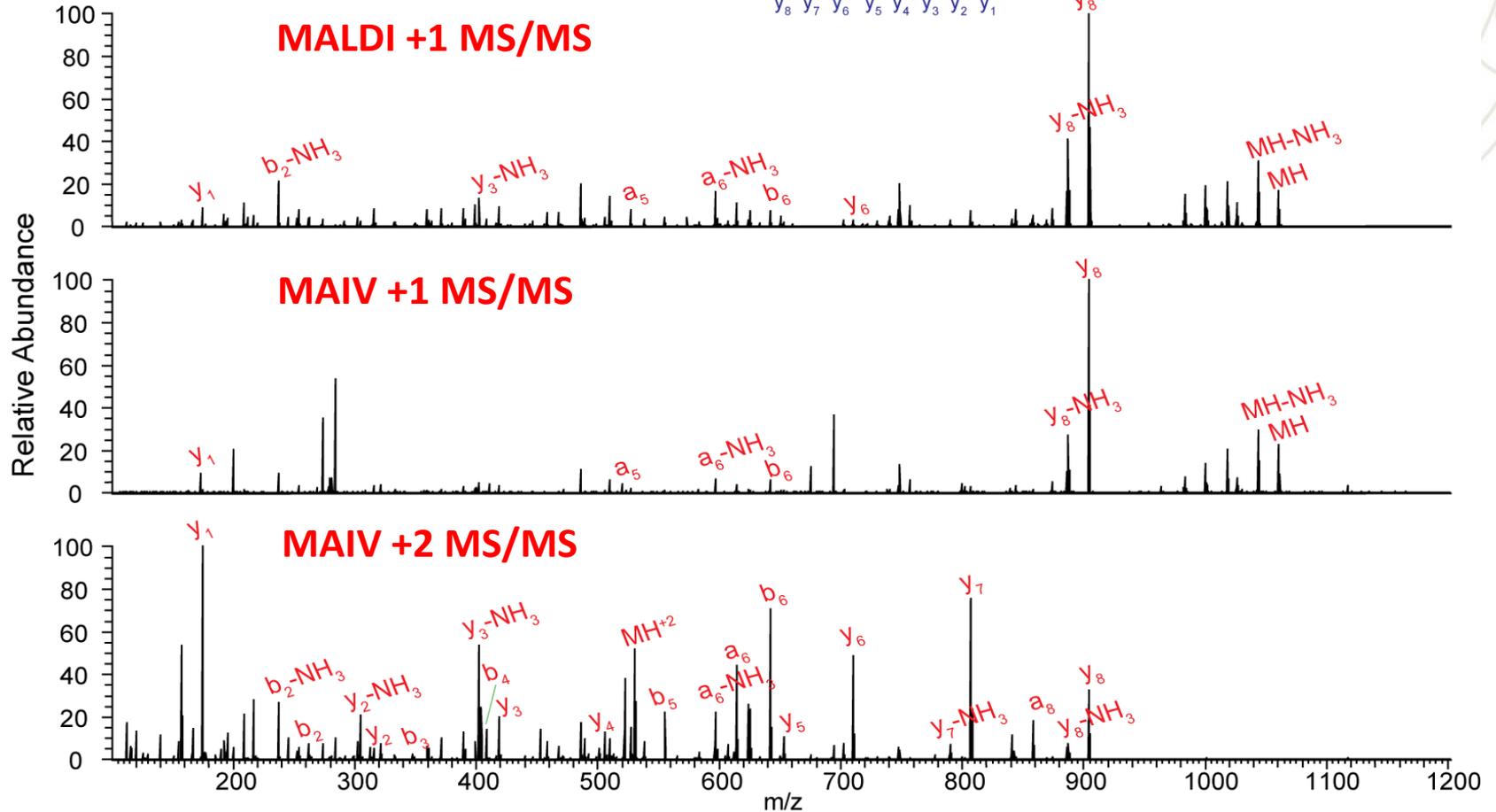
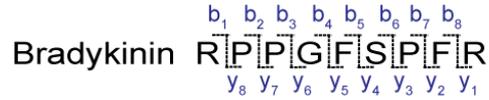
- Myoglobin 17.0 kDa



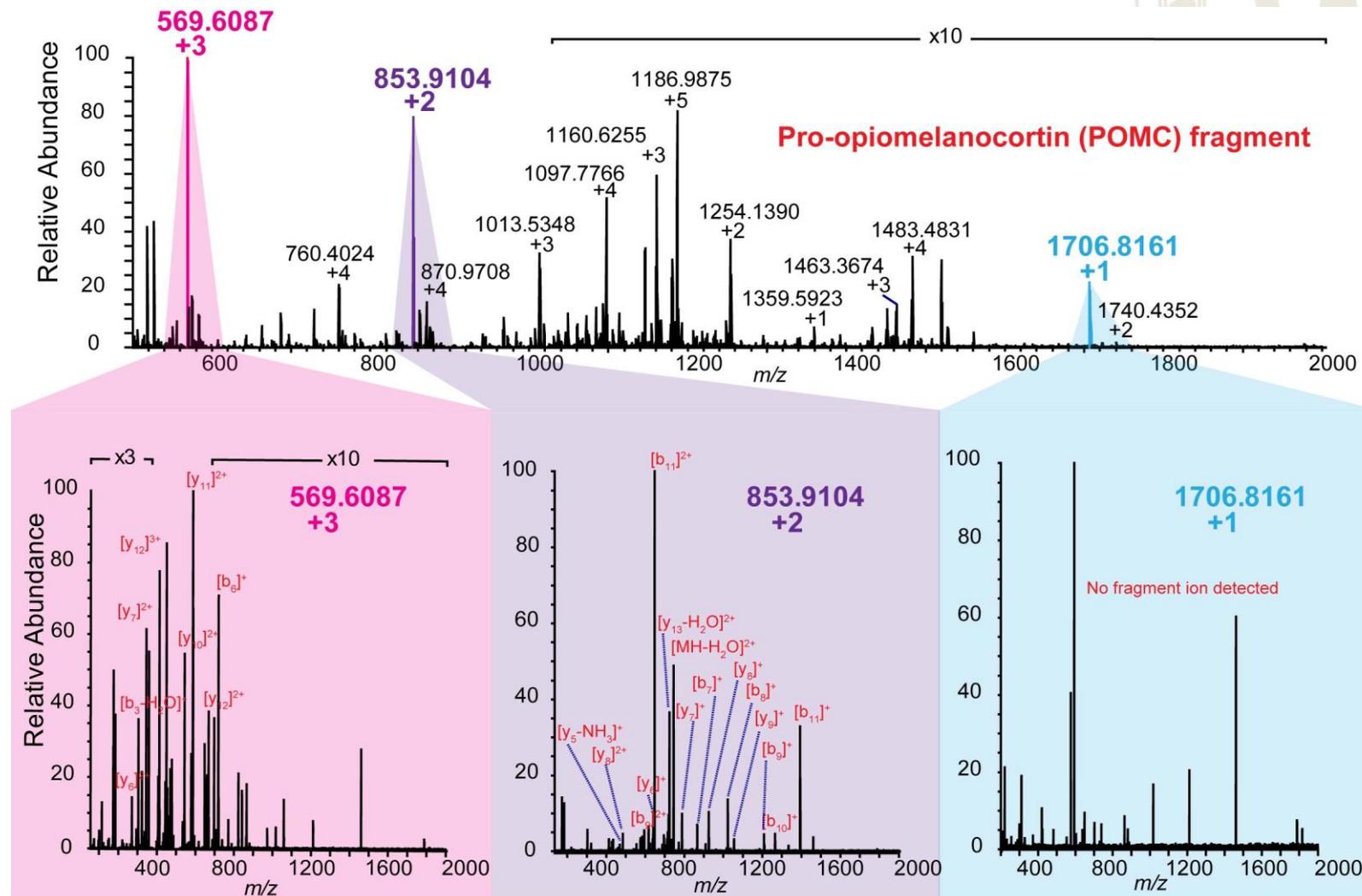
- Rat brain protein extraction



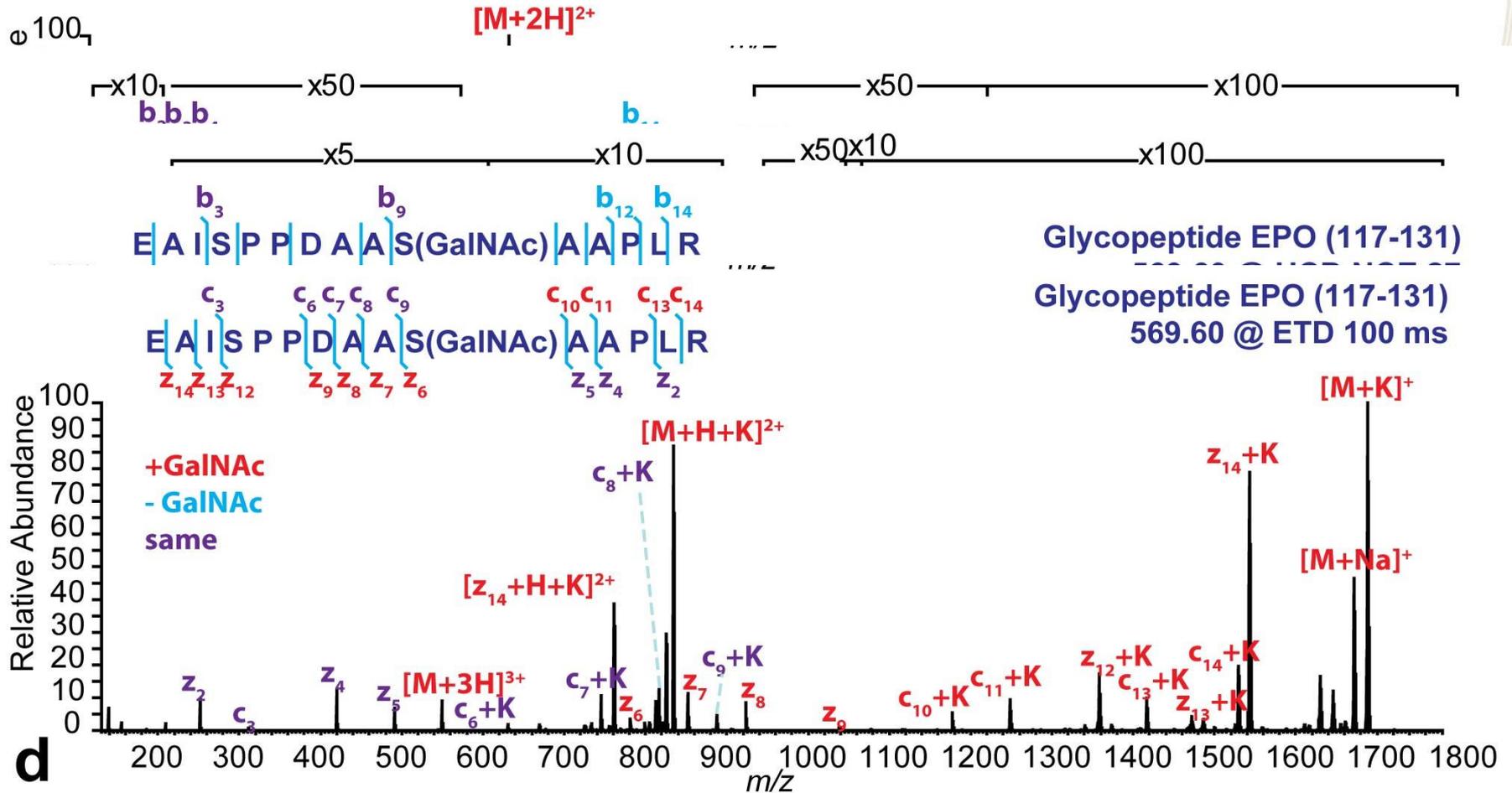
Improved Fragmentation: Bradykinin



Improved Fragmentation Efficiency



Labile Post-Translational Modification Analysis



Project 1 & 2. HR² AP/MALDI MS Conclusions

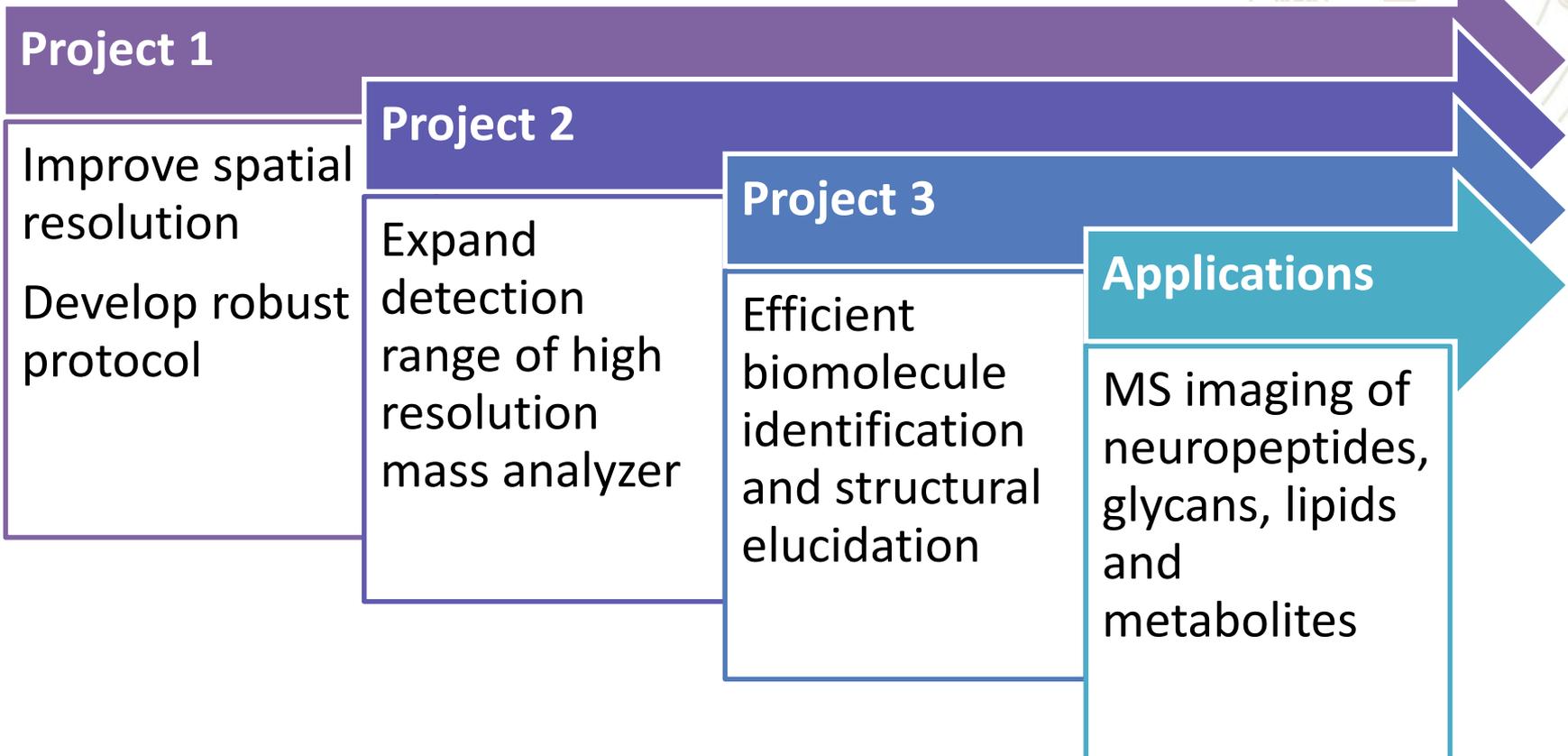
Achieved HR² MS imaging

- In mass (240K at m/z 200)
- In space (< 10 μm)

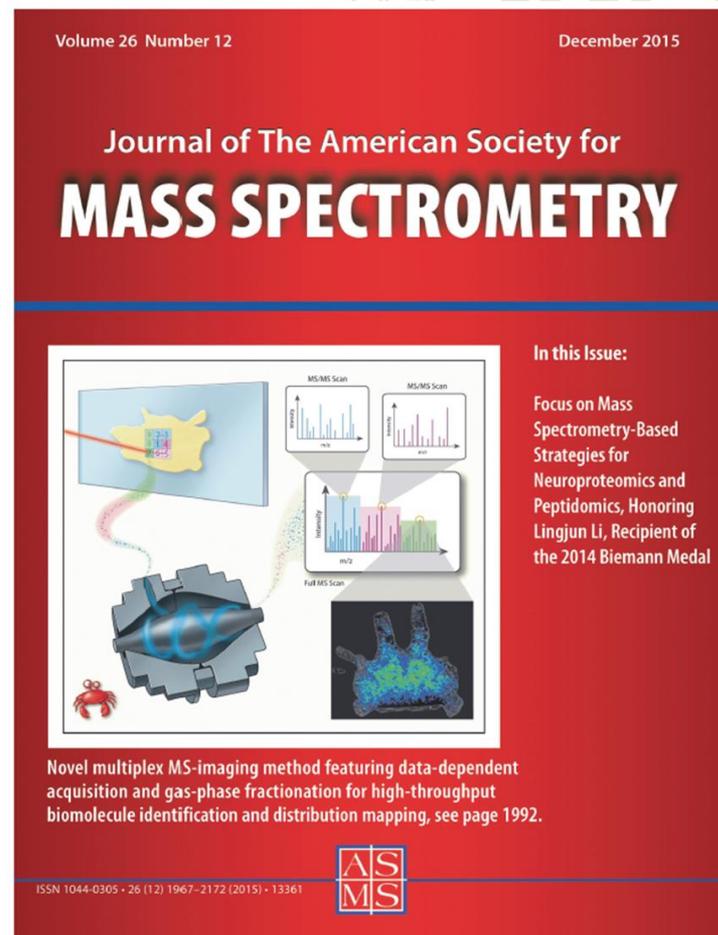
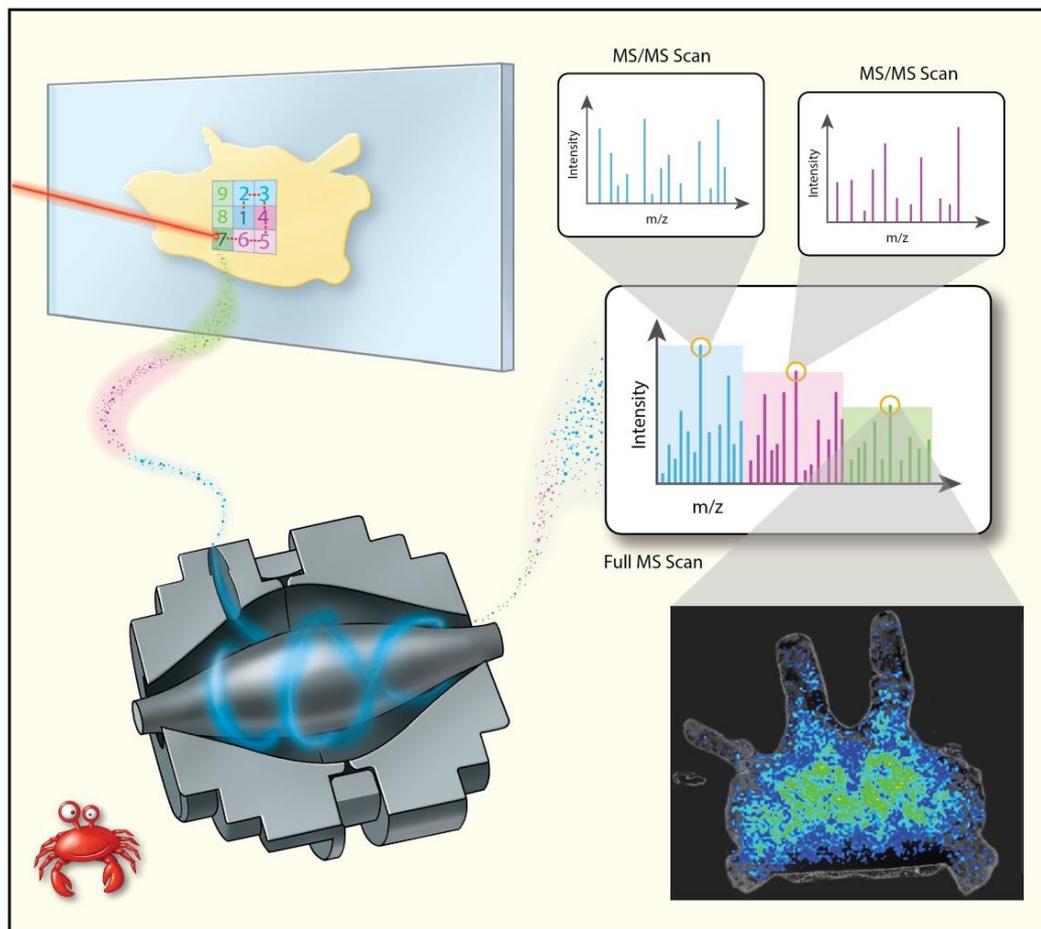
Produced multiply charged ions

- Mass range expanded for protein detection
- Fragmentation efficiency improved
- Possible to study labile PTMs

Ongoing Interest in MS Imaging



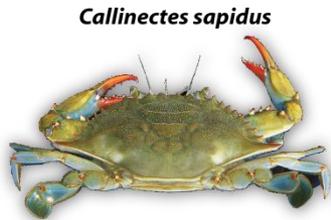
Project 3. High Throughput *In Situ* DDA Analysis of Neuropeptides by Coupling Novel Multiplex MS Imaging with Gas Phase Fractionation



Chen[#], Ouyang[#] and Li (2015). High Throughput *In situ* DDA Analysis of Neuropeptides by Coupling Novel Multiplex Mass Spectrometric Imaging (MSI) with Gas-Phase Fractionation. *JASMS*, vol. 26 (12). p.1992-2001. Cover article ([#]Co-First authors), artwork by Sally Griffith-Oh

MSI Study of the Crustacean Stomatogastric Nervous System (STNS) by MALDI Orbitrap

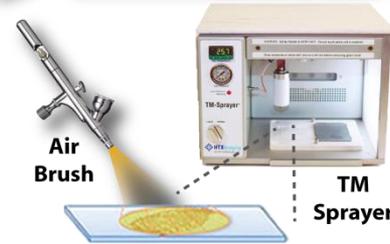
Animal sacrifice and tissue harvest



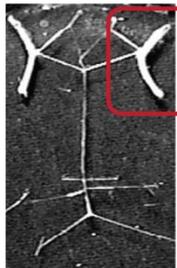
Tissue sections preparation



DHB Matrix application



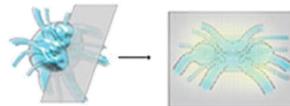
Database search and De Novo sequencing



Commissural ganglia (CoG)

Stomatogastric nervous system (STNS)

Section the tissue into 12µm slices on cryostat



MS imaging acquisition



Distribution mapping

Data analysis

Spiral step: 50µm

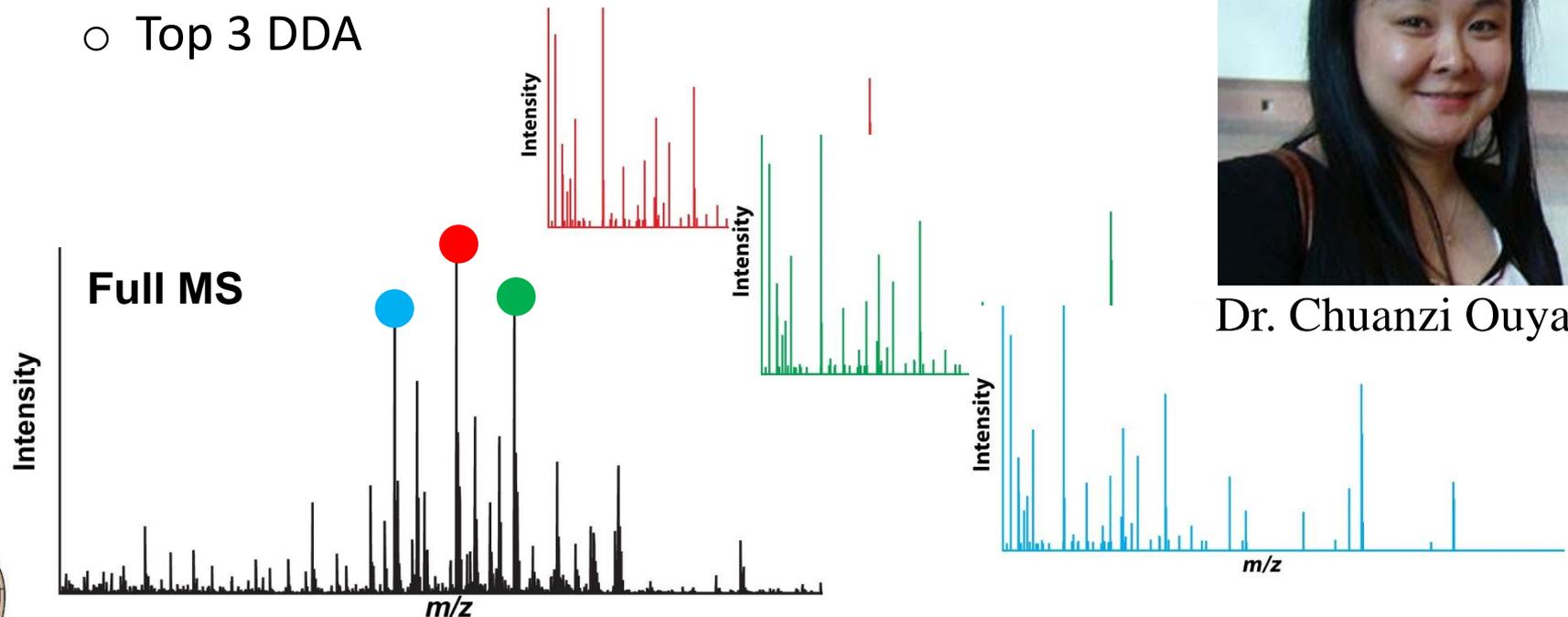


Raster step: 150 µm

Spiral step 1: full MS scan in Orbitrap
Spiral steps 2-9: targeted tandem MS in ion trap

In Situ Biomolecule Identification

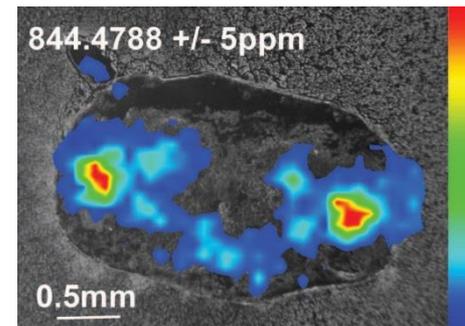
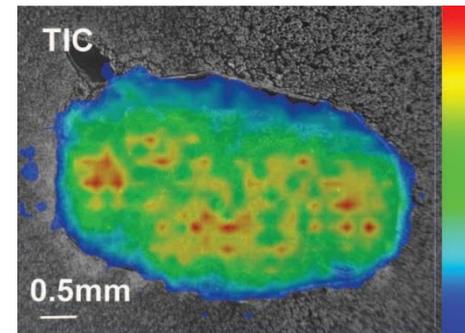
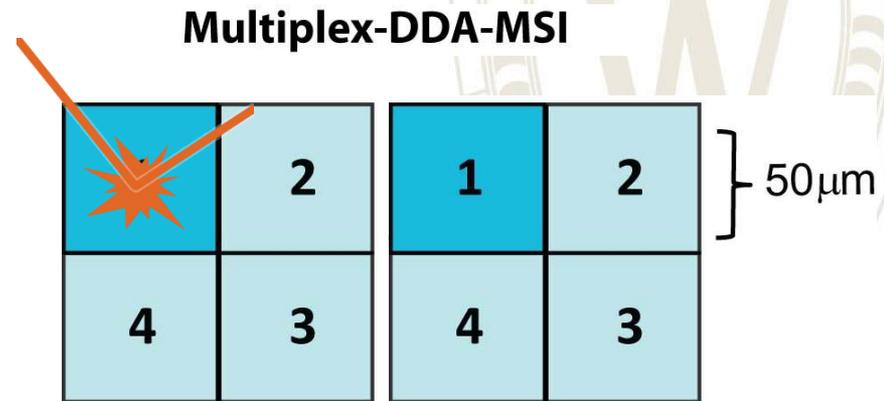
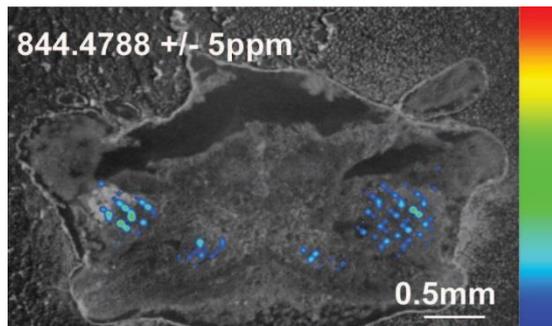
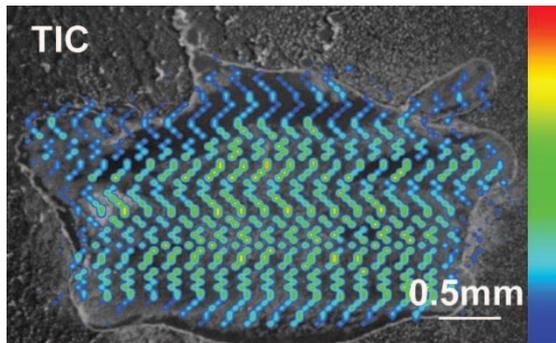
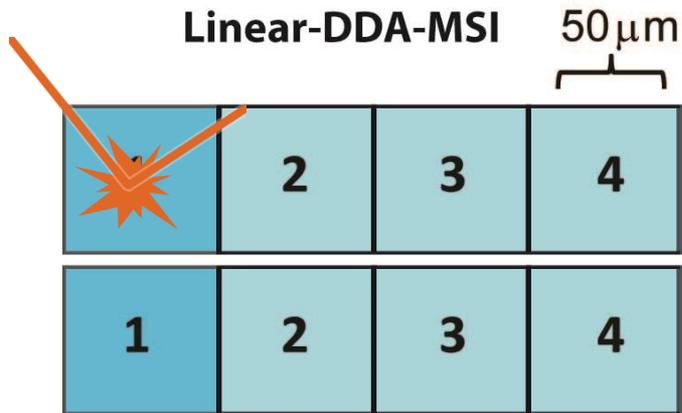
- **A major challenge in MS imaging**
 - Accurate mass matching: putative identification
 - Targeted MS/MS: confident identification and confirmation
- **Data dependent acquisition (DDA)**
 - Fragment top N ions after a full scan
 - Top 3 DDA



Dr. Chuanzi Ouyang



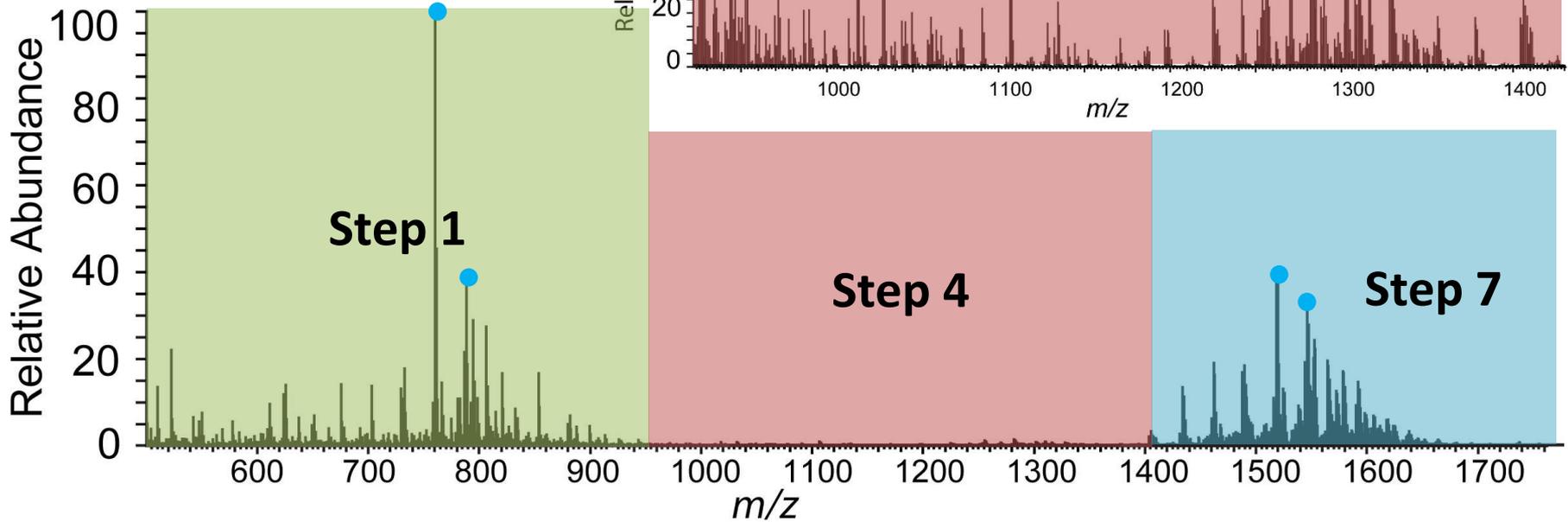
In Situ DDA: Linear vs. Multiplex MSI



Improving MS² Precursor Selection Efficiency

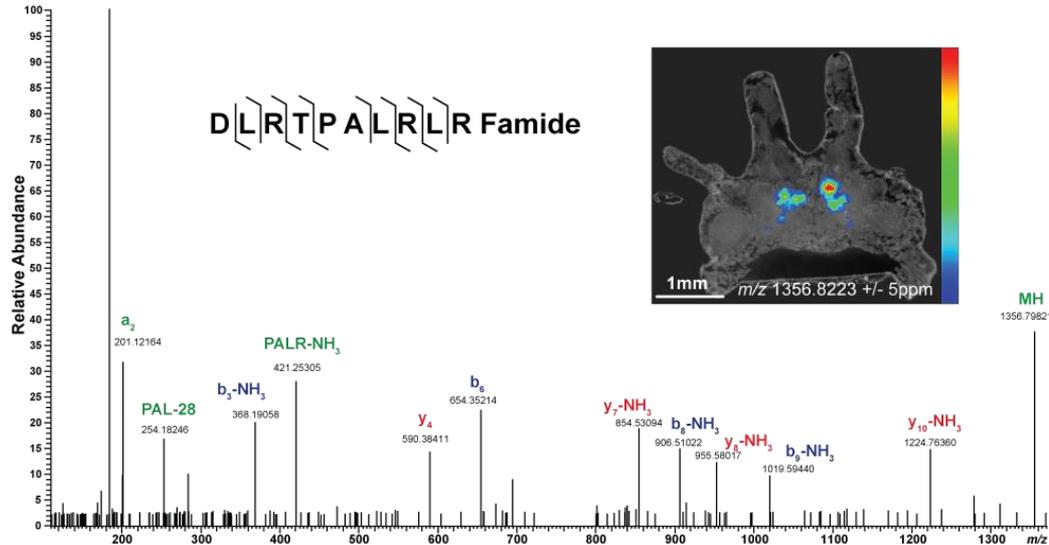
9	2	3
8	1	4
7	6	5

Fraction mass DDA



Multiplex DDA MSI Conclusions

- Simultaneous localization and identification
- Pseudo gas phase separation improved the precursor selection coverage
- Detection of novel neuropeptides by *de novo* sequencing
- **18** novel neuropeptides from crustacean neural tissue were identified by *in situ* MS/MS.

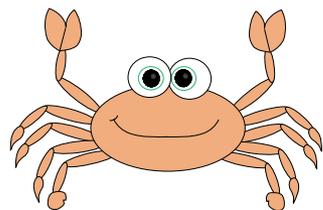


Novel RFamide identified in brain tissue of blue crab



MALDI MSI Workflow

➤ Workflow to achieve complementary metabolite coverage



Organ Dissection



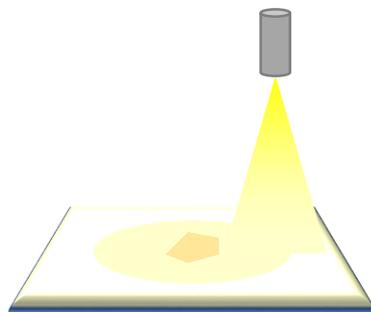
Gelatin Embedding



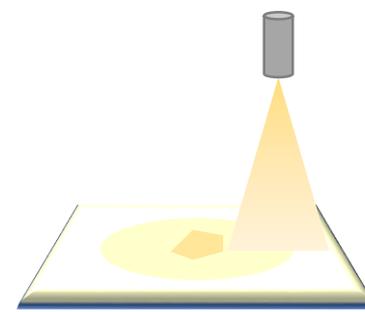
Sectioning to Slices



Imaging w/ MALDI-Orbitrap



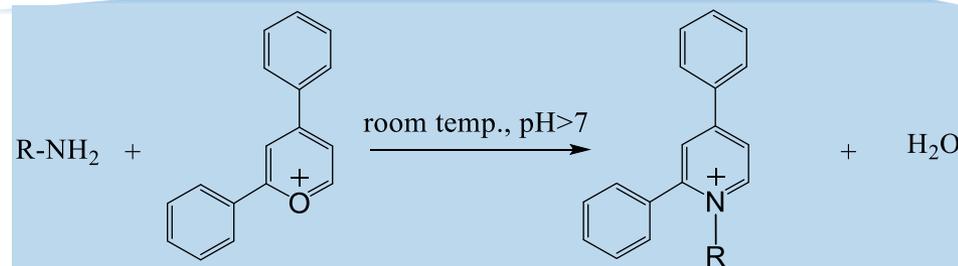
Spraying Matrix



DPP-TFB Derivatization

➤ Metabolite MALDI MSI challenges

- Matrix interference
- Low ionization efficiency

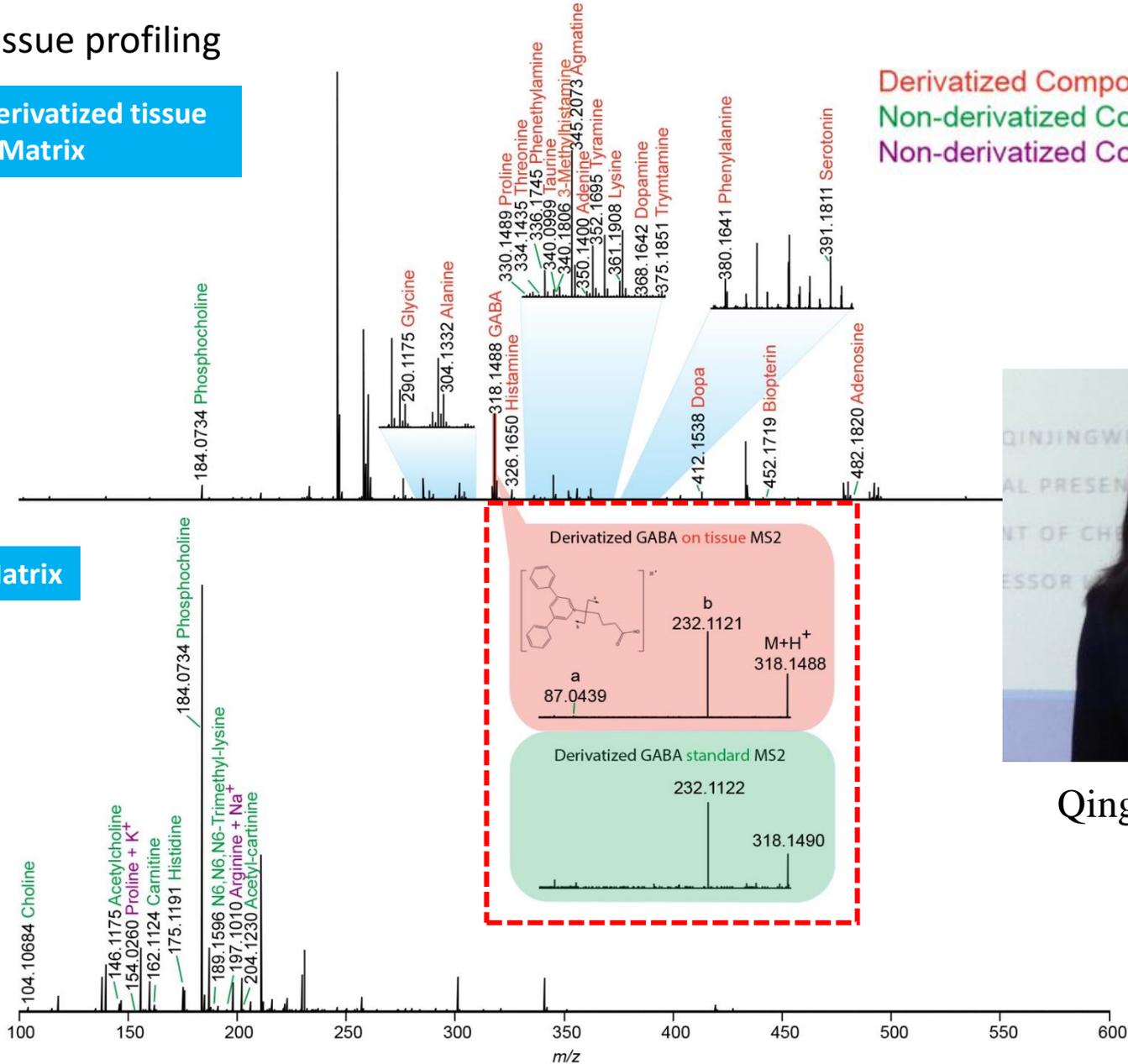


Results

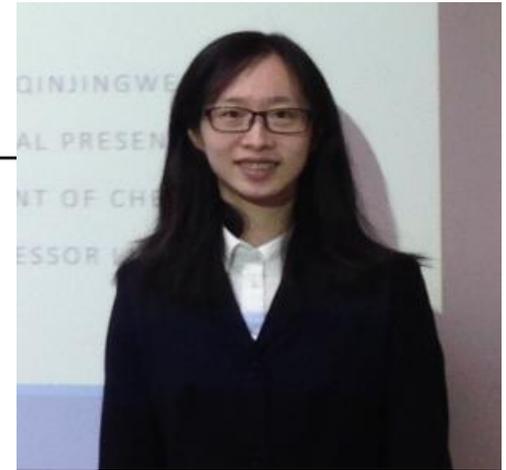
➤ On tissue profiling

Derivatized tissue
+ Matrix

Matrix



Derivatized Compound + DPP⁺
Non-derivatized Compound + H⁺
Non-derivatized Compound + Na⁺/K⁺



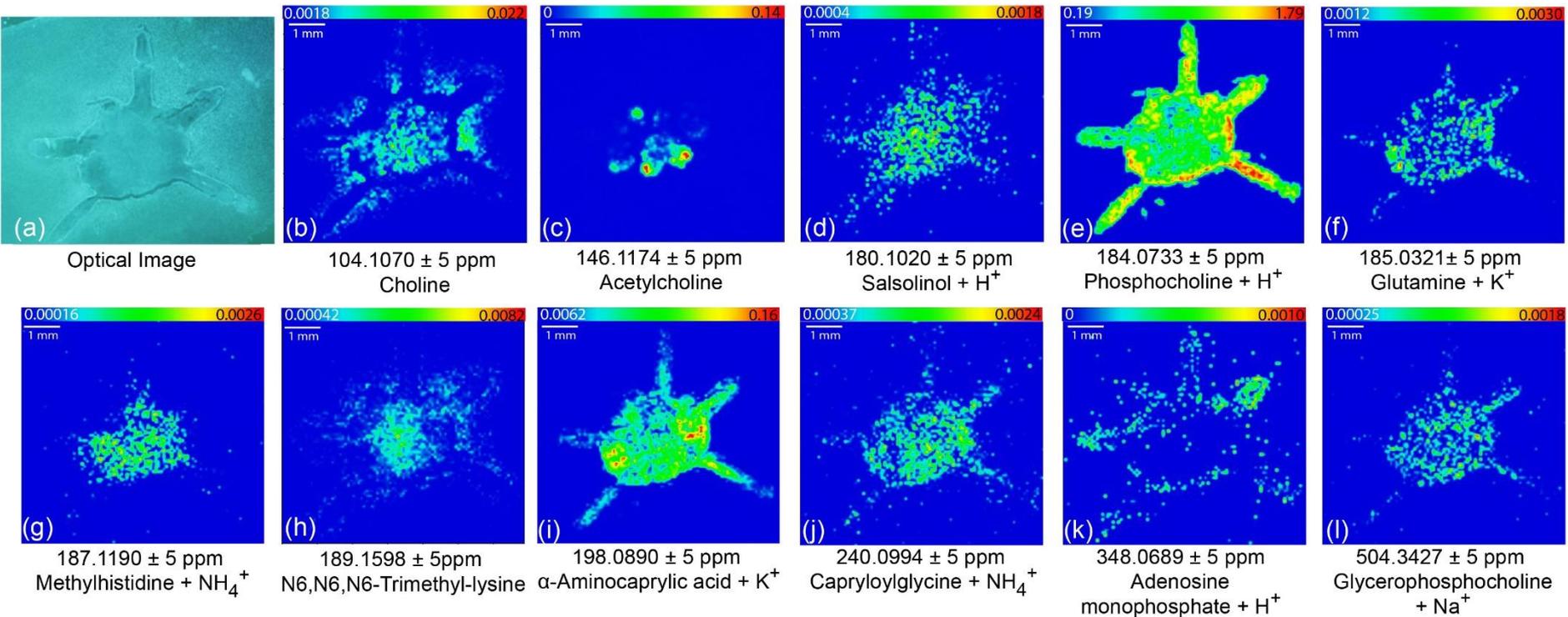
Qingjingwen Cao

Results

➤ Selected ion images from crab brain

Matrix

- Neurotransmitter: Acetylcholine
- Amino acids
- Lipids

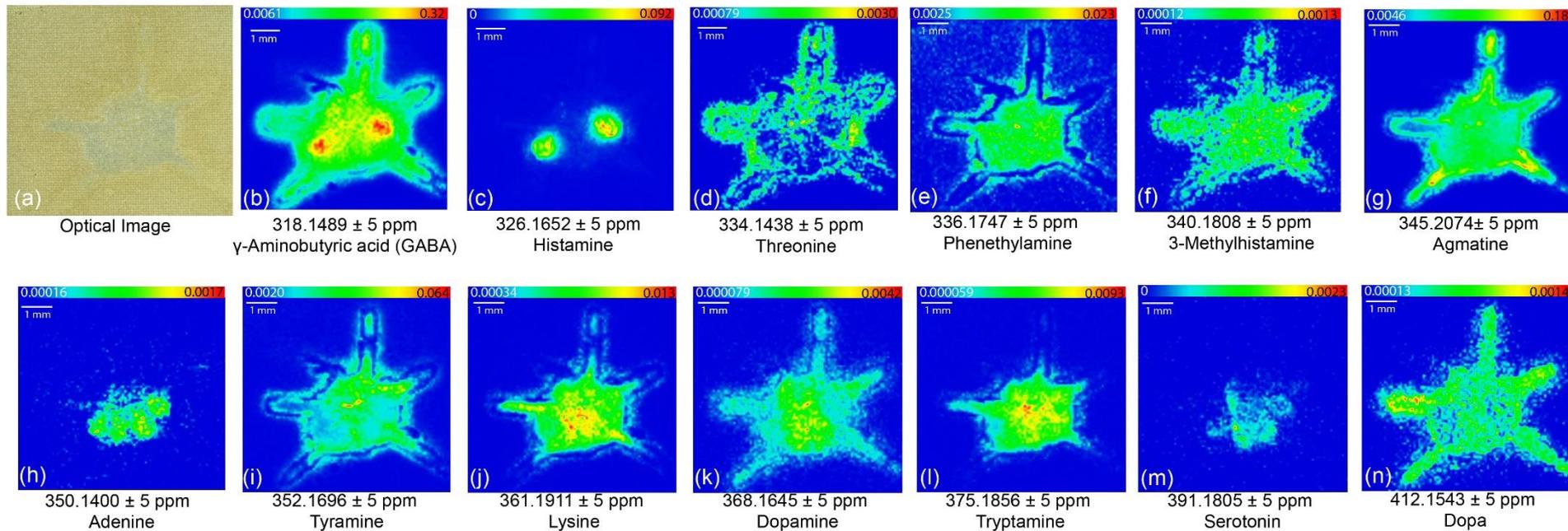


Results

➤ Selected ion images from crab brain

Derivatized tissue
+ Matrix

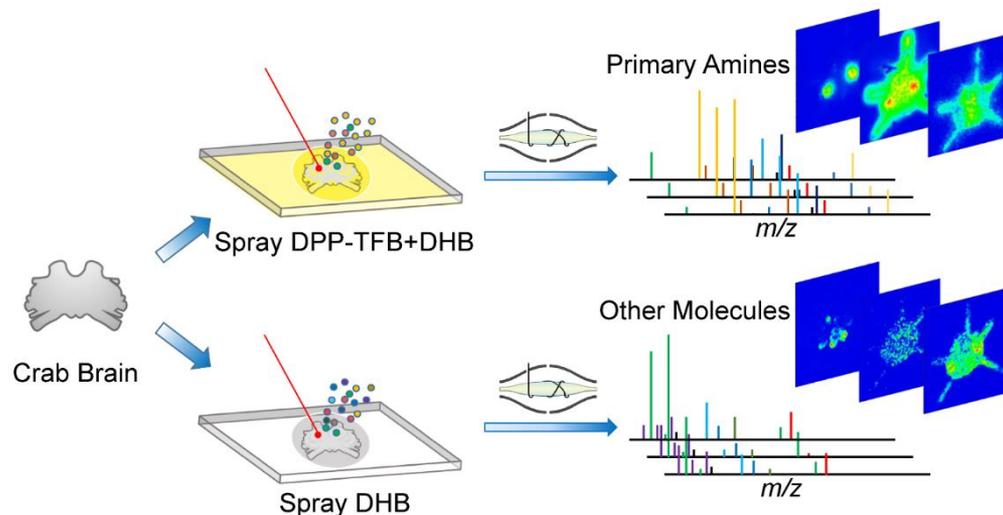
- Many more neurotransmitters!



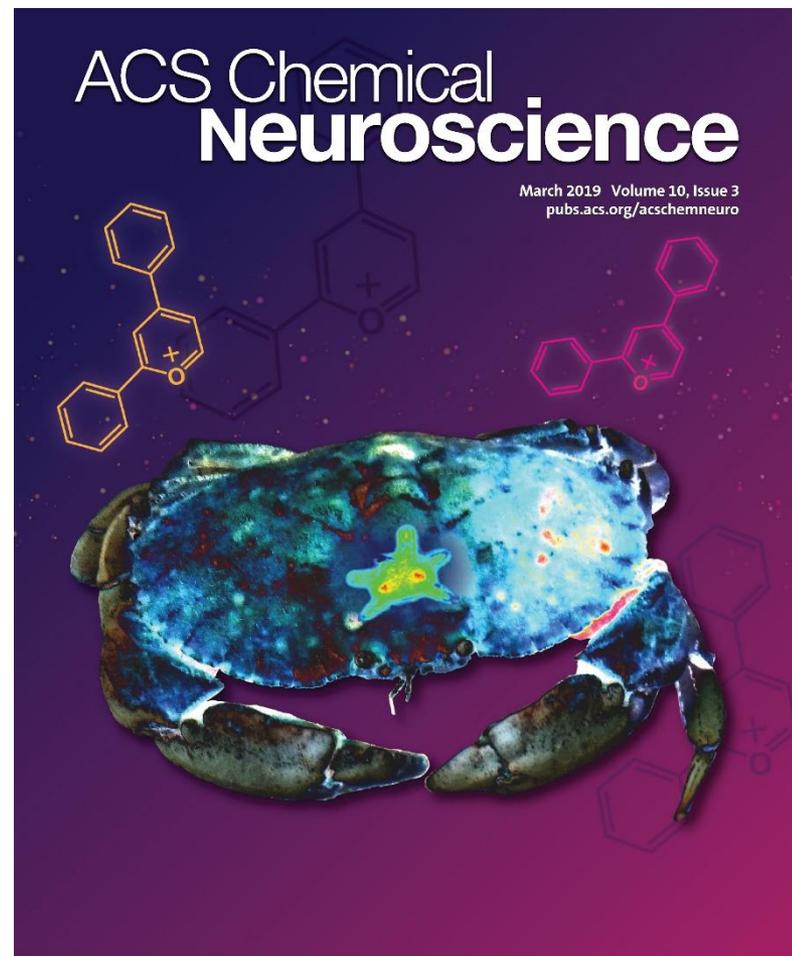
MALDI MSI Visualization and Identification of Neurotransmitters in Crustacean Brain



THE UNIVERSITY
of
WISCONSIN
MADISON



- On tissue chemical derivatization and reaction-free approaches enabled complementary signaling molecule visualization on crab brain sections via MALDI-LTQ-Orbitrap XL platform.
- Pyrylium salt served as a primary amine derivatization reagent and produced prominent signal enhancement of multiple neurotransmitters, including dopamine, serotonin, γ -aminobutyric acid and histamine that were not detected in underivatized tissues.
- Molecules with other functional groups, such as acetylcholine and phosphocholine, were directly imaged after matrix application.

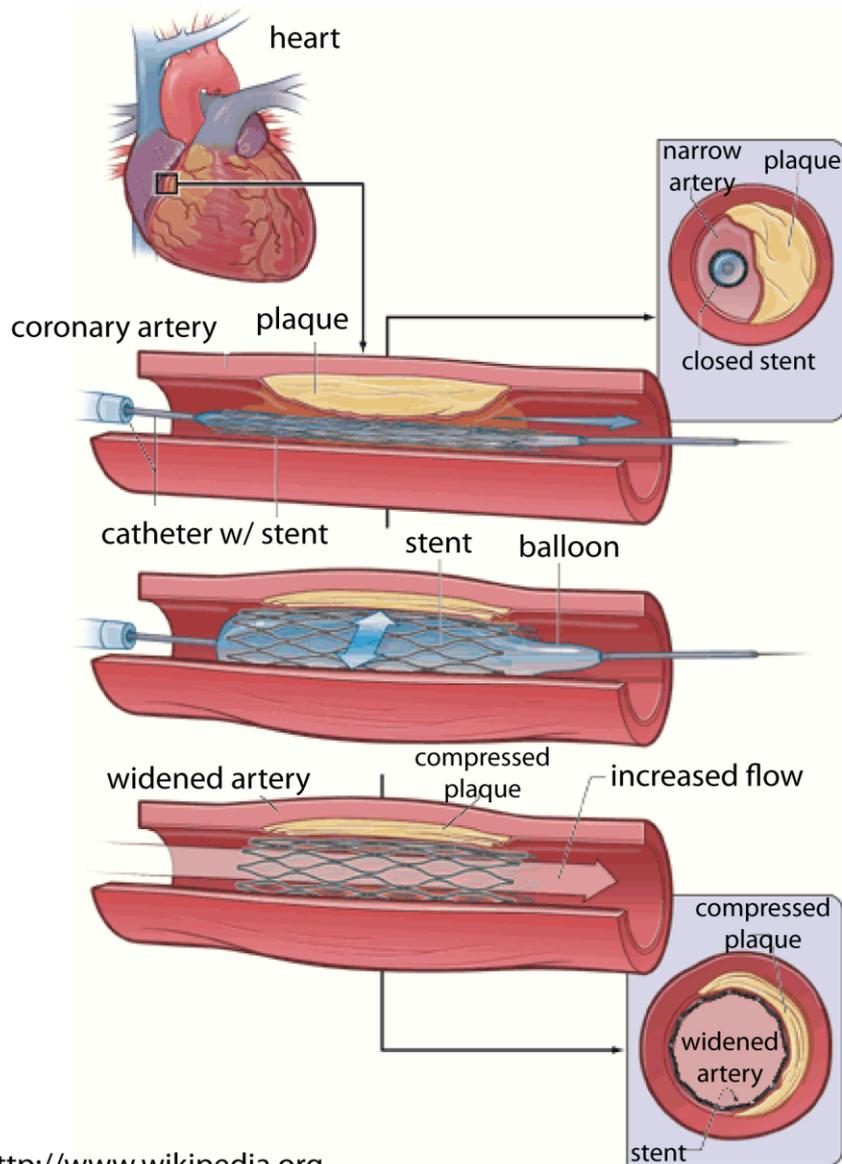


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Cao et al., *ACS Chem Neurosci* 2019 Mar 20;10(3):1222-1229. doi: 10.1021/acscchemneuro.8b00730.

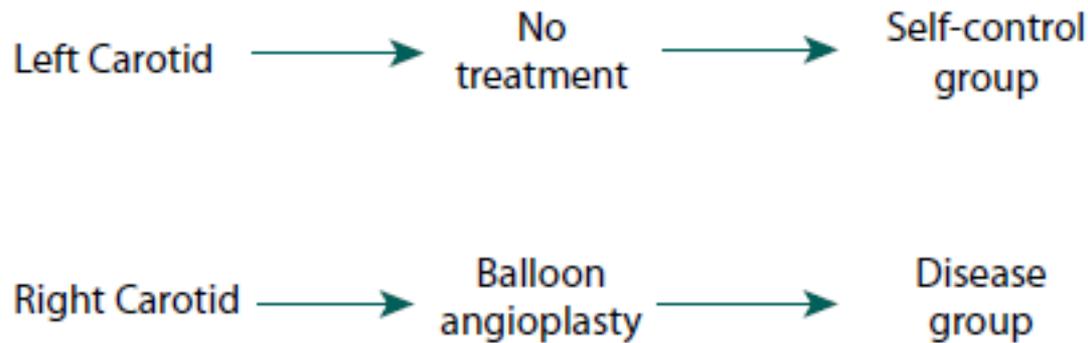
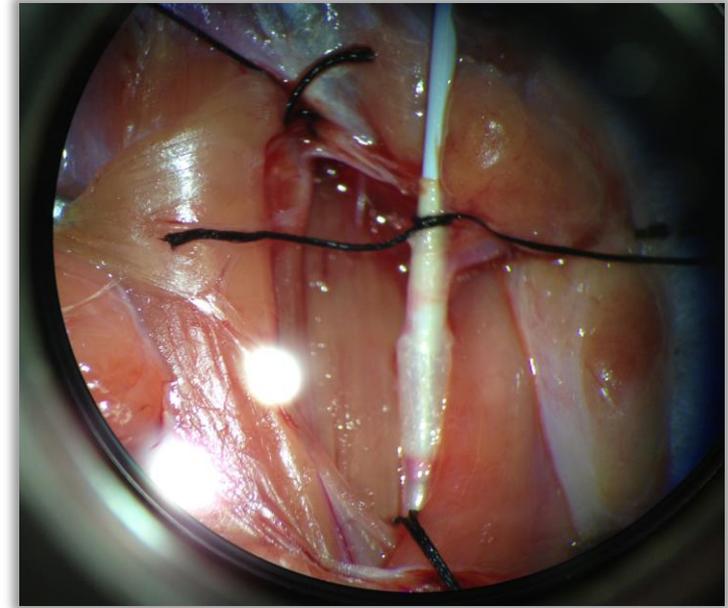
Restenosis: An unintended consequence



- Coronary atherosclerosis occurs when fatty plaques deposit on the interior wall of an artery and inhibits blood flow.
- Angioplasty is a common treatment wherein a balloon is inflated in the artery to clear the blockage.
- A stent is often placed after angioplasty to prevent future re-narrowing.

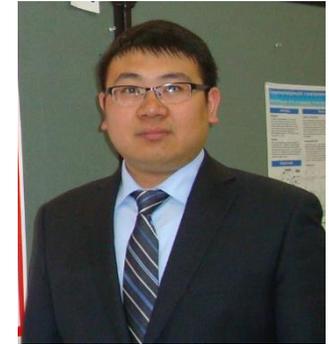
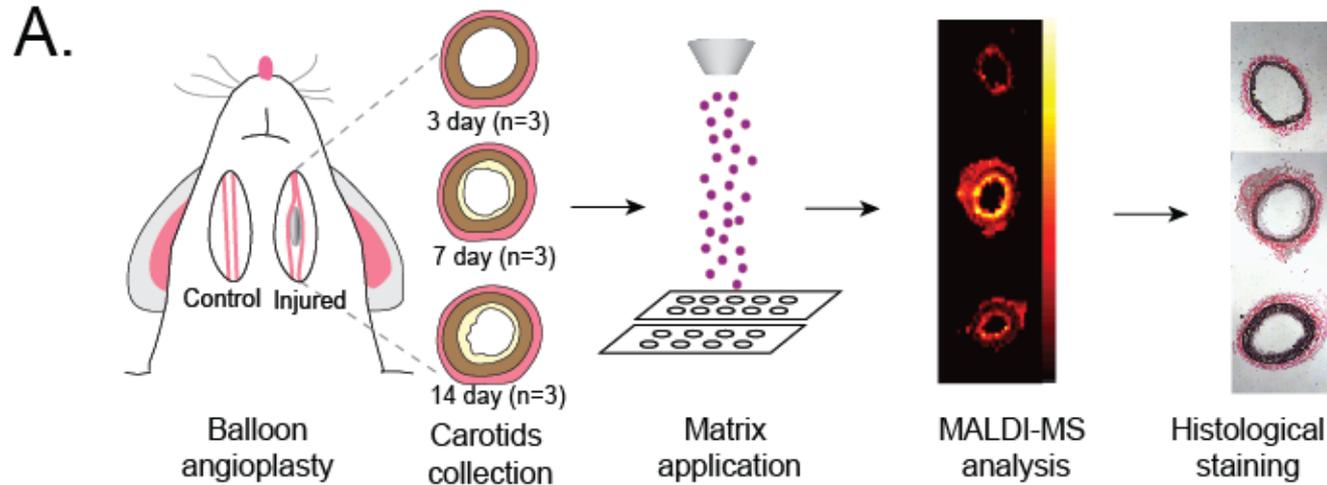
In collaboration with Dr. Craig Kent Lab

Rat Carotid Injury Model for Restenosis

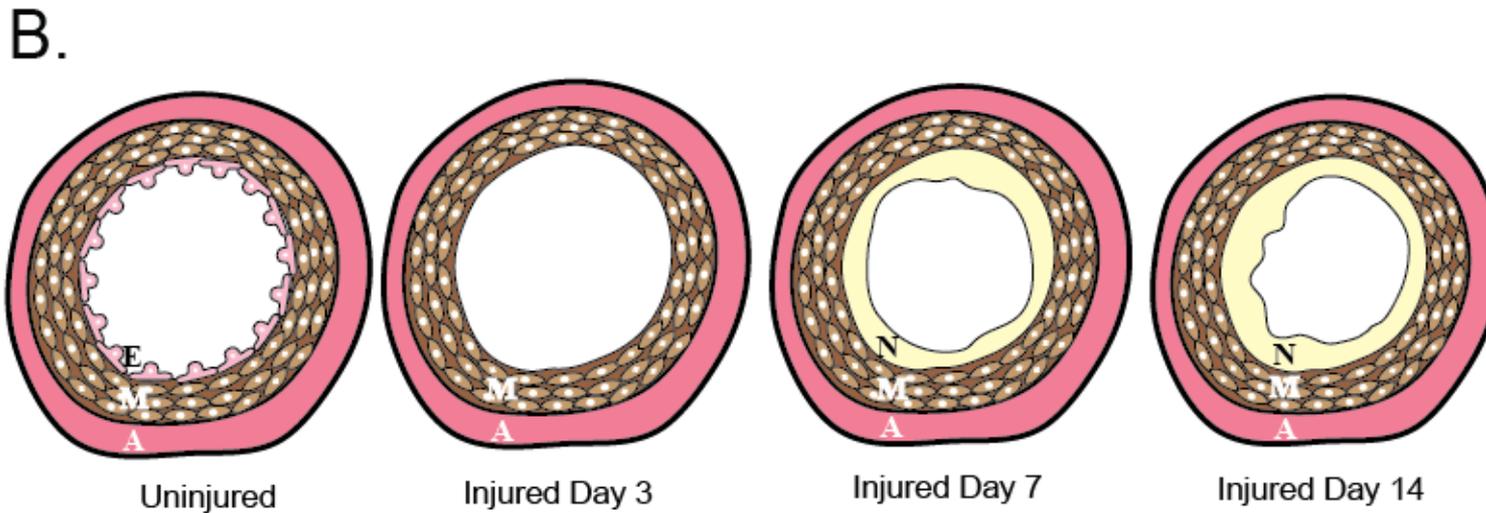


A rat model of restenosis is prepared by treating healthy rat carotid with balloon angioplasty. After surgery, the restenosis will gradually occur and rat carotid samples will be collected on 3, 7, 14 days after surgery, respectively.

MALDI MSI Workflow to Reveal Dynamic Changes of Signaling Lipids in Restenosis



Yatao Shi



Jill Johnson

m/z

3 days

7 days

14 days

control

disease

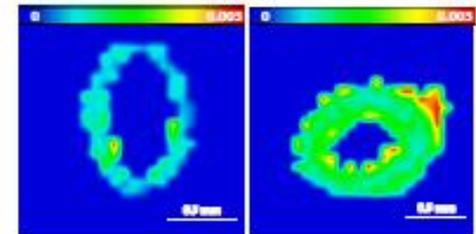
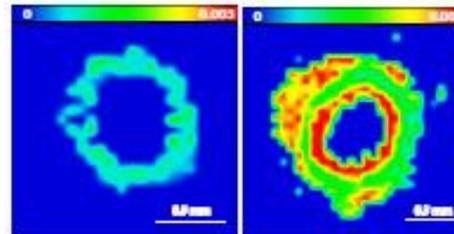
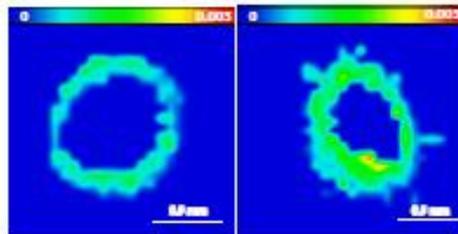
control

disease

control

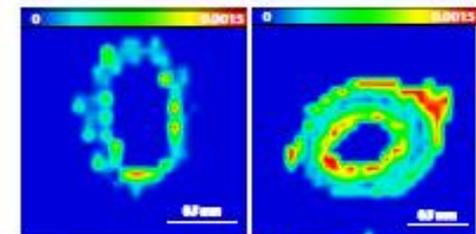
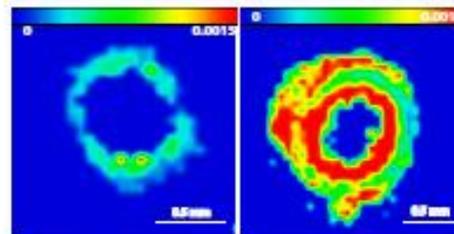
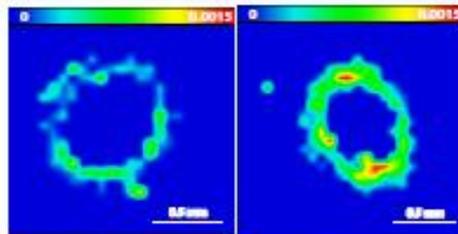
disease

551.5043
DAG(32:0)

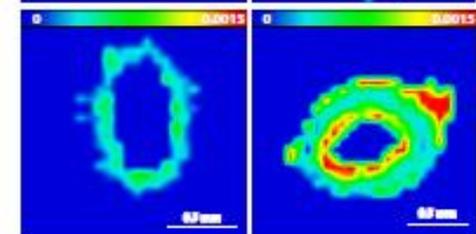
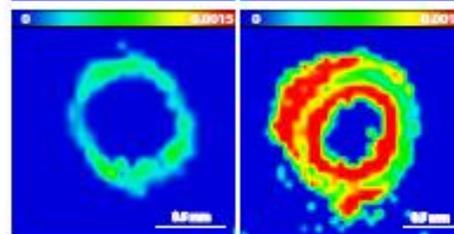
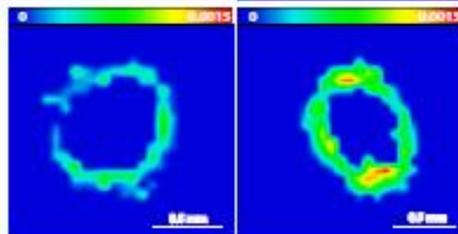


DAGs

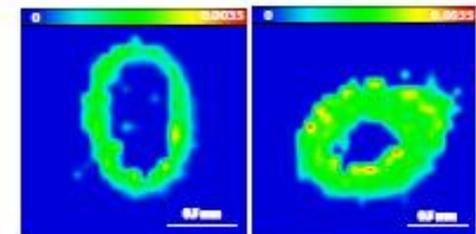
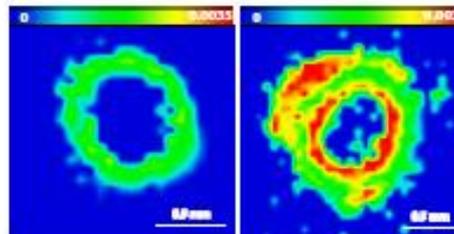
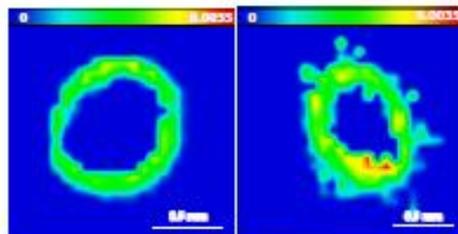
575.5035
DAG(34:2)



603.5352
DAG(36:2)

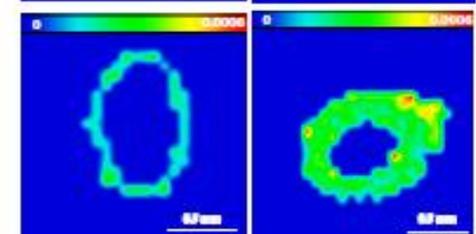
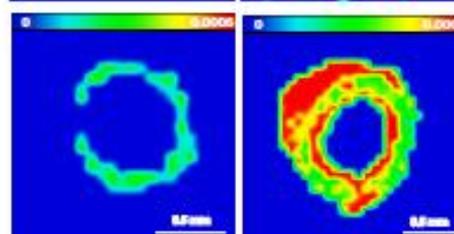
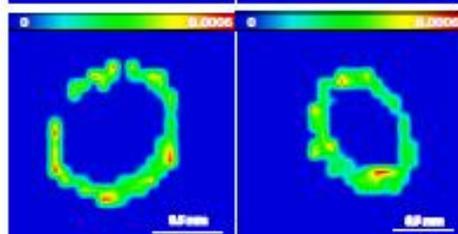


478.3292
LysoPC(16:0)

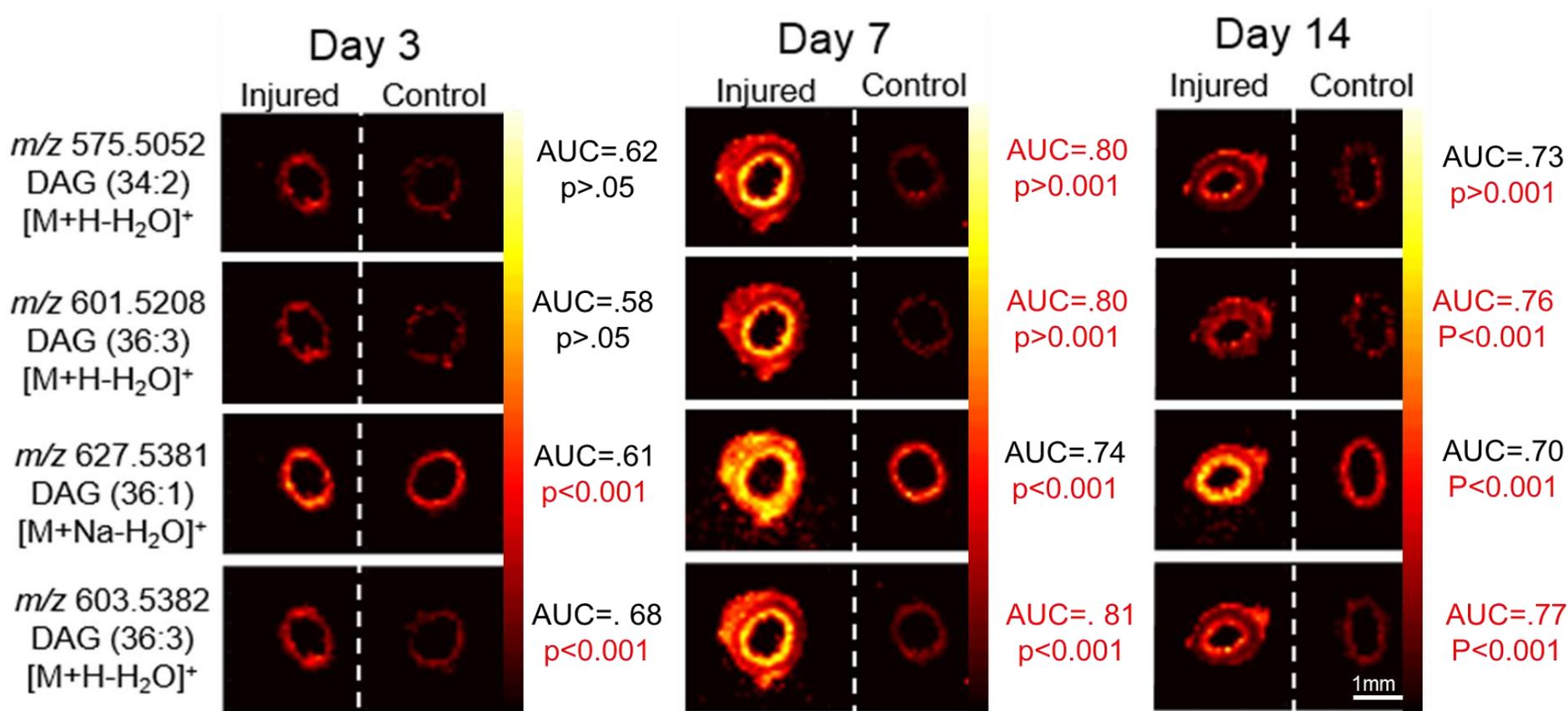


LysoPCs

504.3447
LysoPC(18:1)



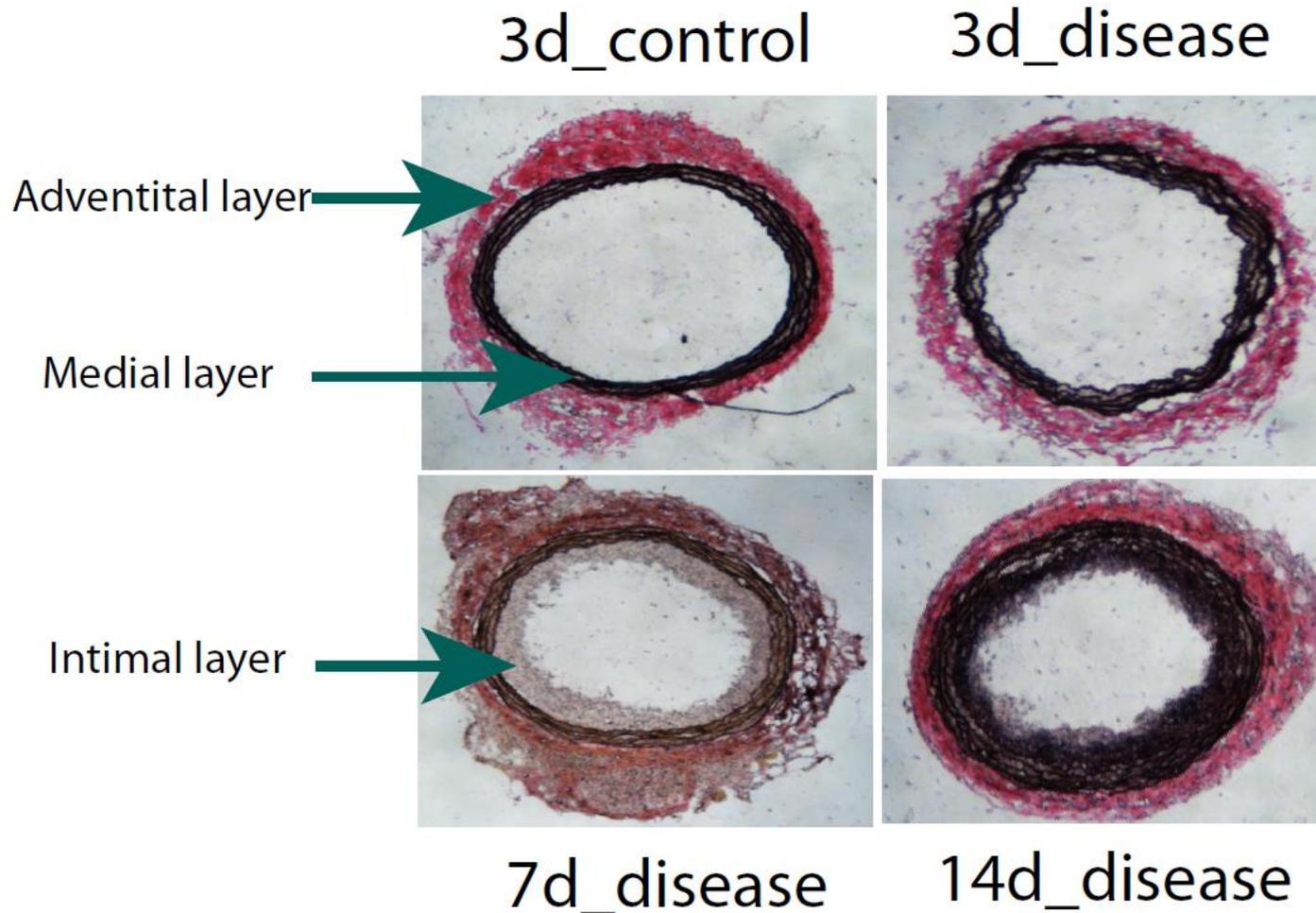
Diacylglycerols Upregulation: Statistics



Mass Spectrometric Imaging Reveals Temporal and Spatial Dynamics of Bioactive Lipids in Arteries Undergoing Restenosis.

Shi Y, Johnson J, Wang B, Chen B, Fisher GL, Urabe G, Shi X, Kent KC, Guo LW, Li L. *J Proteome Res.* 2019 Apr 5;18(4):1669-1678. doi: 10.1021/acs.jproteome.8b00941. Epub 2019 Mar 11.

Verhoeff-van Gieson Staining



Bowen Wang



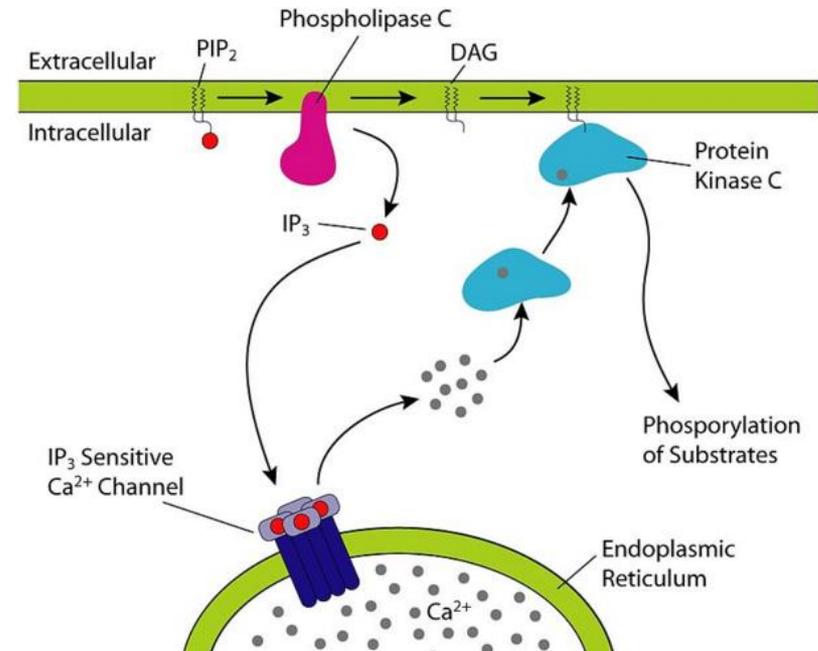
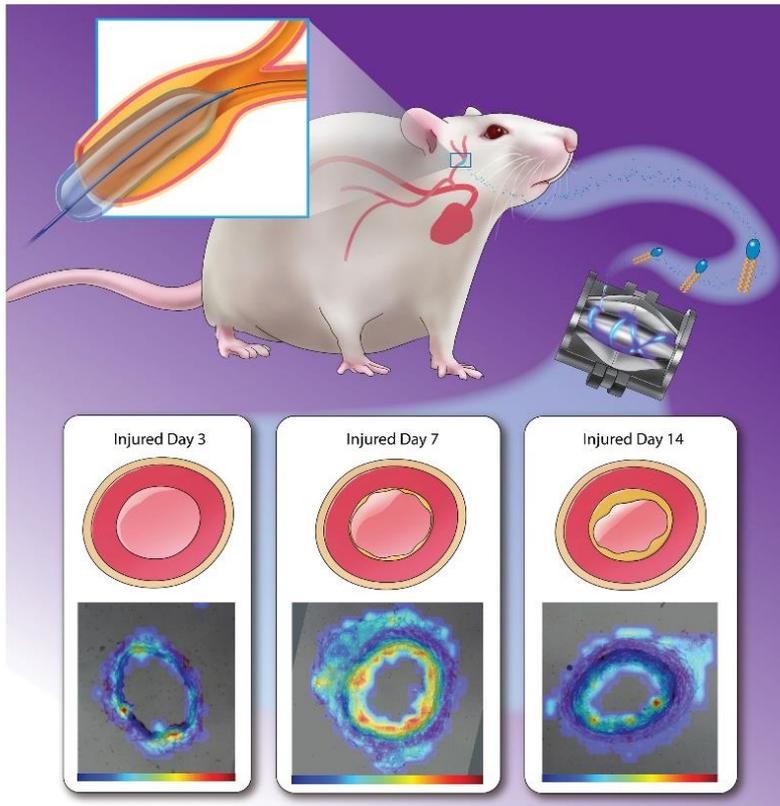
Dr. Xudong Shi

MALDI MSI Reveals Unique Neointimal Distributions of DAGs

Journal of
proteome
research

April 2019

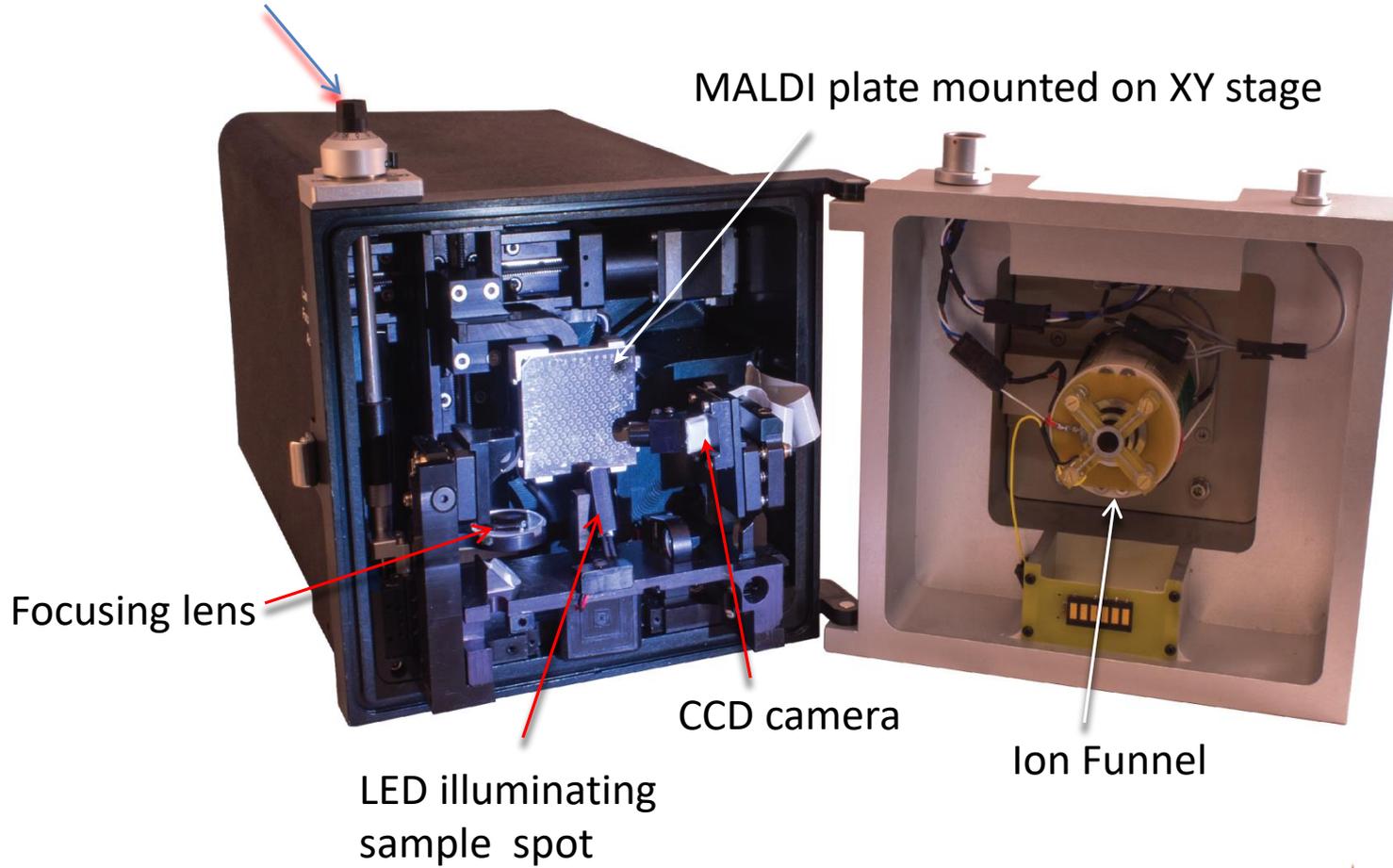
Volume 18 Number 4 pubs.acs.org/jpr



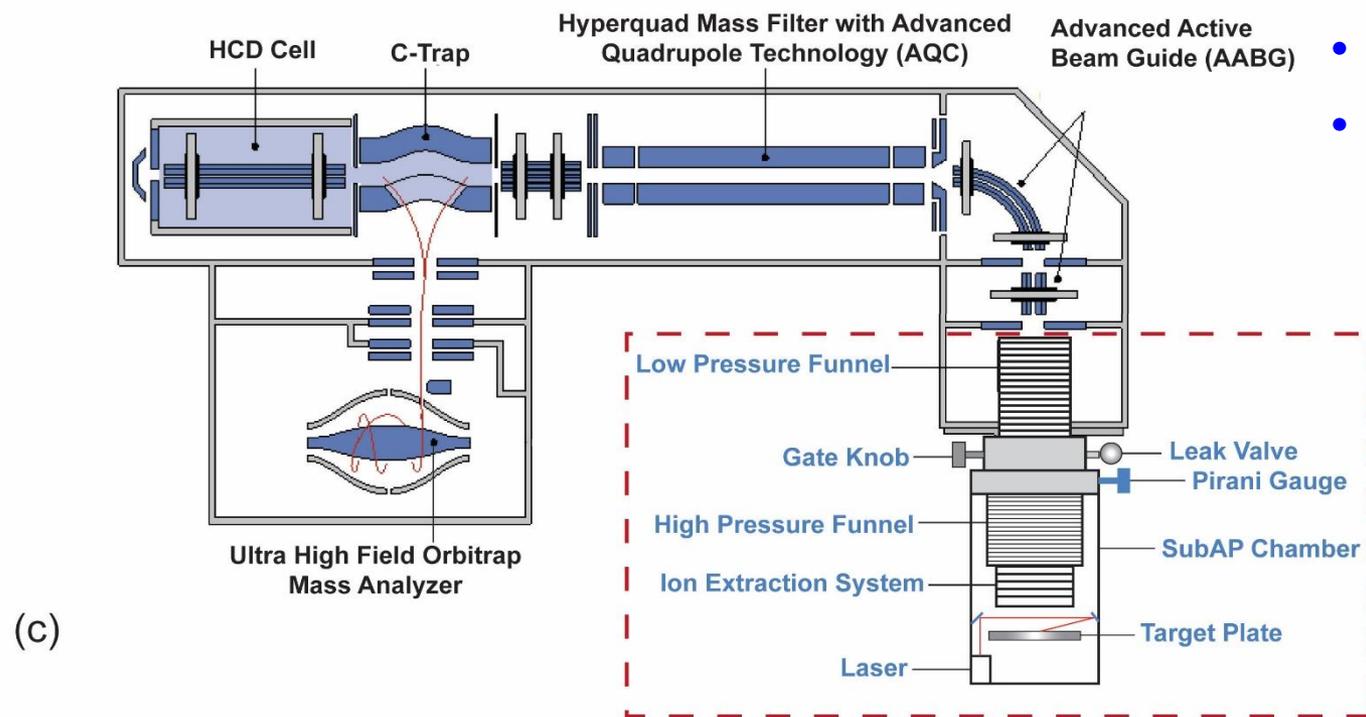
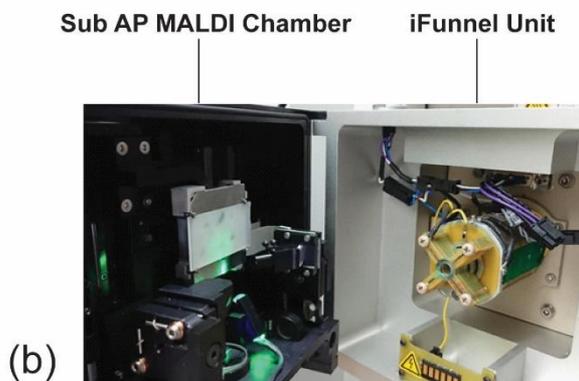
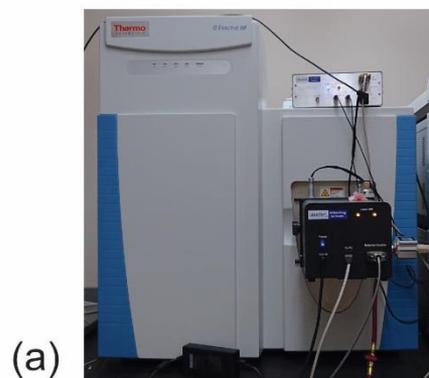
- DAGs were up-regulated during restenosis
- DAG-mediated signal pathway was proven to be involved in the process of restenosis
- LysoPCs are upregulated during the process of restenosis

SubAP-MALDI source

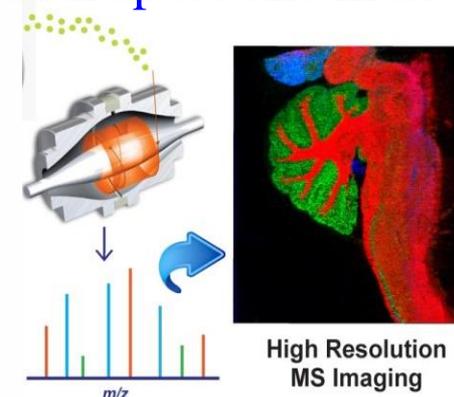
Dial controlling laser spot size
by changing a distance between focusing lens and sample plate



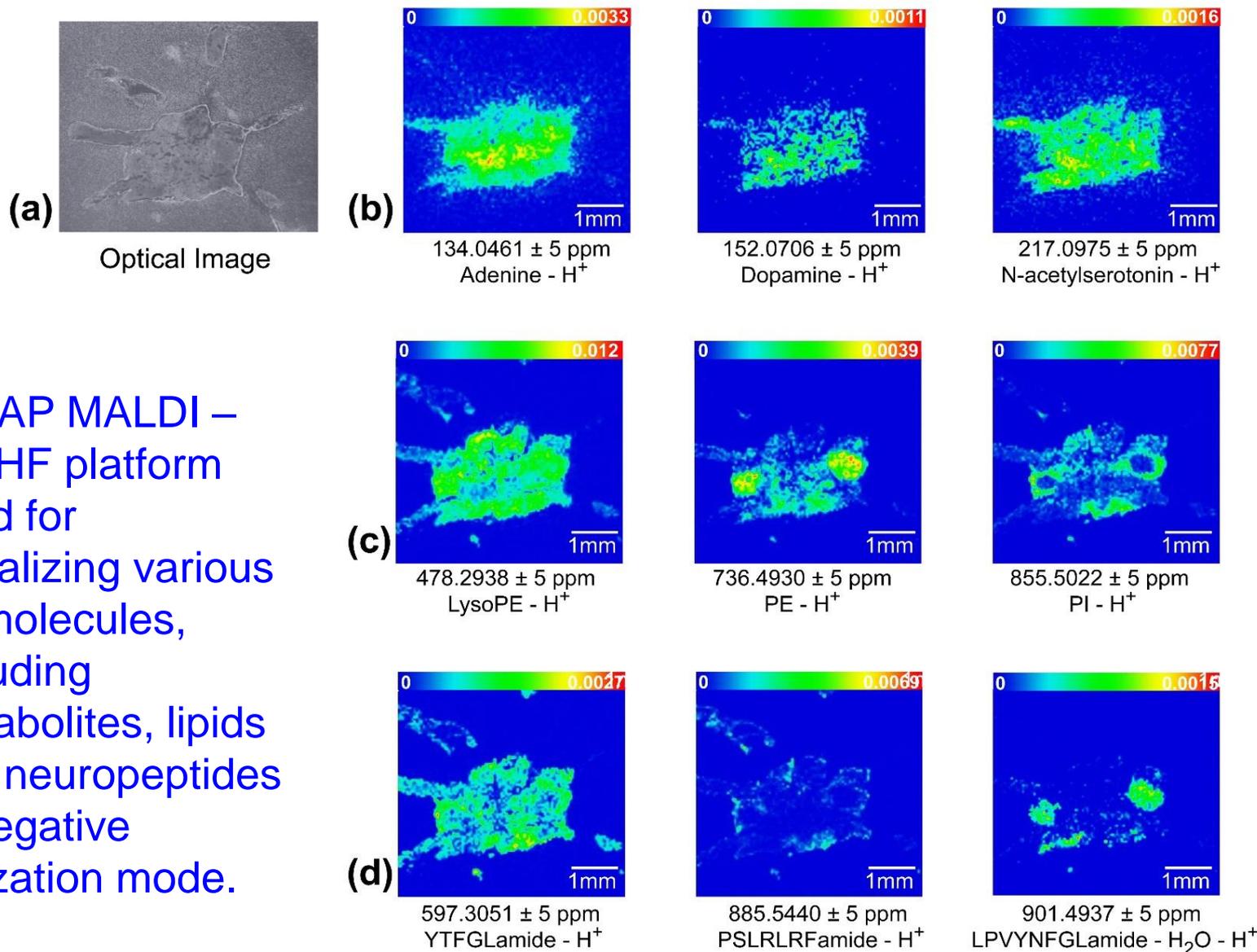
A High Resolution SubAP/MALDI(ng) UHR Source Coupled to a Quadrupole-Orbitrap Platform for MSI



- Developed by MassTech
- Solid state laser
- <10 um laser diameter
- Up to 10 kHz firing rate
- Operated at 3~5 Torr
- Interfaced to QE-HF to achieve high resolution imaging in both space and mass



A High Resolution SubAP/MALDI-Q-Orbitrap Platform Enables *In Situ* Analysis of Biomolecules by Negative Mode Ionization and Acquisition



SubAP MALDI –
QE HF platform
used for
visualizing various
biomolecules,
including
metabolites, lipids
and neuropeptides
in negative
ionization mode.



Comparison of Vacuum MALDI and AP-MALDI Platforms for the Mass Spectrometry Imaging of Metabolites Involved in Salt Stress in *Medicago truncatula*

Caitlin Keller¹, Junko Maeda², Dhileepkumar Jayaraman³, Sanhita Chakraborty⁴, Michael R. Sussman⁵, Jeanne M. Harris⁴, Jean-Michel Ané^{2,3} and Lingjun Li^{1,6*}

¹ Department of Chemistry, University of Wisconsin–Madison, Madison, WI, United States, ² Department of Agronomy, University of Wisconsin–Madison, Madison, WI, United States, ³ Department of Bacteriology, University of Wisconsin–Madison, Madison, WI, United States, ⁴ Department of Plant Biology, University of Vermont, Burlington, VT, United States, ⁵ Department of Biochemistry, University of Wisconsin–Madison, Madison, WI, United States, ⁶ School of Pharmacy, University of Wisconsin–Madison, Madison, WI, United States

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Matrix-assisted laser desorption/ionization-mass spectrometry imaging (MALDI-MSI) is routinely used to determine the spatial distributions of various biomolecules in tissues. Recently, there has been an increased interest in creating higher resolution images using sources with more focused beams. One such source, an atmospheric pressure (AP) MALDI source from MassTech, has a laser capable of reaching spatial resolutions of 10 μm . Here, the AP-MALDI source coupled with a Q Exactive HF Orbitrap platform is compared to the commercial MALDI LTQ Orbitrap XL system using *Medicago truncatula* root nodules. AP-MALDI parameters, such as the S-lens value, capillary temperature, and spray voltage, were optimized on the Q Exactive-HF platform for optimal detection of plant metabolites. The performance of the two systems was evaluated for sensitivity, spatial resolution, and overall ability to detect plant metabolites. The commercial MALDI LTQ Orbitrap XL was superior regarding the number of compounds detected, as at least two times more m/z were detected compared to the AP-MALDI system. However, although the AP-MALDI source requires a spatial resolution higher than 10 μm to get the best signal, the spatial resolution at 30 μm is still superior compared to the 75 μm spatial resolution achieved on the MALDI platform. The AP-MALDI system was also used to investigate the metabolites present in *M. truncatula* roots and root nodules under high salt and low salt conditions. A discriminative analysis with SCiLS software



Dr. Caitlin Keller

A High Resolution AP/MALDI-Q-Orbitrap Platform Enables *In Situ* Analysis of Metabolites involved in *Medicago truncatula*

AP-MALDI-ORBITRAP-MSI

- Atmospheric pressure
- High spatial resolution

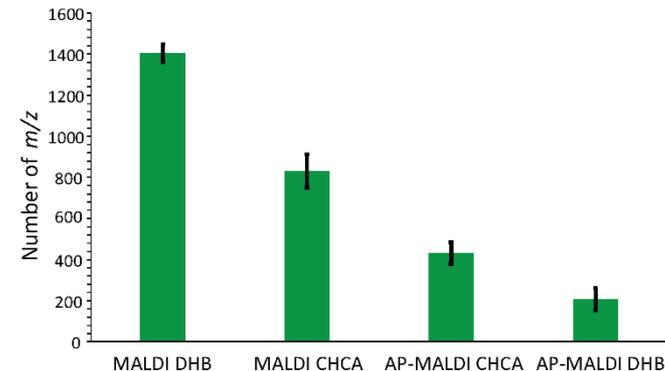


MALDI-Orbitrap-MSI

- Under vacuum
- Higher sensitivity



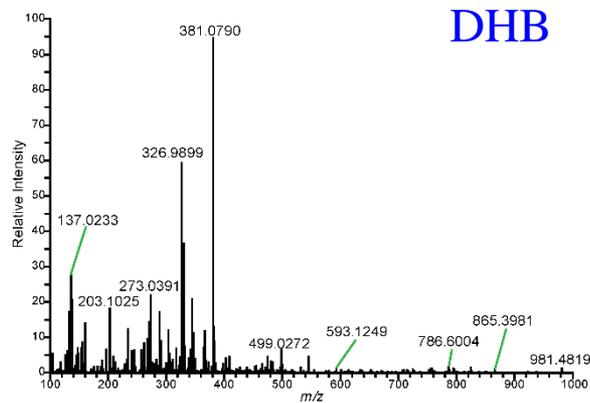
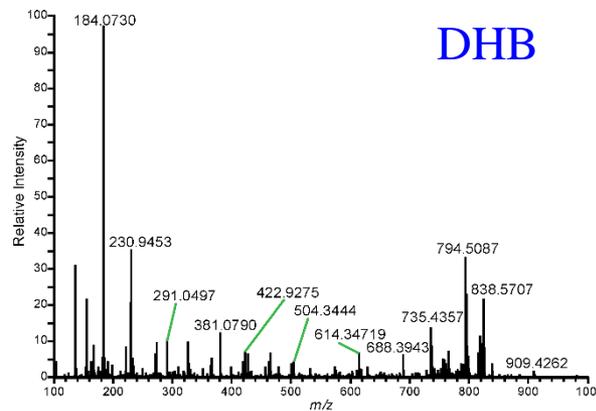
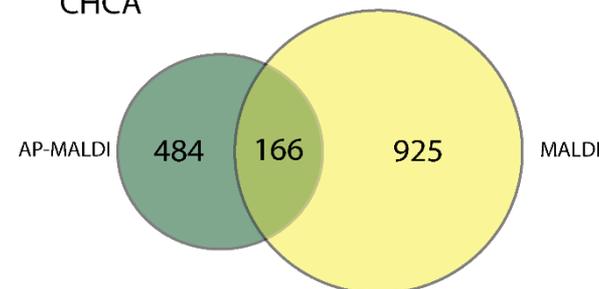
Number of m/z Detected Between MALDI and AP-MALDI Platforms



DHB

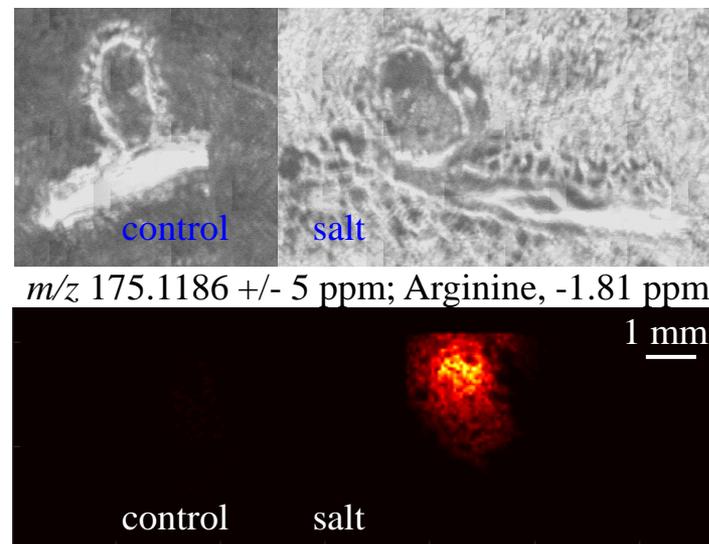


CHCA



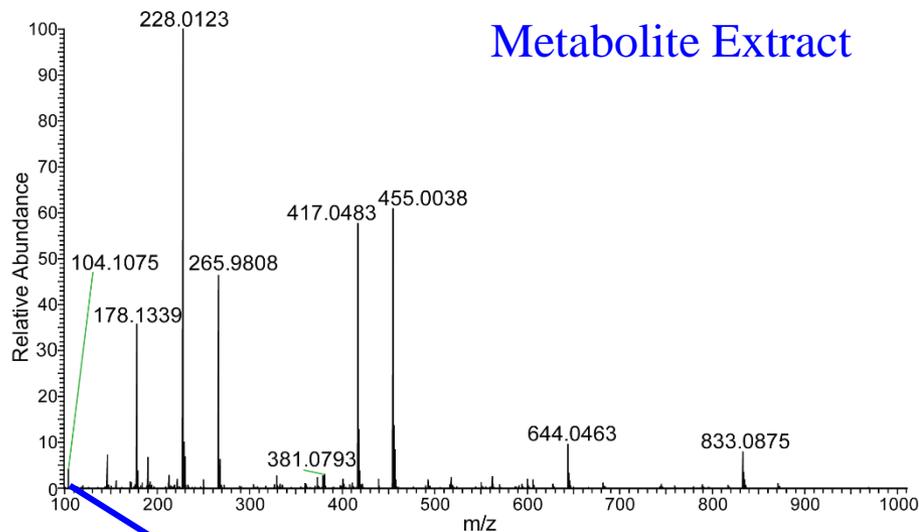
A High Resolution AP/MALDI-Q-Orbitrap Platform Enables *In Situ* Analysis of Metabolites involved in Salt Stress in *Medicago truncatula*

- Salt stress
 - Decreased plant growth
 - Poor development of symbiosis in root-nodule
 - Reduced nitrogen-fixation capacity
- Appearance of salt stress nodules: fewer, smaller, globular, white/brown



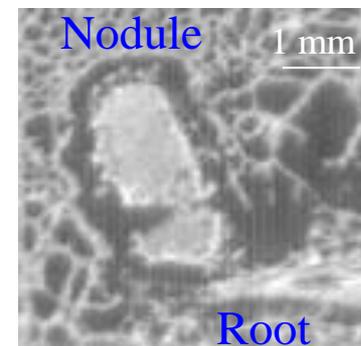
m/z; Retention time (min)	Distribution	AUC >0.75 Location	Identification; Adduct Identified	Literature Molecular Weight	Delta ppm
268.1034; 3.55	Control Nodule and Root	Root	Adenosine [M+H] ⁺	267.0968	-2.53
175.1186; 1.04	Salt Nodule	Nodule	Arginine [M+H] ⁺	174.1117	-1.81
965.5076; 20.46	Salt Root and Outer Nodule	Root	Soyasaponin I [M+Na] ⁺	942.5188	-0.44

A High Resolution Sub-AP/MALDI-Q-Orbitrap Platform Enables Identification and *In Situ* Analysis of Metabolites in *Medicago Truncatula*

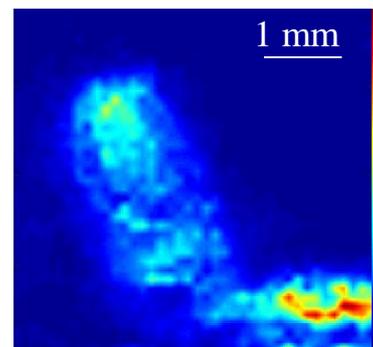


Imaging

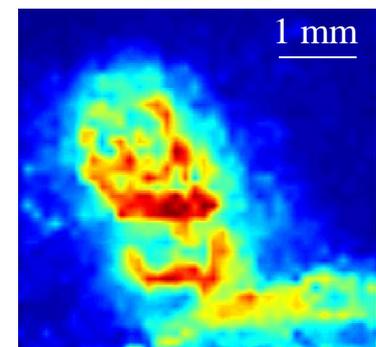
- 50 μm raster
- 16.91 minutes to acquire 2200 pixels



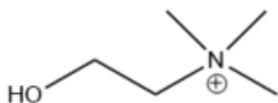
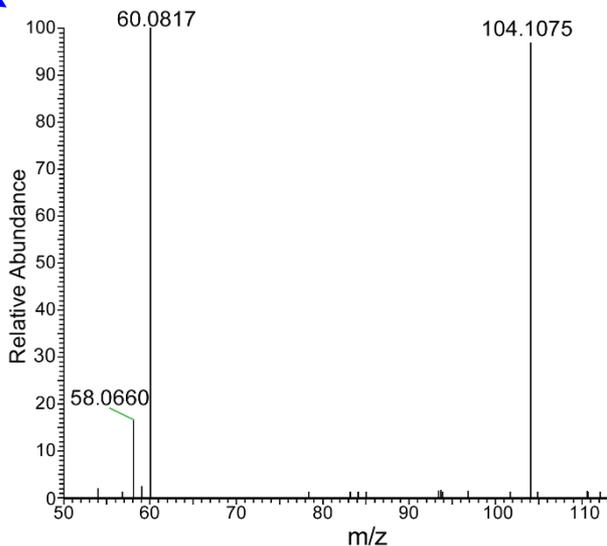
m/z 104.1077 \pm 5 ppm



m/z 455.0043 \pm 5 ppm

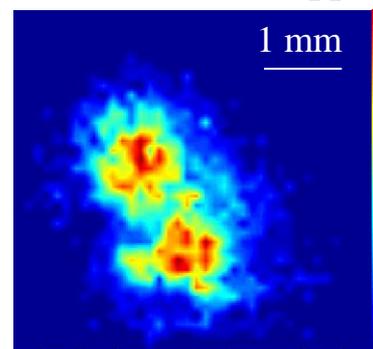


MS/MS
HCD

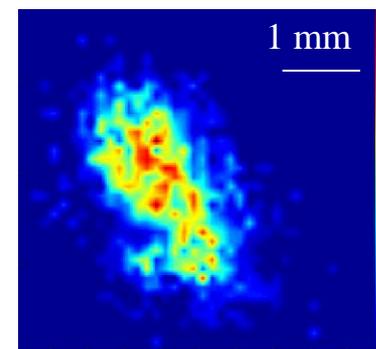


Choline

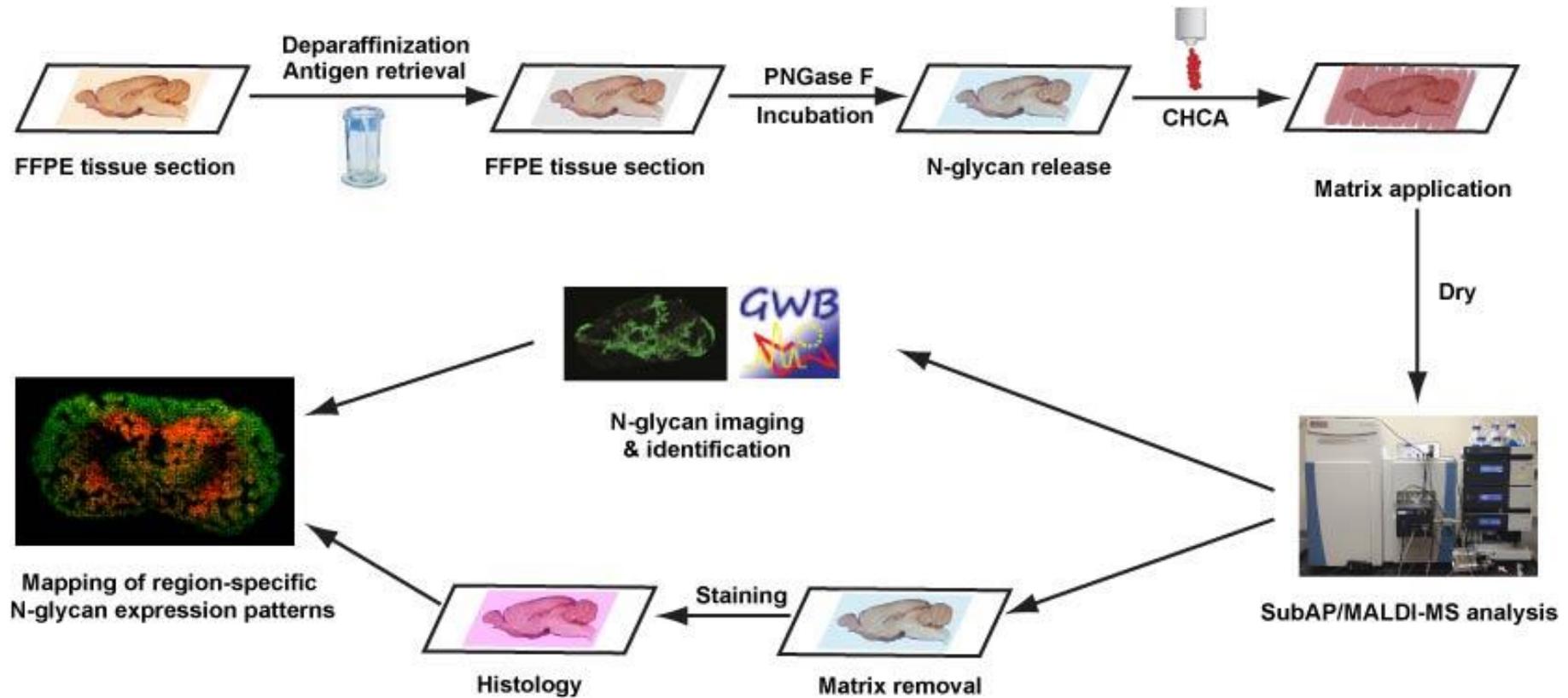
m/z 233.0525 \pm 5 ppm



m/z 241.0691 \pm 5 ppm

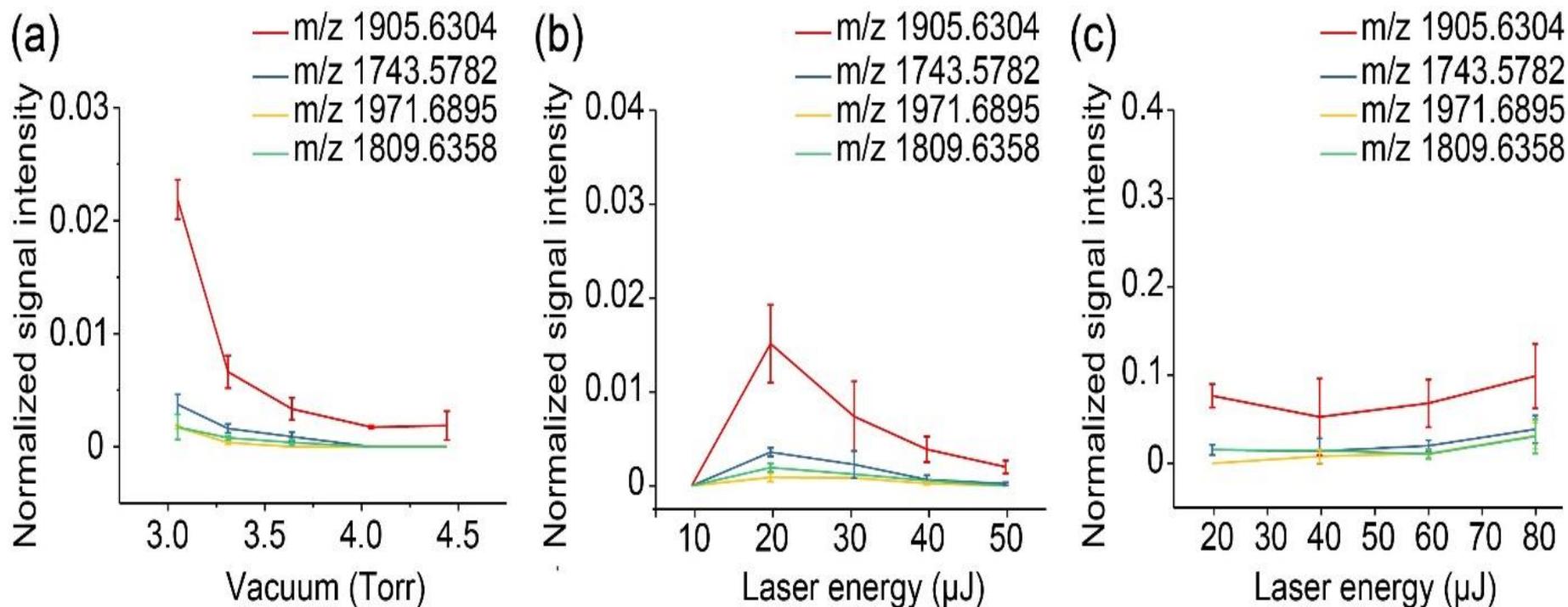


In Situ N-glycan Imaging of Mouse Brain with SubAP/MALDI-Q-Orbitrap Platform



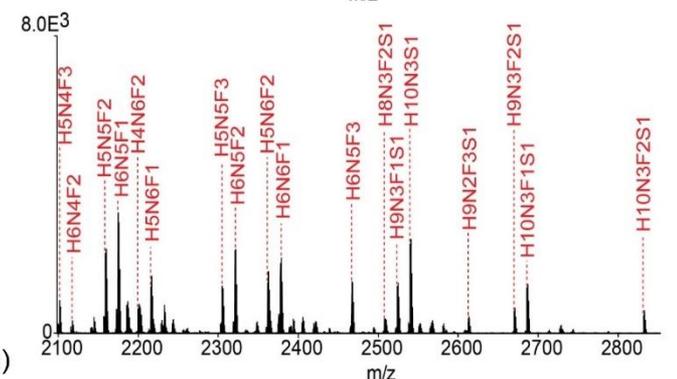
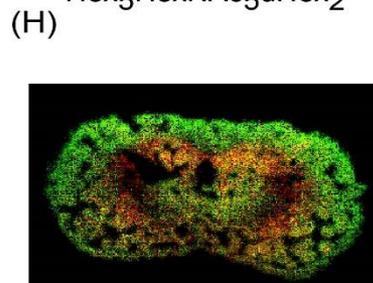
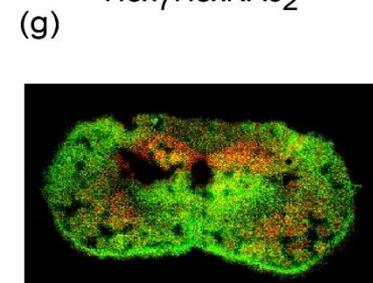
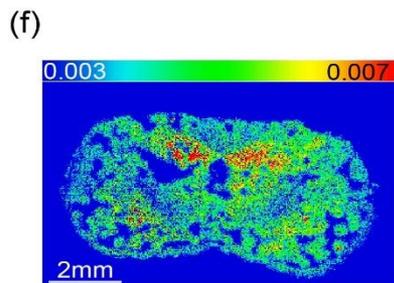
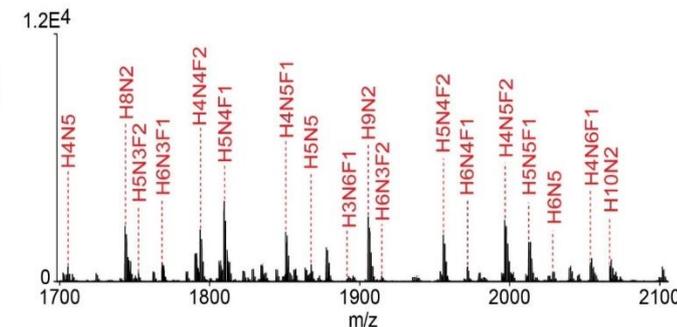
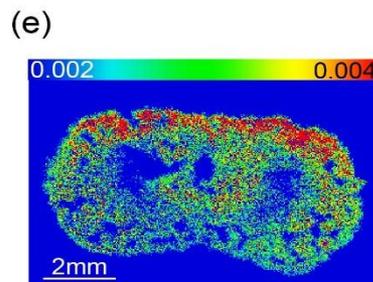
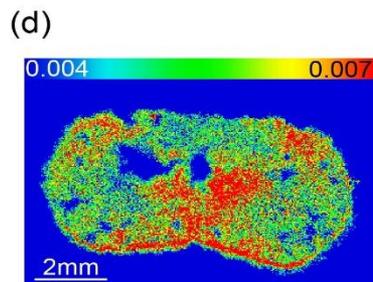
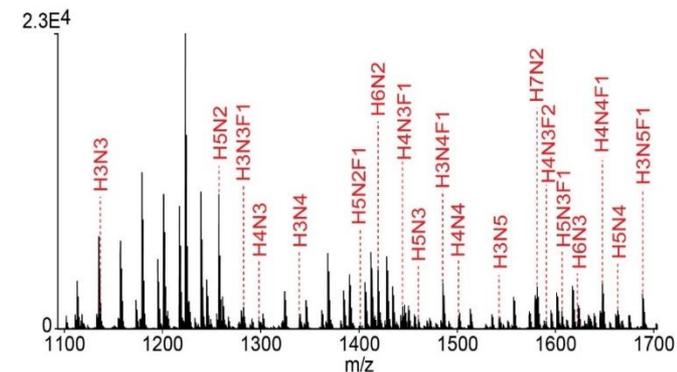
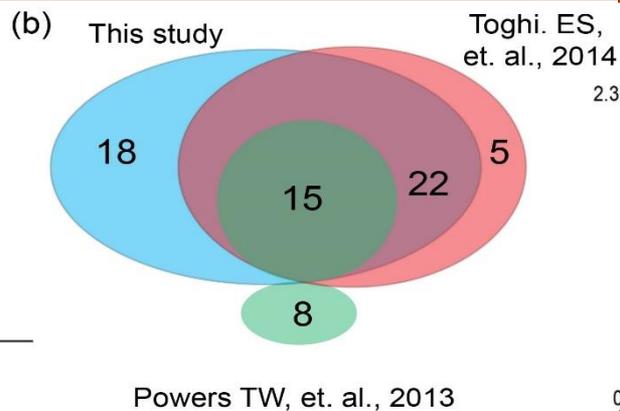
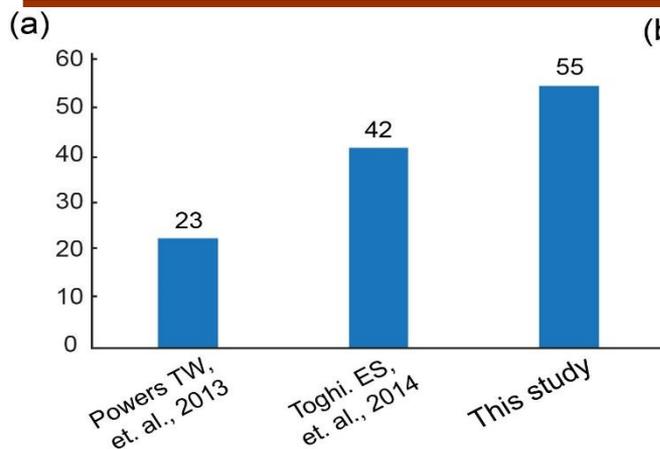
SubAP/MALDI imaging of N-glycans from formalin-fixed, paraffin-embedded (FFPE) tissue sections

Optimization of the SubAP/MALDI Source Parameters for N-linked Glycan Analysis



(a) TIC-normalized signal intensities of N-glycans significantly dropped following the decrease of source vacuum; (b) When CHCA used as matrix, laser energy of 20 μJ showed the highest TIC-normalized N-glycan signal intensities; (c) TIC-normalized N-glycan signal intensities were independent of laser energy when DHB was used.

In Situ N-glycan Imaging of Mouse Brain with SubAP/MALDI-Q-Orbitrap Platform

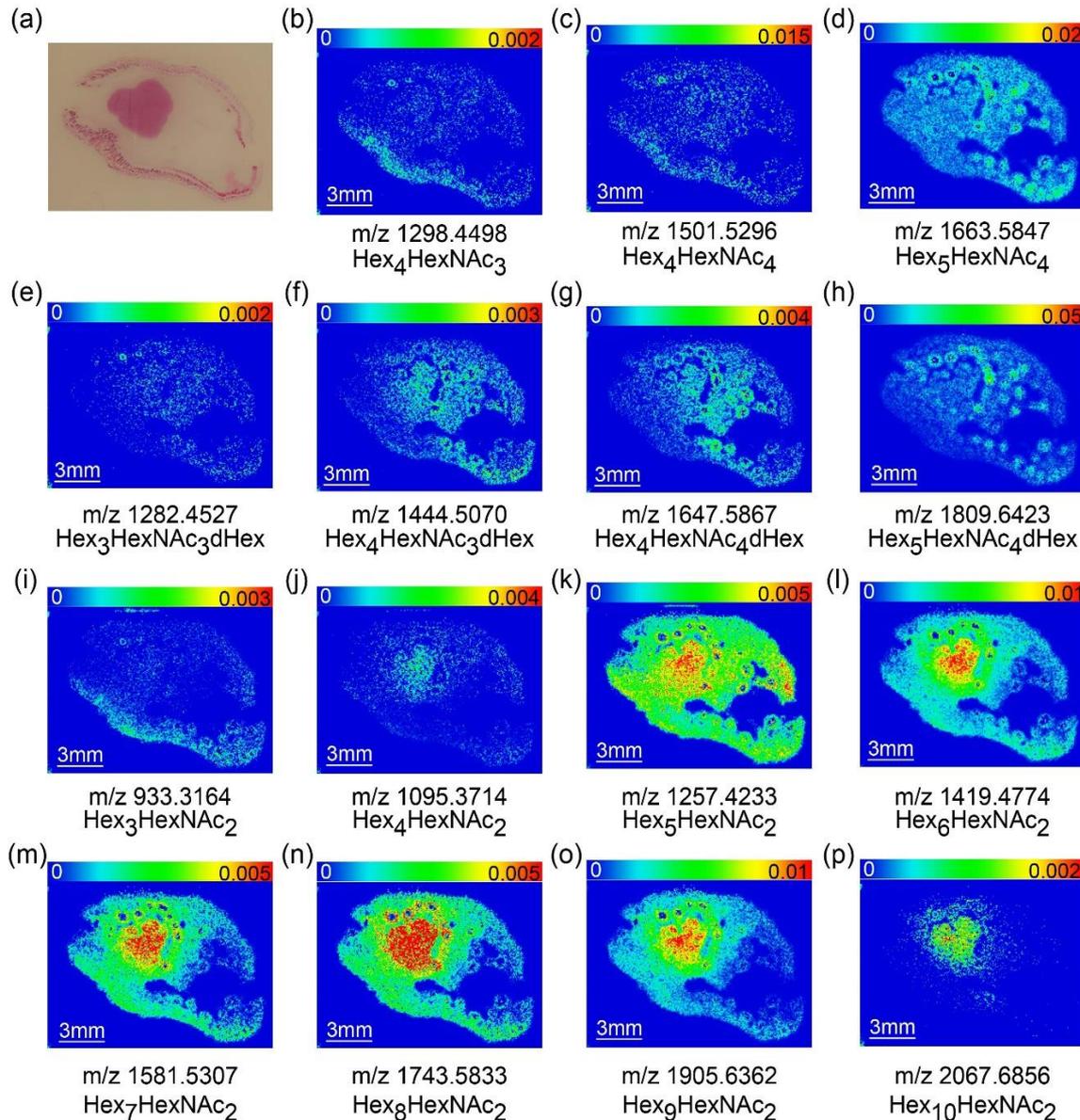


m/z 2174.7576
Hex₆HexNAc₅dHex₁

Hex₇HexNAc₂ (Green)
Hex₆HexNAc₅dHex₁ (Red)

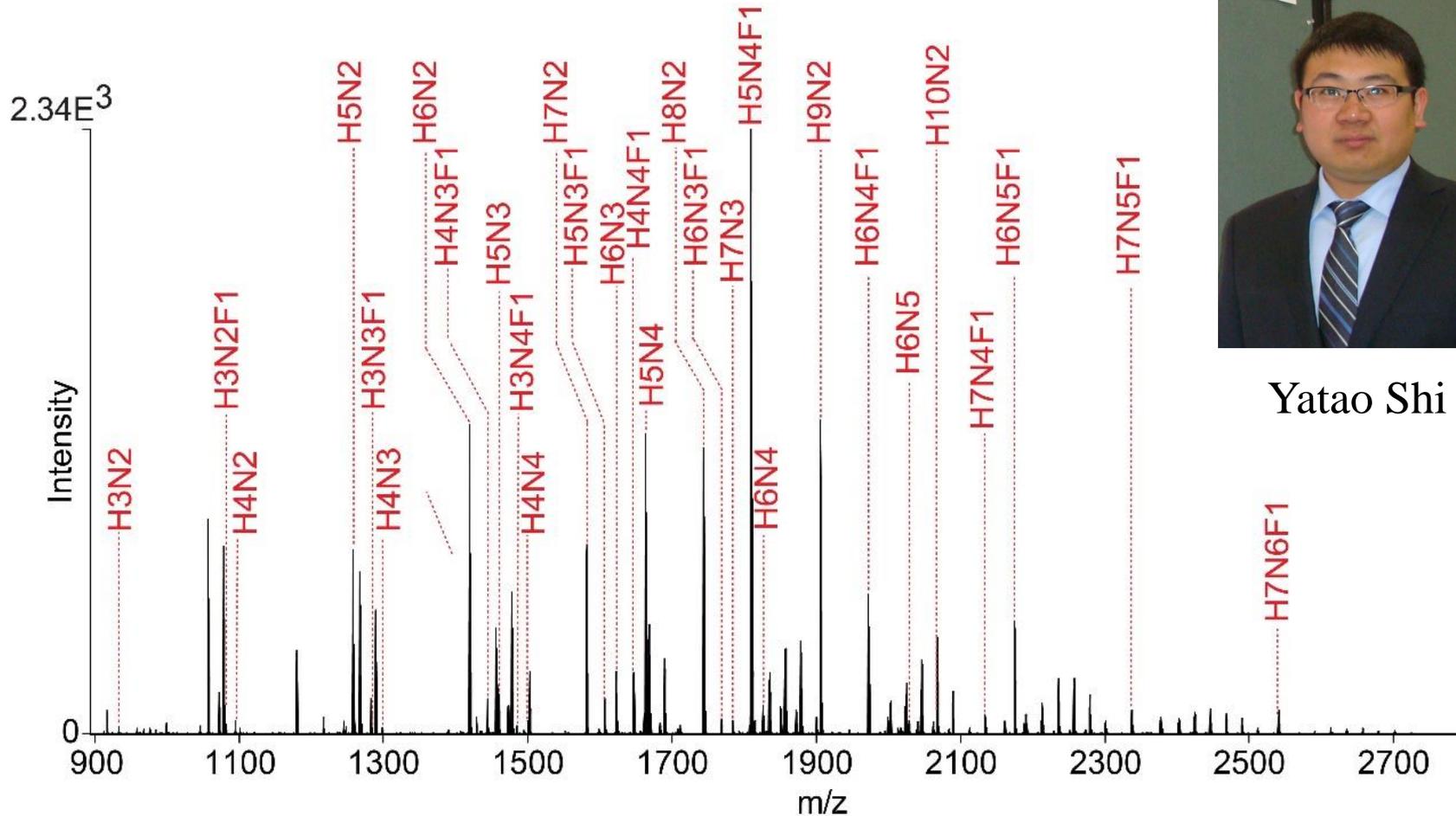
Hex₅HexNAc₅dHex₂ (Green)
Hex₆HexNAc₅dHex₁ (Red)

MSI of N-Glycan Analysis Reveals Distinct Distribution Patterns for Ovarian Cancer



Images of N-glycans showing different spatial distribution patterns on FFPE mouse tissue section with ovarian cancer. (a) H&E stained FFPE mouse tissue section with ovarian cancer. (b-h) Complex N-glycans showed similar distribution in cancer area in comparison to peripheral area; (i-p) High mannose N-glycans accumulated in cancer area except Hex₃HexNAc₂

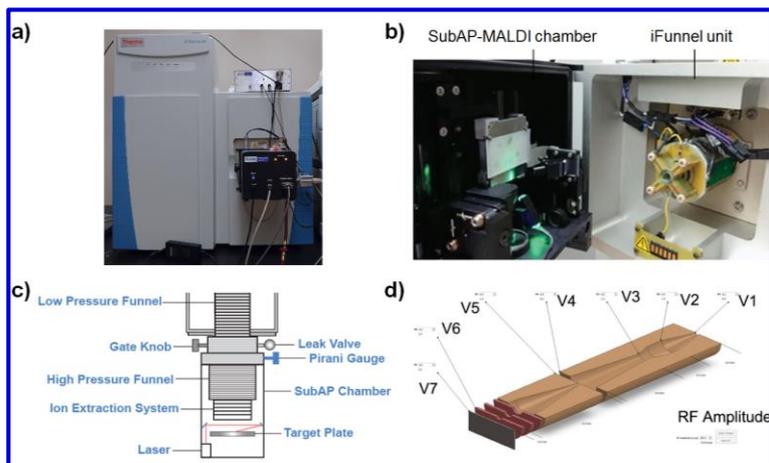
N-glycans detected from FFPE mouse tissue section with ovarian cancer



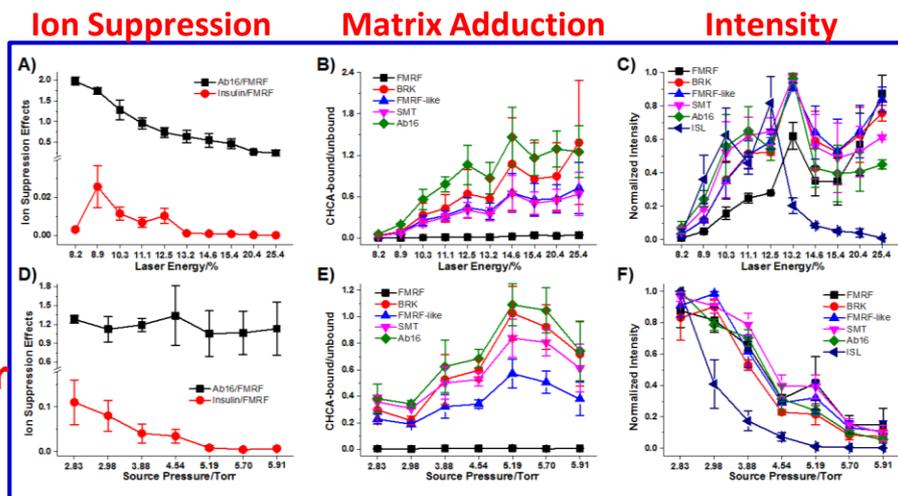
Yatao Shi

N-glycans detected from FFPE mouse tissue section with ovarian cancer. H: Hexose; N; N-Acetylglucosamine; F: Fucose; S: Sialic acid

AP-MALDI Sensitivity: SubAP + Ion Funnel + Optimization



Laser
Gas/Torr



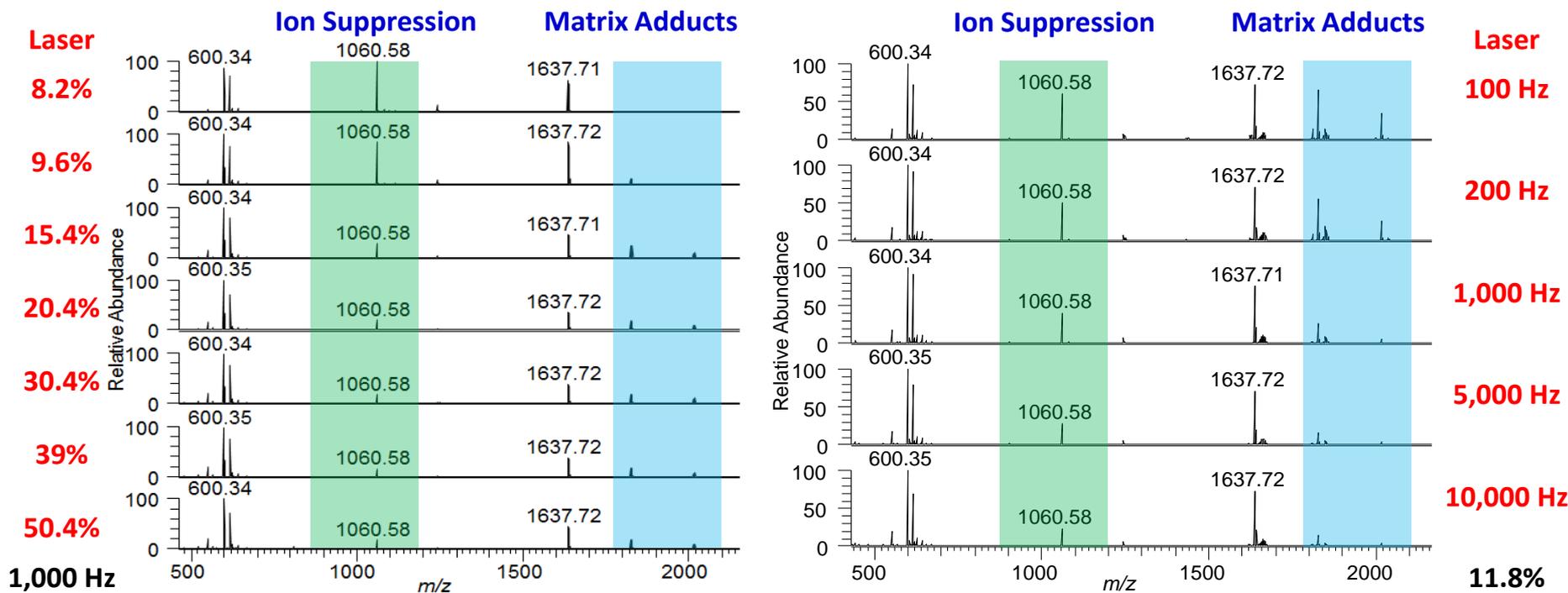
SubAP-MALDI Platform

Optimization

- ◆ SubAP-MALDI was employed as a model system for AP-MALDI sensitivity study, due to its feasibility in parameter optimization.
- ◆ The sensitivity/performance of SubAP-MALDI system is laser energy- and gas pressure-dependent.
- ◆ In addition, laser firing frequency and ion funnel voltages are other important parameters to optimize.

Li, G., Cao, Q., Liu, Y., DeLaney, K., Tian, Z., Moskovets, E., & Li, L. (2019). Characterizing and alleviating ion suppression effects in atmospheric pressure matrix-assisted laser desorption/ionization. *Rapid Communications in Mass Spectrometry*, 33(4), 327-335.

SubAP-MALDI: Ion Suppression + Matrix Adducts



- ◆ Neuropeptide mixtures were used to observe the frequency- and laser energy-dependency of ion suppression effects and matrix adduction effects.
- ◆ Upon increasing laser energy, both ion suppression effects and matrix adduction increase.
- ◆ Upon increasing laser frequency, ion suppression increases but matrix adduction decreases.
- ◆ Optimized laser: frequency of ~1,000 Hz, energy less than ~15% (CHCA).

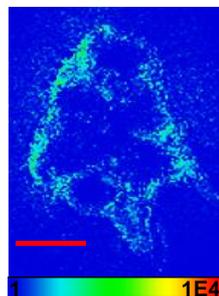
Li, G., Cao, Q., Liu, Y., DeLaney, K., Tian, Z., Moskovets, E., & Li, L. (2019). Characterizing and alleviating ion suppression effects in atmospheric pressure matrix-assisted laser desorption/ionization. *Rapid Communications in Mass Spectrometry*, 33(4), 327-335.

AP-MALDI imaging: reproducible lipid analysis of crab brain tissue section (#1)

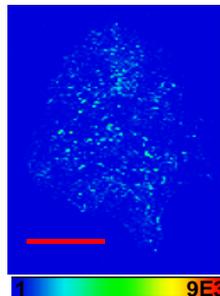
Optical Image



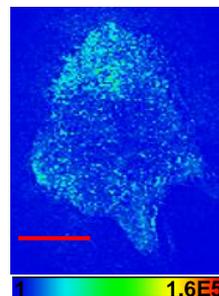
[d18:1 Cer/Na]⁺
m/z 822.6492



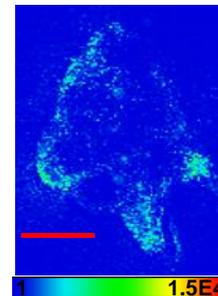
[LPC/Li]⁺
m/z 552.3670



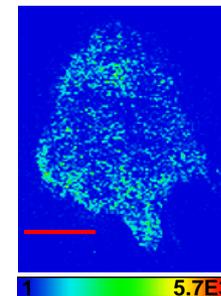
[PC(16:0-18:1)/H]⁺
m/z 760.5861



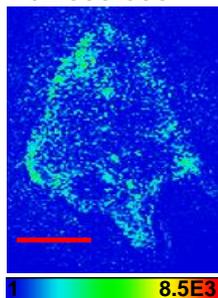
[MMPE+57+Li]⁺
m/z 796.6399



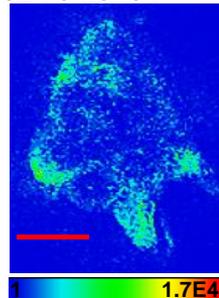
[SPE(23:2)/Na]⁺
m/z 777.5830



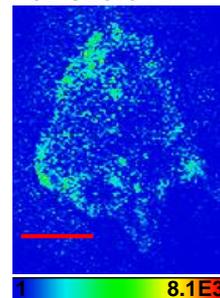
[TAG(51:8)/Na]⁺
m/z 855.6507



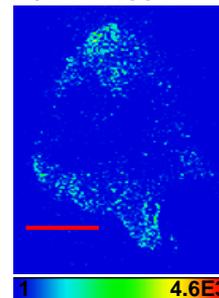
[DAG + C₄H₇ON/H]⁺
m/z 704.5794



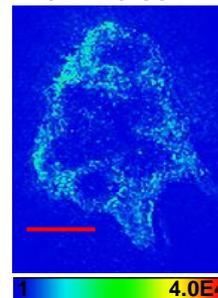
[DMPE+17+Li]⁺
m/z 825.6127



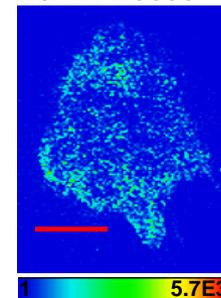
[FAHFA+BMA-ATP]⁺
m/z 717.5914



[FAHFA+AMPP]⁺
m/z 729.5914

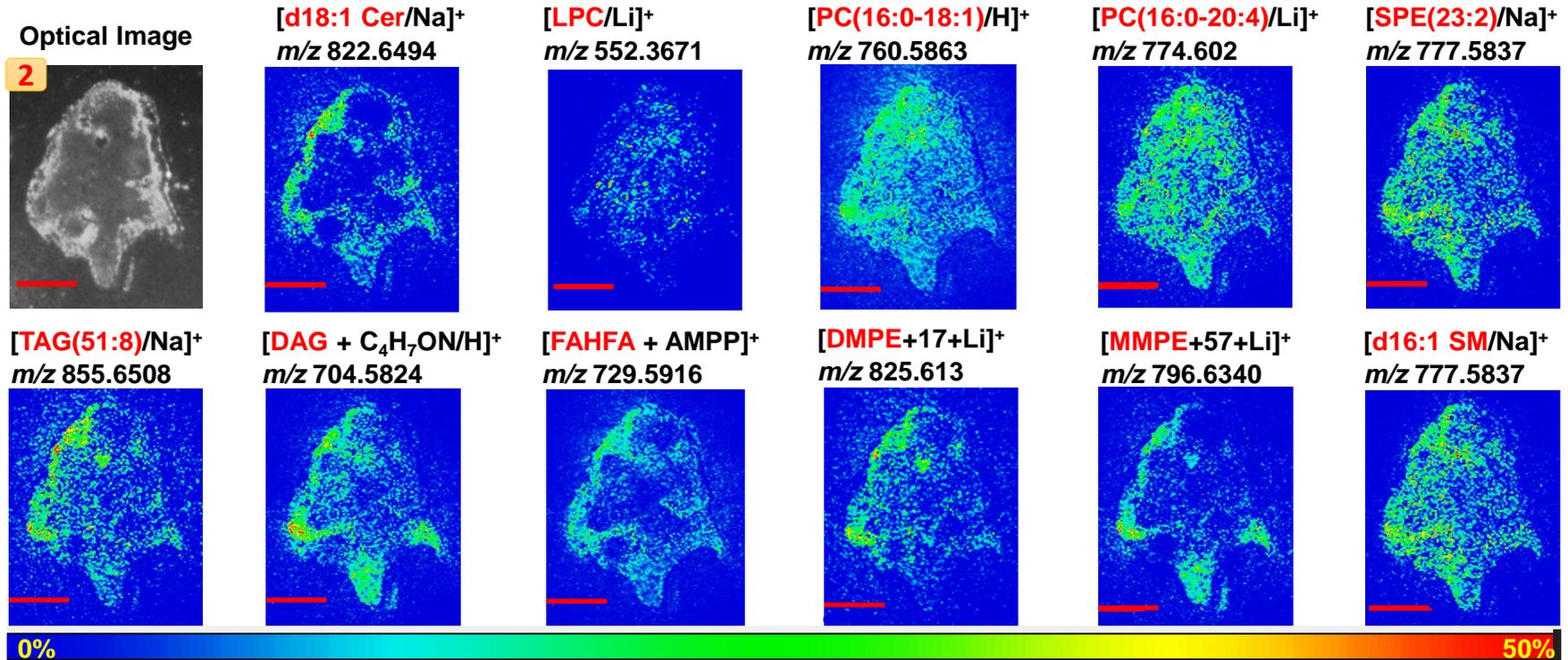


[d16:1 SM + Na]⁺
m/z 777.5830



Lipid AP-MALDI-MSI, all ions selected with 10 ppm tolerance. Scale bar, 1 mm. Step size: 30 μ m. Images are generated with the normalization to raw data in Thermo ImageQuest. In total, **68 lipids** are identified with accurate mass match, isotopic distribution, charge state comparison and S/N control (>3). **CHCA**, 20171206

AP-MALDI imaging: reproducible lipid analysis of crab brain tissue section (#2)



Lipid AP-MALDI-MSI, all ions selected with 10 ppm tolerance. Scale bar, 1 mm. Step size: 30 μ m. Images are generated with the normalization to raw data in Thermo ImageQuest. In total, **77 lipids** are identified with accurate mass match, isotopic distribution, charge state comparison and S/N control (>3). [CHCA](#), 20171206

AP-MALDI: reliable/reproducible lipid imaging of crab brain tissue

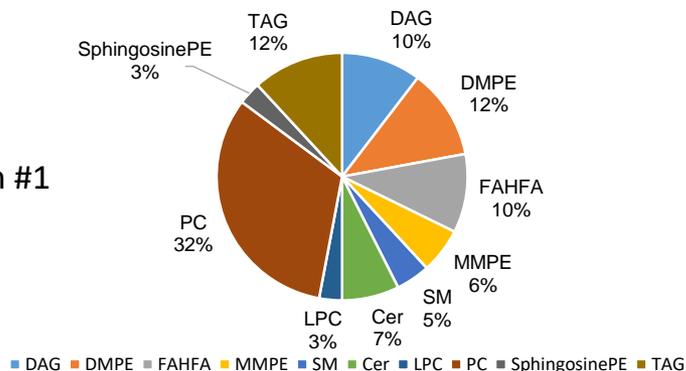


Table 1. Lipid AP-MALDI-MSI analysis of crab brain tissue.

Tissue Section #	DAG	DMPE	FAHFA	MMPE	SM	Cer	LPC	PC	SphingosinePE	TAG	SUM
S1	7	8	7	4	3	5	2	22	2	8	68
S2	9	7	8	8	4	4	3	26	2	6	77
in total	9	9	8	8	4	6	3	29	2	8	86

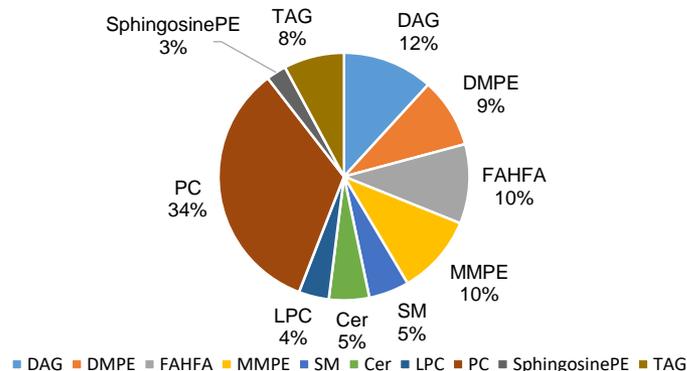
Lipid feature distribution of crab brain via AP-MALDI-MSI

Section #1



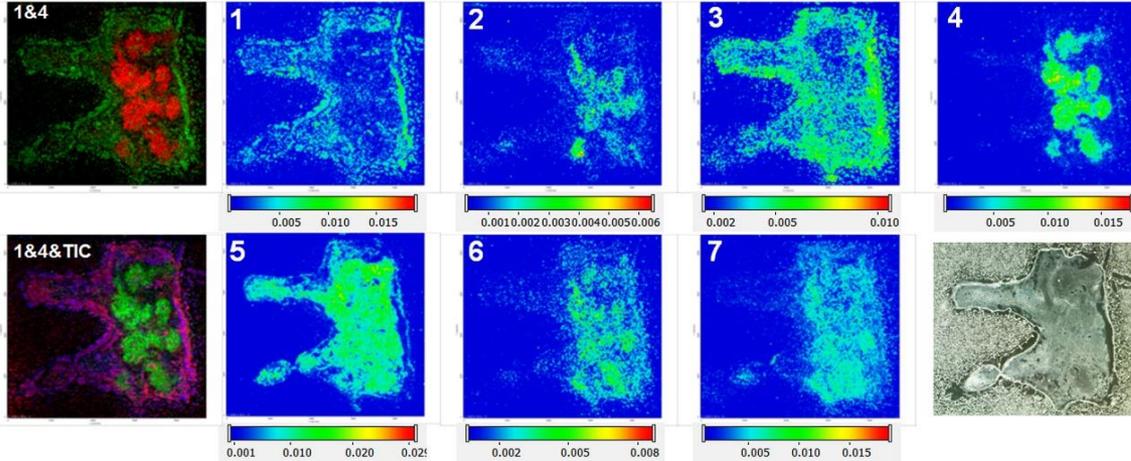
Lipid feature distribution of crab brain via AP-MALDI-MSI

Section #2



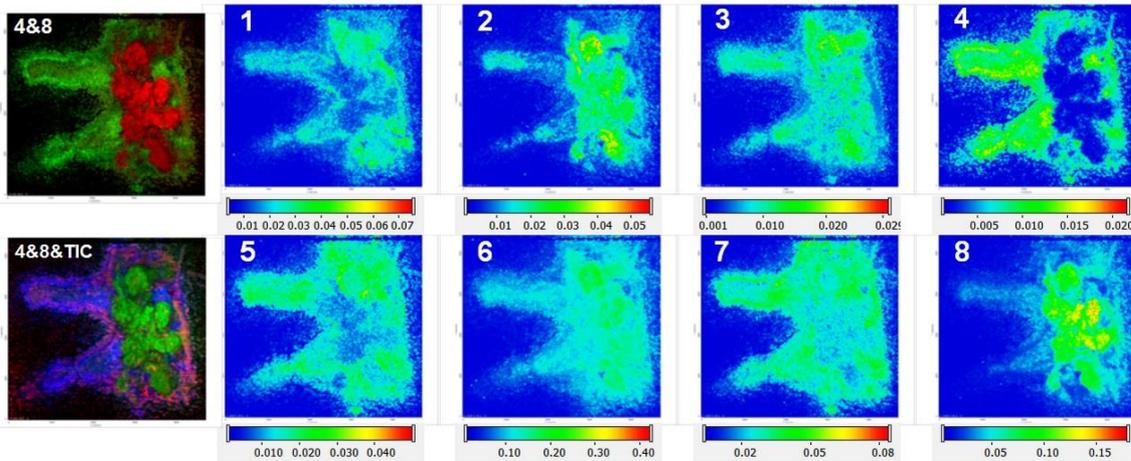
◆ Both slides share similar lipid features and imaging patterns.

AP-MALDI: Multidimensional, High-resolution Visualization



LPC

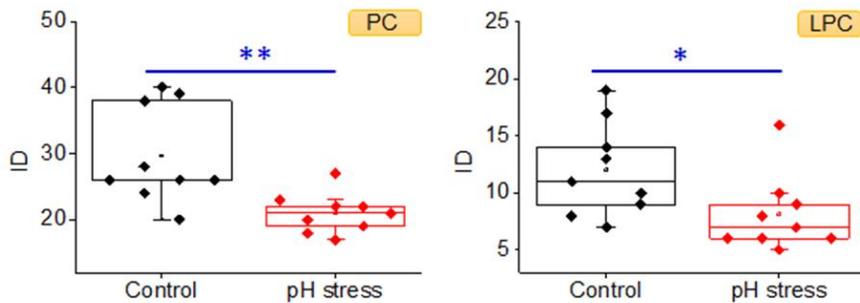
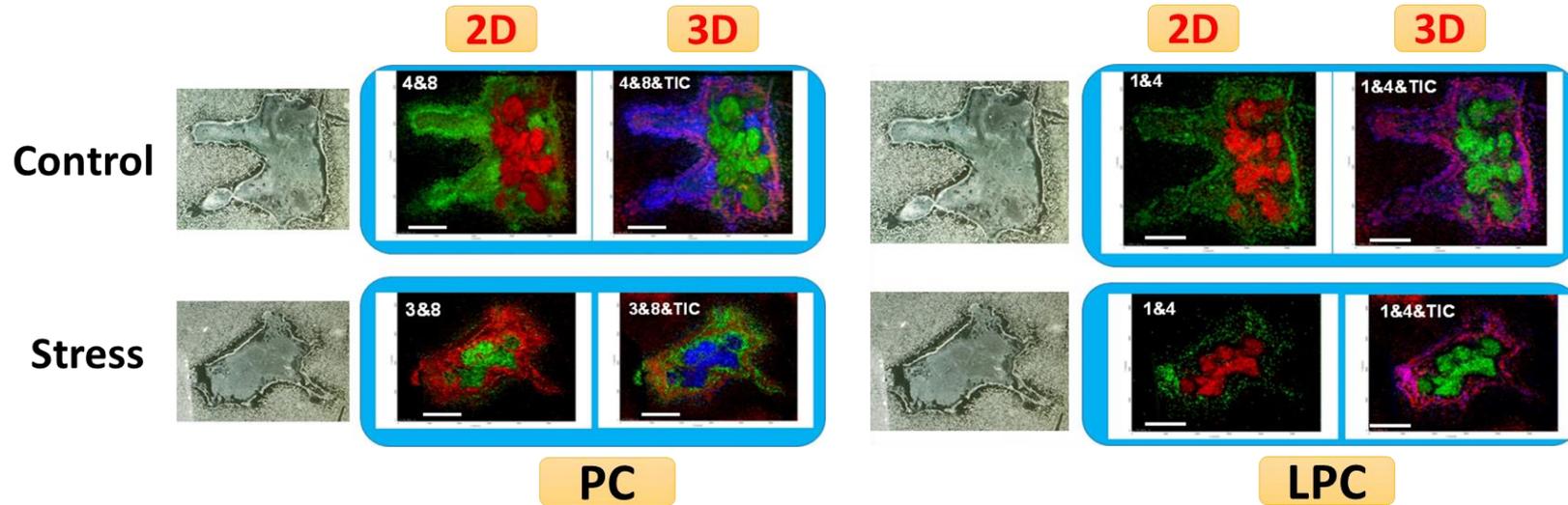
	Species	lipid features	calculated m/z	detected m/z	Delta m/z (ppm)	charge	S/N
1	LPC/Na+	A16:0	504.3424	504.3445	4.07	1	104.94
2	LPC/Li+	LPC18:3	524.3323	524.3348	4.79	1	34.08
3	LPC/Li+	X20:4	550.3479	550.3503	4.29	1	118.43
4	LPC/Li+	X20:3	552.3636	552.3660	4.36	1	86.49
5	LPC/Na+	P20:0	558.3894	558.3941	8.42	1	145.00
6	LPC/Li+	X22:4	578.3792	578.3815	3.91	1	42.41
7	LPC/Li+	X22:3	580.3949	580.3971	3.81	1	80.63



PC

	Species	lipid features	calculated m/z	detected m/z	Delta m/z (ppm)	charge	S/N
1	PC/H+	D16:1-16:0/D14:1-18:0	732.5538	732.5540	0.30	1	402.79
2	PC/H+	D16:0-16:0	734.5694	734.5686	1.13	1	281.44
3	PC/Li+	P16:0-18:1/A16:0-18:2	746.5670	746.5691	2.76	1	131.22
4	PC/H+	8:0/P18:0-16:0/A18:1-16:0/A16:0-18:2	746.6058	746.6060	0.24	1	145.54
5	PC/H+	D16:0-18:2	758.5694	758.5694	0.04	1	239.56
6	PC/H+	D16:0-18:1	760.5851	760.5846	0.63	1	2241.00
7	PC/H+	D18:0-18:2/D18:1-18:1	786.6007	786.6006	0.17	1	471.26
8	PC/H+	D18:0-18:1	788.6164	788.6158	0.74	1	1138.95

AP-MALDI: Multidimensional Visualization (Control vs. pH Stress)



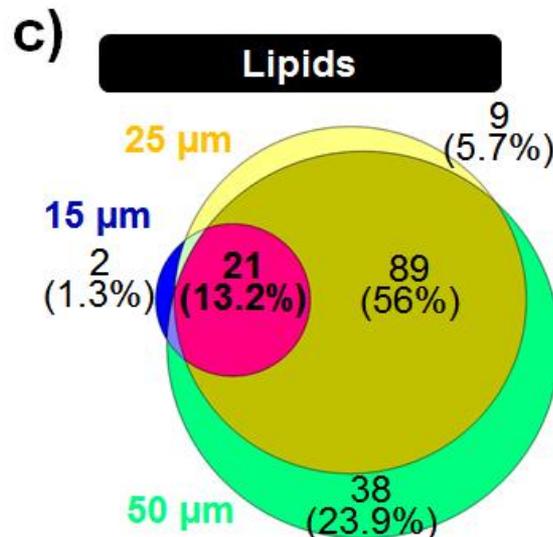
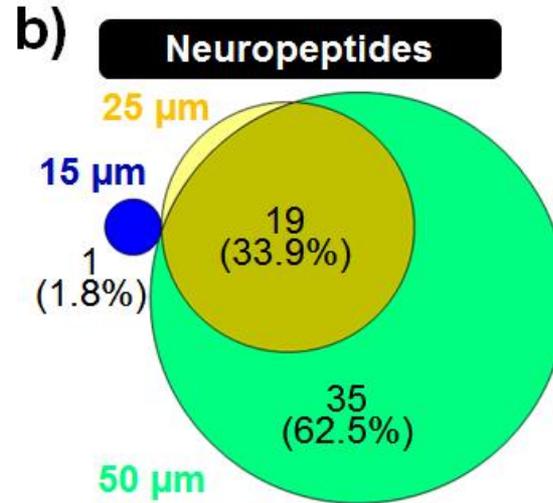
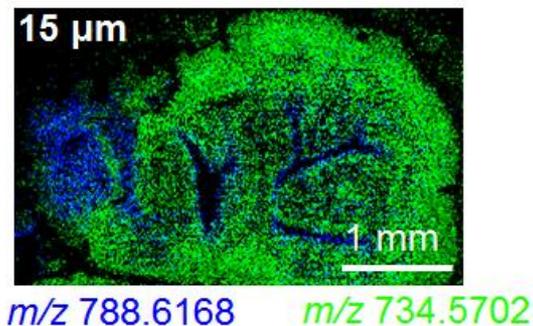
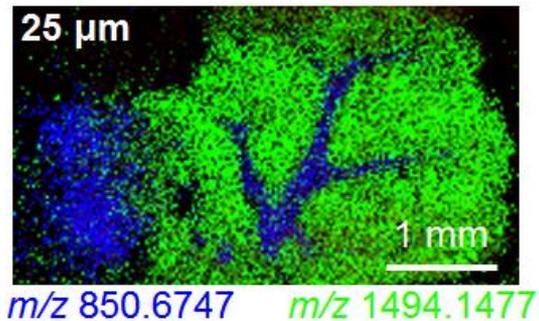
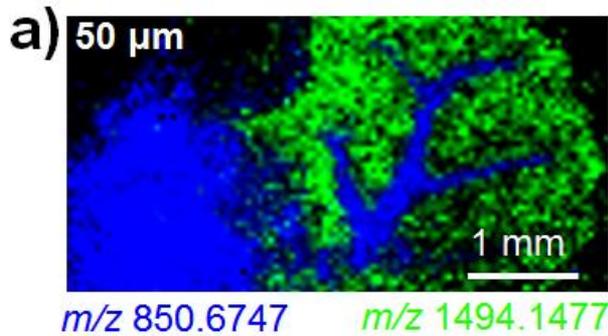
Multidimensional AP-MALDI-MSI for high-resolution microstructure visualization. Scale bar, 1000 μm . Crab brain. AP-MALDI, 30 μm step size. 2D, reconstruction of brain model by overlay of two lipids: like lipid 4 & lipid 8. 3D, reconstruction of brain model by overlay of three ions, two lipids + TIC.

- ◆ Lipids are down-regulated in response to pH stress.
- ◆ Neurons, just as imaged, shrink in response to pH stress.

Statistical comparisons are performed using student's *t*-tests; * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, n.s., not significant. All error bars denote SD; $n = 9$.

High-resolution SubAP-MALDI-MSI

Spatial Resolution & Sensitivity



Mouse brain cerebellum tissue section MSI:

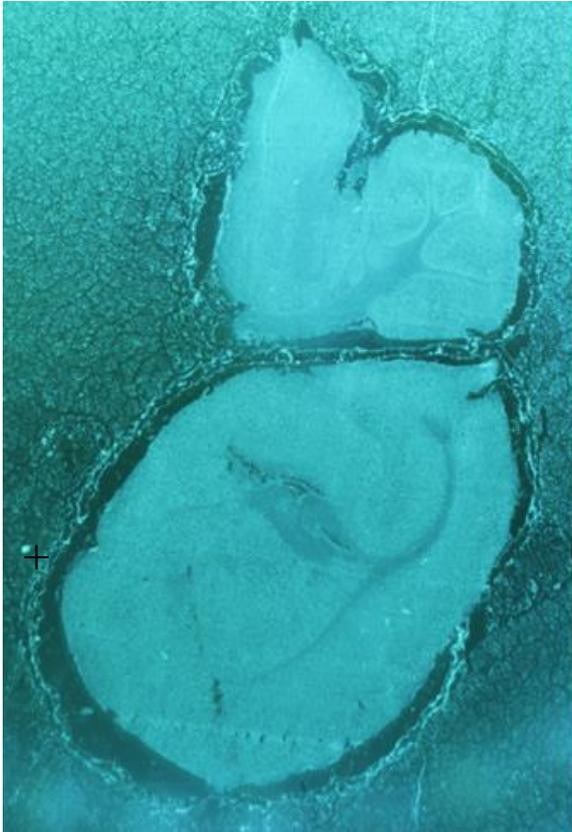
- a) Step size: 15/25/50 μm .
- b) Neuropeptide distribution.
- c) Lipid distribution.

Higher spatial resolution,
Lower sensitivity.

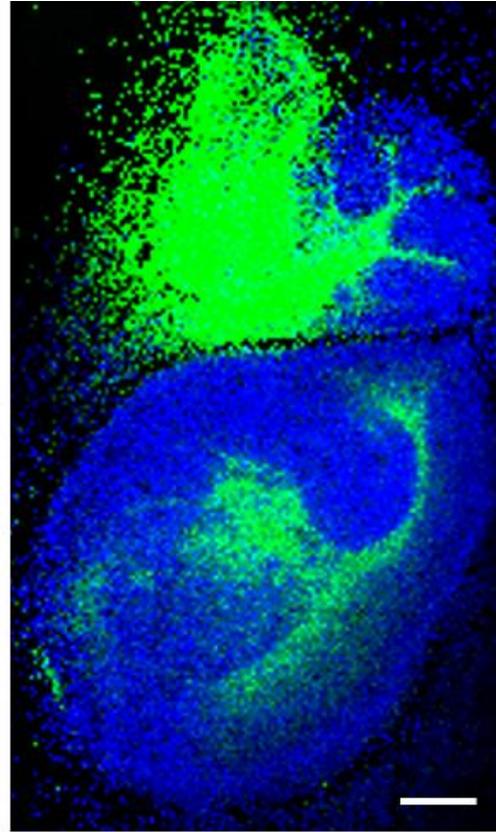
Optimal: 25 μm .
(lipids + neuropeptides)

High-resolution SubAP-MALDI-MSI

Mass Resolution & Sensitivity



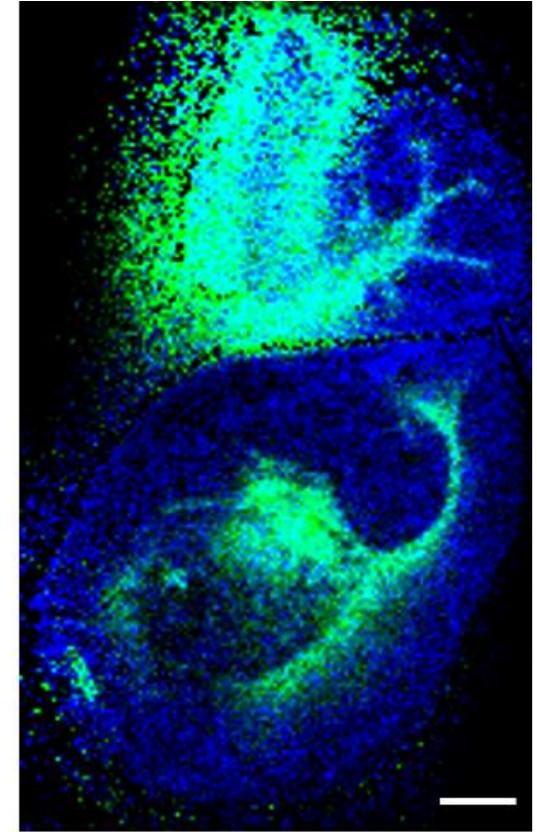
Optical image



TAG(52:9)/Na+
Lactosyle Ceramide(16:0)/Li+
 m/z 867.6530 vs 867.6239



High mass resolution

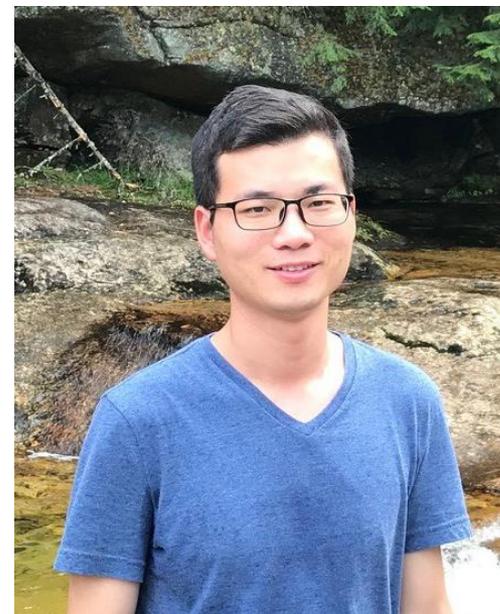
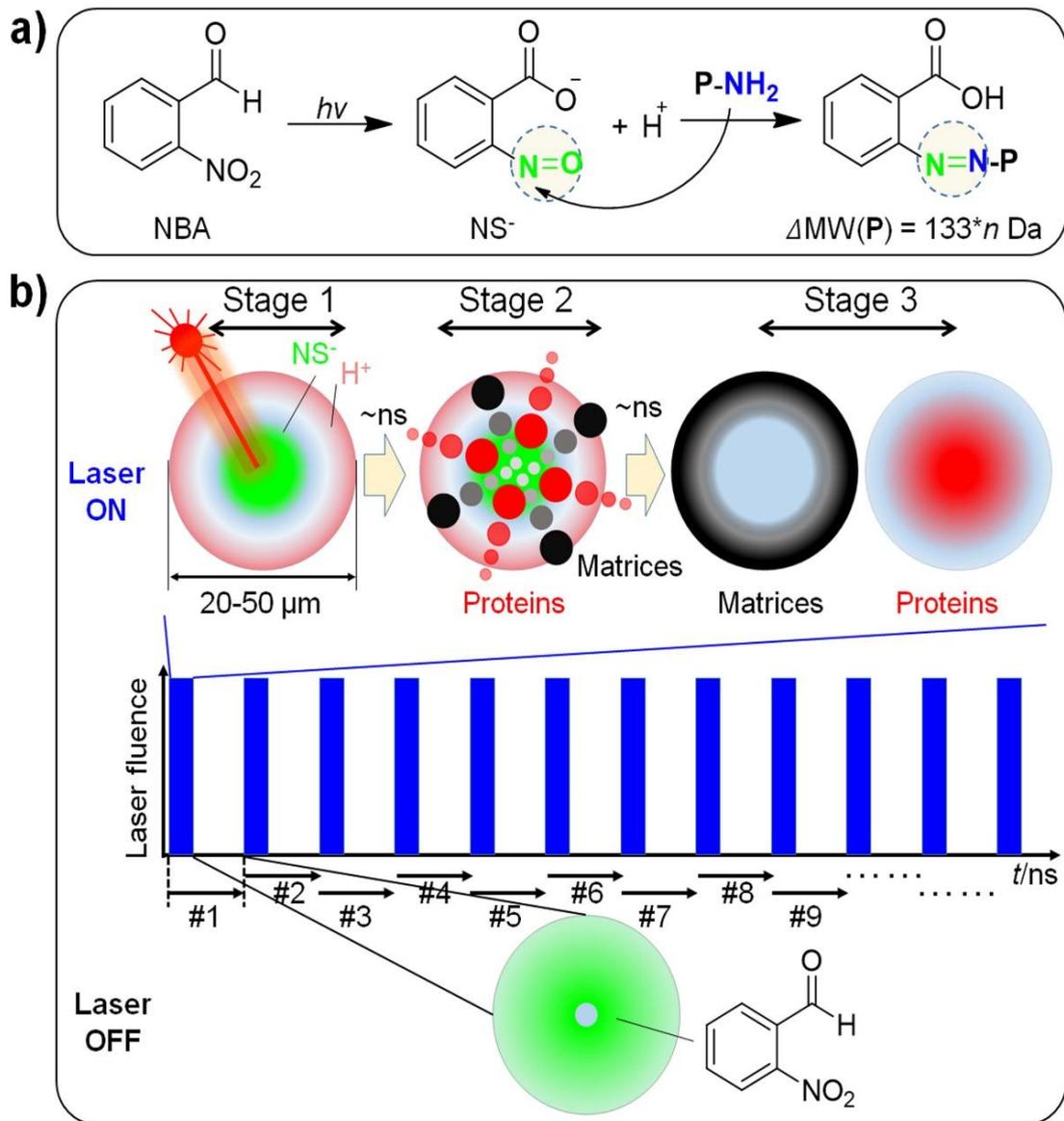


LFDDFLRFamide/NH4+
[MMPE(38:4)+70+Li]+
 m/z 1088.59 vs 822.647



neuropeptides+lipids

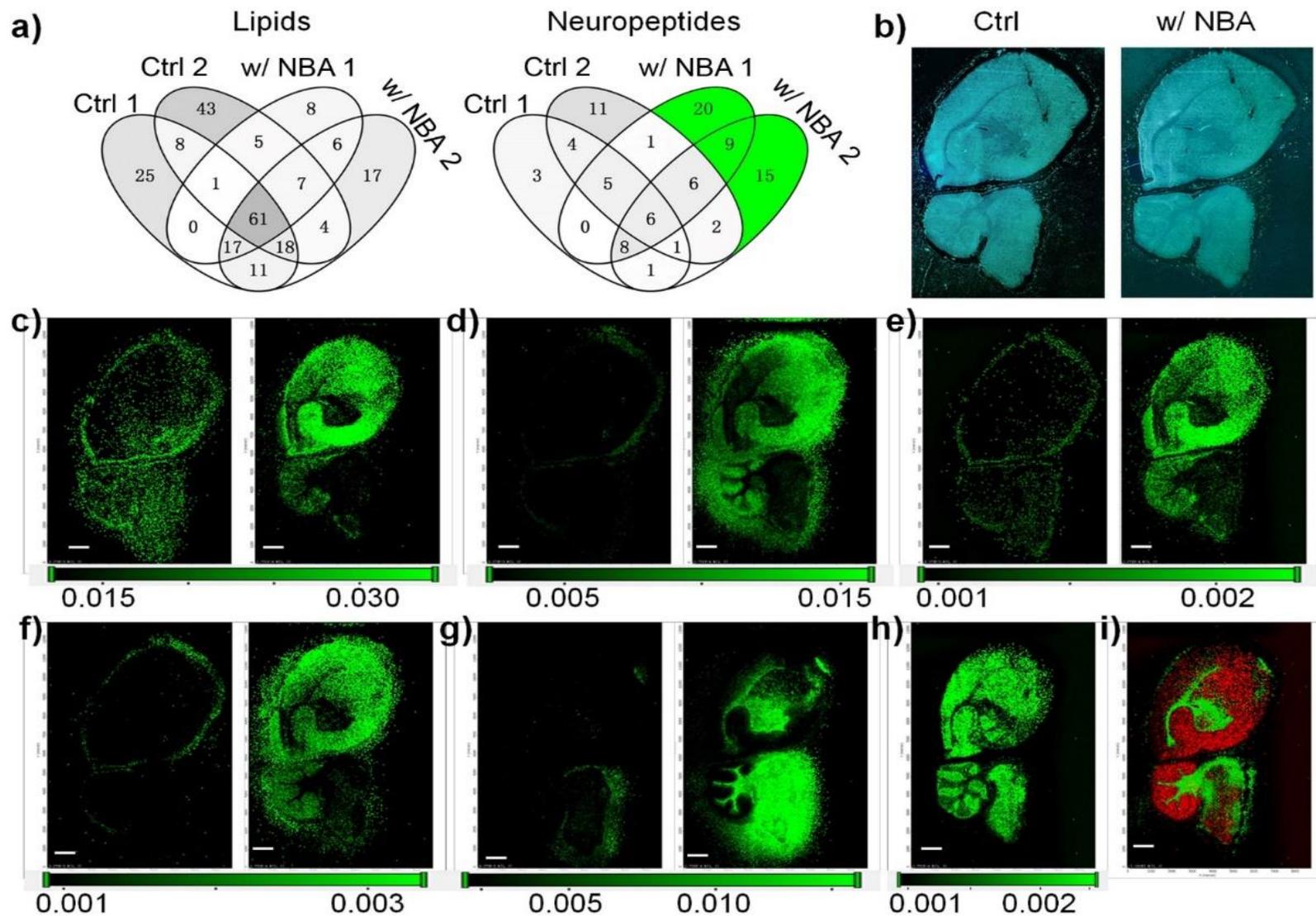
Nanosecond Photochemical Reaction (nsPCR) Enables On-demand Matrix Removal



Dr. Gongyu Li

a) Nanosecond photochemistry on NBA, creating reactive NS⁻ for localized micro-electric field.
b) On-demand three-stage matrix removal regulated by laser ON/OFF switch.

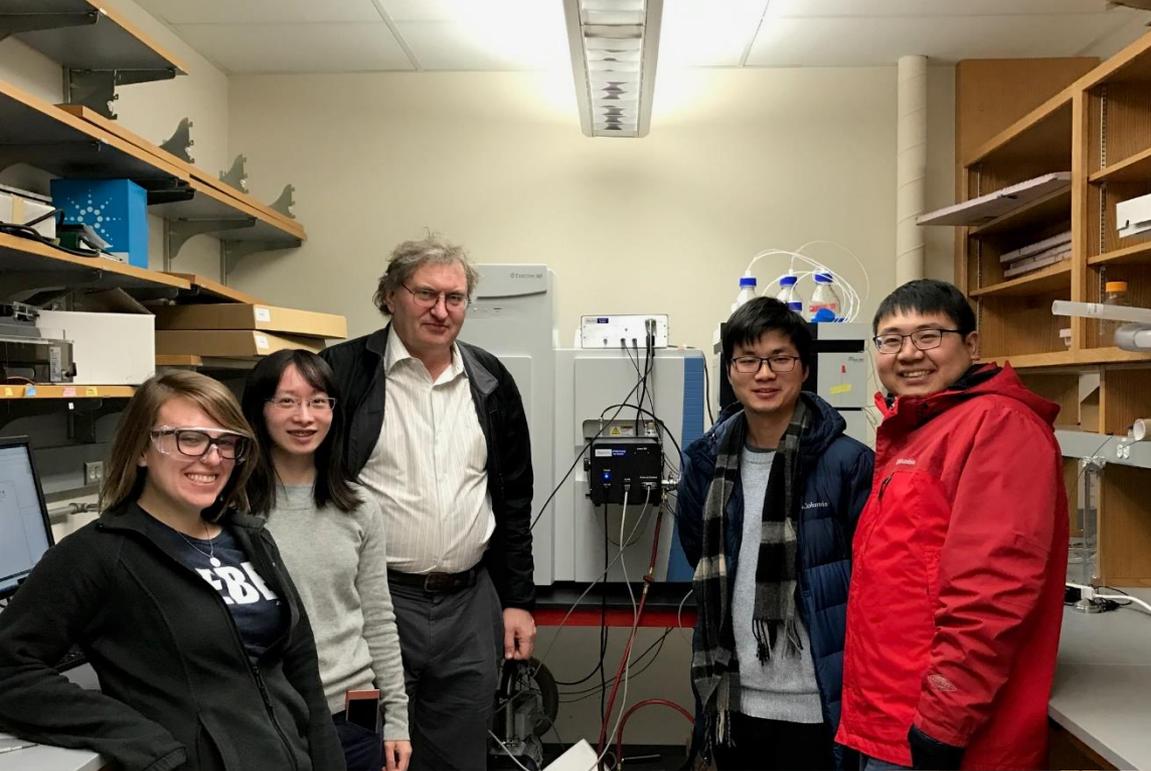
Enhanced Neuropeptide Identification and Visualization via nsPCR



Take Home Messages...

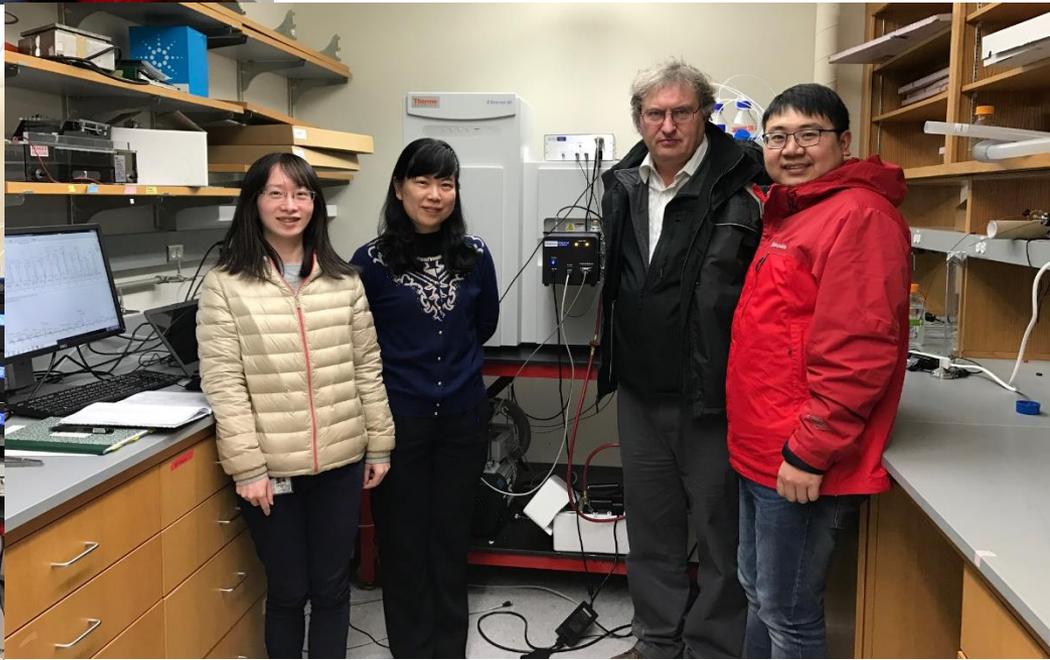
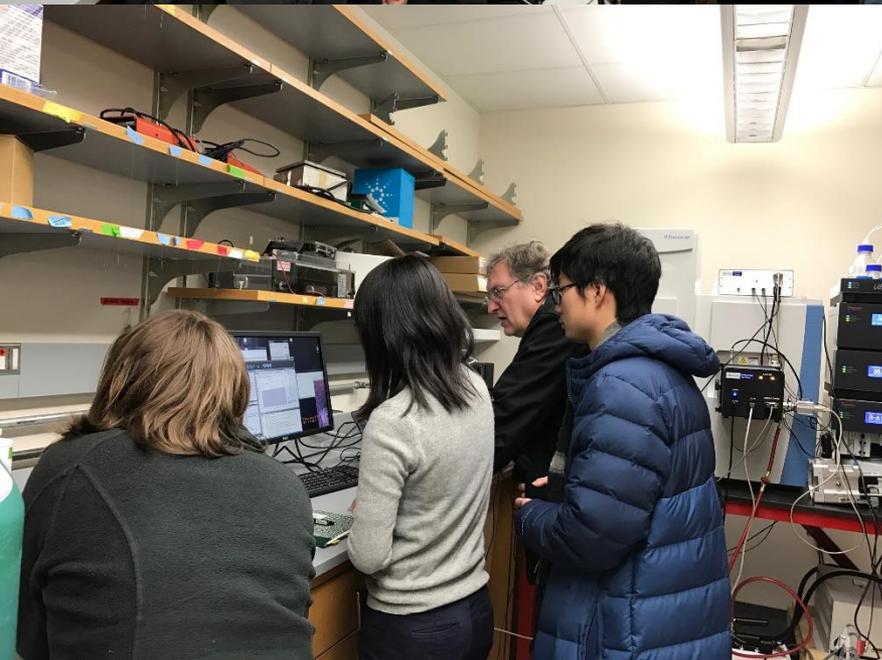


- ✿ LSI/MAIV-MS combines the benefit of MALDI and ESI to achieve expanded mass range, improved MS/MS efficiency, MS imaging capability and labile PTM protein analysis.
- ✿ Novel combination of multiplexed MSI with DDA on a MALDI Orbitrap platform enables enhanced *in situ* neuropeptide identification.
- ✿ AP/MALDI-Q-Orbitrap platform enables high resolution MSI in mass and space by multi-mode ionization and acquisition
- ✿ SubAP/MALDI-Q-Orbitrap platform provides promising solution to combine high spatial and mass spectral resolution with sensitivity for MSI of diverse biomolecules



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The Lingjun Li Research Group at UW-Madison

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<i>Dr. Xuefei Zhong</i>	<i>Dr. Chris Lietz</i>	Dr. Xudong Shi (UW-Madison)

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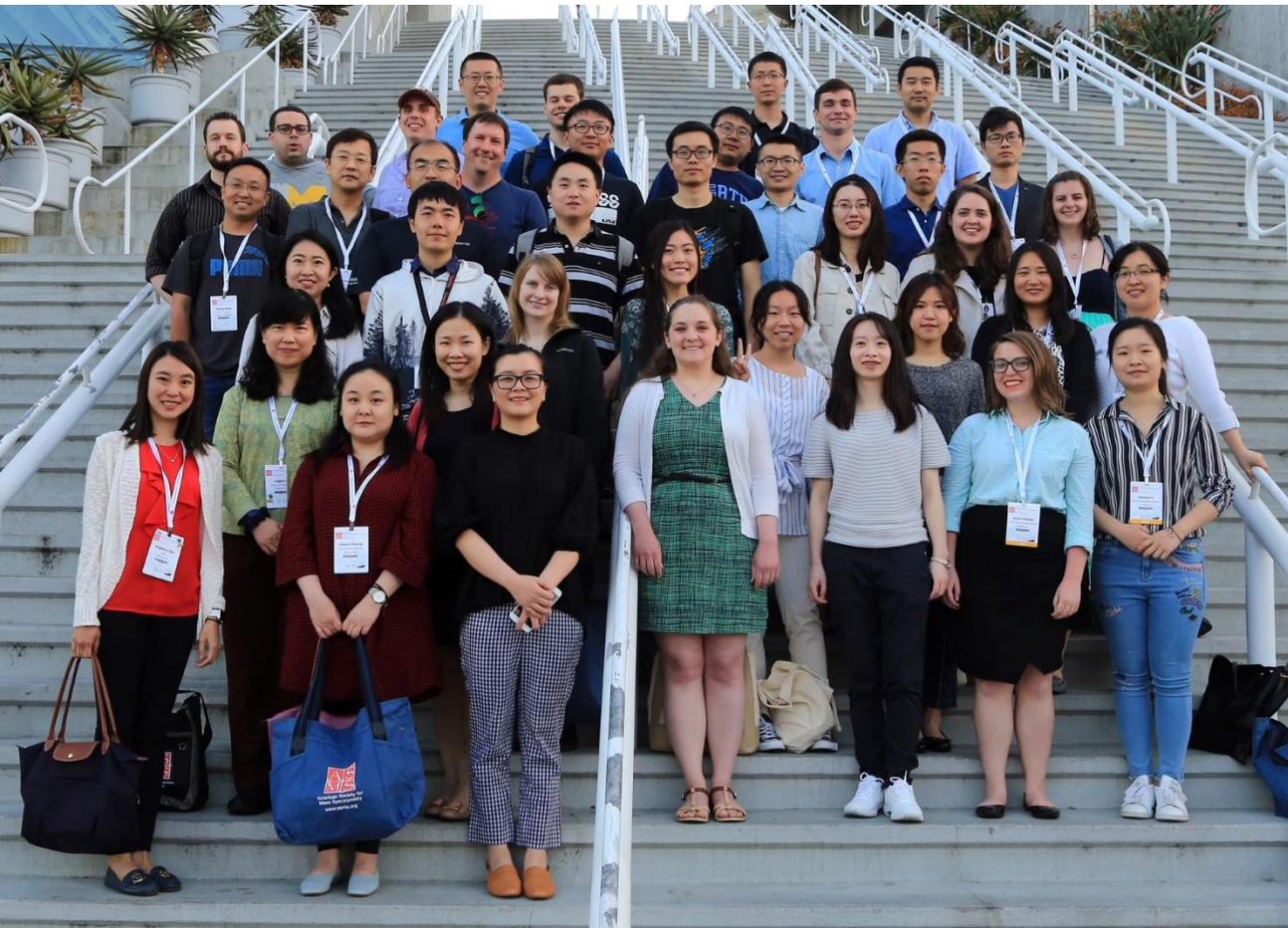
UW Graduate School, School of Pharmacy, WARF
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The Lingjun Li Research Group



THE UNIVERSITY
of
WISCONSIN
MADISON

The Lab started in 2003; Successfully trained and graduated 43 Ph.D. students; Currently training 24 grad students; 5 postdocs; 6 undergrads; 1 HS





Organizers

Julia Laskin

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Lingjun Li

*University of
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Jeffrey Spraggins

Vanderbilt University

35TH ASILOMAR CONFERENCE

Mass Spectrometry Imaging: New Developments and Applications

October 11 – 15, 2019

Asilomar Conference Center • Pacific Grove, CA





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IMPORTANT DEADLINES

August 9

Travel stipend application for
students and post-docs

September 6

Abstract submissions for
contributed posters and hot
topic talks

September 10

Room reservations
at Asilomar

September 13

Conference Registration



35th Asilomar Conference on Mass Spectrometry

Mass Spectrometry Imaging New Developments and Applications

October 11 - 15, 2019

ASILOMAR CONFERENCE CENTER, PACIFIC GROVE, CALIFORNIA

Mass spectrometry imaging is a vibrant, rapidly developing area of research that attracts researchers from numerous fields including instrumentation development, drug discovery, biotechnology, clinical research, forensics, and beyond. The 2019 ASMS Asilomar conference on mass spectrometry imaging is timely as there will be discussion on the progress made over the last decade. This conference will bring together researchers from multiple disciplines to discuss key challenges, innovative developments, and emerging applications in mass spectrometry imaging.

Detailed program at www.asms.org/asilomar-conference/program.

INVITED SPEAKERS

The program concludes with an after-dinner talk by **Richard Caprioli** (Vanderbilt University).

Nathalie Agar (Brigham and Women's
Hospital, Harvard Medical School)

Theodore Alexandrov (EMBL)

Per Andren (Uppsala University)

Josephine Bunch (The National Physical
Laboratory, UK)

Zongwei Cai (Hong Kong Baptist University)

Pierre Chaurand (University of Montreal)

Bingming Chen (Merck)

Pieter Dorrestein (University of California,
San Diego)

Richard R. Drake (Medical University of
South Carolina)

Livia Eberlin (University of Texas, Austin)

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Ingela Lanekoff (Uppsala University)

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David Muddiman (North Carolina State
University)

Olga Ovchinnikova (ORNL)

Bernhard Spengler (University of Giessen)

Jonathan Sweedler (University of Illinois
Urbana-Champaign)

Raf Van de Plas (TU Delft)

Olga Vitek (Northeastern University)

Zhibo Yang (University of Oklahoma)

PROGRAM HIGHLIGHTS AND SPECIAL FEATURES

Pre-conference workshop hosted by
Imaging Mass Spectrometry Society (IMSS)

Posters and Hot Topic short talks selected from submitted abstracts.

Go to www.asms.org to register, apply for stipend, submit
an abstract, or reserve a room at Asilomar.